Reduced 5-HT$_{2A}$ receptor signaling following selective bilateral amygdala damage

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Neurobiological evidence implicates the amygdala as well as serotonergic (serotonin, 5-HT) signaling via postsynaptic 5-HT$_{2A}$ receptors as essential substrates of anxiety behaviors. Assuming a functional interdependence of these substrates, we hypothesized that a low-fear behavioral phenotype due to bilateral lesion of the amygdala would be associated with significant 5-HT$_{2A}$ receptor changes. Thus, we used [$^{18}$F]altanserin positron emission tomography (PET) referenced to radioligand plasma levels and corrected for partial volume effects to quantify the spatial distribution of 5-HT$_{2A}$ receptor binding potential (BP$_{P}$) in a rare patient with Urbach-Wiethe disease and selective bilateral amygdala calcification damage relative to 10 healthy control subjects. Consistent with our a priori hypothesis, we observed a 70% global decrease in 5-HT$_{2A}$ receptor BP$_{P}$ in the Urbach-Wiethe patient relative to controls. Thus, brain abnormalities in this patient are not restricted to the amygdala, but extend to overall 5-HT neurotransmission via 5-HT$_{2A}$ receptors. Our findings provide important insights into the molecular architecture of human anxiety behaviors and suggest the 5-HT$_{2A}$ receptor as a promising pharmacological target to control pathological anxiety.

Keywords: Amygdala; fear; anxiety; serotonin; 5-HT$_{2A}$ receptor; PET; Urbach-Wiethe disease

INTRODUCTION

Over the past decade, interest in the human amygdaloid complex (henceforth referred to as the amygdala) has increased tremendously, driven by the progress in animal research and functional imaging techniques. Although the amygdala has been implicated in myriad affective and motivational functions including reward–reinforcement behavior, fear is still the emotion best characterized in terms of its dependence on a neural circuitry centered around the amygdala and its adaptive impact on behavior (LeDoux, 2007). However, when fear is disproportional in intensity, chronic or unrelated to genuine threats, it may become maladaptive and precipitate pathological anxiety (Millan, 2003). Anxiety spectrum disorders represent highly prevalent mental illnesses, tend to run a chronic course and are as disabling as physical diseases (Kessler et al., 1994). Consequently, there is a continuous search for novel drug targets enabling potent pharmacological control of pathological anxiety.

Serotonin [5-hydroxytryptamine (5-HT)] neurotransmission has long been implicated in the modulation of anxiety behaviors, and indeed, many anxiolytic drugs currently available interfere with 5-HT reuptake or target 5-HT receptors (Millan, 2003). Accumulating evidence points to a specific role of the postsynaptic 5-HT$_{2A}$ receptor (5-HT$_{2A}$R) in the expression of anxiety behaviors: polymorphisms of the 5-HT$_{2A}$R-coding gene (HTR2A) in humans form a haplotype that increases susceptibility for panic disorder (Unschuld et al., 2007; Yoon et al., 2008), whereas HTR2A knock-out produces a low-anxiety behavioral phenotype in mice (Weissaub et al., 2006). Moreover, personality risk factors for anxiety disorders positively correlate with frontolimbic 5-HT$_{2A}$R expression (Hurlemann et al., 2008), whereas a 70% global decrease in 5-HT$_{2A}$ receptor BP$_{P}$ in the Urbach-Wiethe patient relative to controls. Thus, brain abnormalities in this patient are not restricted to the amygdala, but extend to overall 5-HT neurotransmission via 5-HT$_{2A}$ receptors. Our findings provide important insights into the molecular architecture of human anxiety behaviors and suggest the 5-HT$_{2A}$ receptor as a promising pharmacological target to control pathological anxiety.
disorder typified by cutaneous, mucosal and visceral deposits of periodic acid-Schiff (PAS)-positive hyaline (glycoprotein) material that pathognomonically presents itself in early infancy through hoarse cries due to laryngeal infiltration and follows a slowly progressive, yet often benign course. About 250 cases have been reported worldwide (Hofer, 1973). LP has been mapped to a locus on chromosome 1q21, and pathogenic loss-of-function mutations have been identified within the extracellular matrix protein 1 gene (ECM1) (Hamada et al., 2002; Chan et al., 2007).

Selective bilateral calcification damage to the amygdala occurs in 50–75% of LP cases and has been shown to cause impaired fear recognition from facial expressions, reduced fearfulness in social contexts and a failure to acquire conditioned fear responses (Adolphs et al., 1994, 1998, 2005). These findings beg the question whether or not the documented amygdala pathology in patient A.M. is associated with localized 5-HT$_{2A}$R changes consistent with the symptomatology, or alternatively, whether the symptomatology is explainable by network disruption centred on the amygdala. We therefore used $[^{18}F]$altanserin PET referenced to radioligand plasma levels and corrected for partial volume effects to quantify the spatial distribution of 5-HT$_{2A}$R $B_P$ in a rare patient with Urbach-Wiethe disease and selective bilateral amygdala calcification damage relative to 10 healthy control subjects.

**METHODS**

**Participants**

Patient A.M. and 10 healthy control subjects matched for age (age range 26.3–34.4 years; mean age 32.0 ± 3.7 years), gender and education underwent thorough psychiatric exploration by two experienced clinicians blinded for LP diagnosis to exclude either current or past Diagnostic and Statistical Manual of Mental Disorders IV (DSM-IV) axis I and axis II disorders as potential confounding factors. Controls had no history of neurological or severe physical disorders and were, as A.M., naive to neuro- and psychopharmacological treatment. Written informed consent according to the latest revision of the 1964 Declaration of Helsinki was obtained from all participants, and study protocols were approved by the ethics committees of the Medical Faculties of the Universities of Bonn and Duesseldorf, the German Federal Office for Radiation Protection (BfS) and the German Federal Institute for Drugs and Medical Devices (BfArM).

The neuroradiological assessment demonstrated that brain calcification damage in patient A.M. was amygdala-selective, with particular emphasis on the BLA and no involvement of other nearby tissues (Hurlemann et al., 2007). Consistent with observations in other LP patients (Tranel et al., 2006), A.M. denied any subjective familiarity with fear, including the feelings of anxiety, worry, fright or panic. She claimed to never have experienced such feelings and even could not imagine situations where these feelings might afflict or disturb her. She did not appear to have a normal sense of danger and would skydive from a plane without hesitation, if given the opportunity.

Interestingly, A.M. showed much less cognitive deviation from controls than initial studies of patients with this rare etiology had suggested (Siebert et al., 2003). Neuropsychological testing showed her to be of normal IQ (Wechsler Adult Intelligence Scale-III full-scale IQ 100) (Wechsler, 1981) and within normal limits for a wide range of brain functions, including linguistic, visuo-perceptual, visuospatial and visuoconstructual faculties, and executive control such as planning and decision-making (Strange et al., 2003; Hurlemann et al., 2007). Declarative episodic memory was intact, however, an emotion-induced modulation of declarative episodic memory was absent (Strange et al., 2003; Hurlemann et al., 2007). The Positive and Negative Affect Schedule (Watson et al., 1988), the Hamilton Depression Rating Scale (Hamilton, 1960), the Hamilton Anxiety Rating Scale (Hamilton, 1959) and the State-Trait Anxiety Inventory for Adults (Spielberger, 1983) have been administered to A.M., all yielding no evidence of abnormal affective functioning. In brief, she had neither notable neuropsychological impairments nor notable psychopathology on conventional testing when undergoing PET scanning.

**PET scanning**

The $[^{18}F]$altanserin PET procedure has been detailed elsewhere (Hurlemann et al., 2008).

In brief, T1-weighted cranial magnetic resonance (MR) images Magnetization Prepared Rapid Gradient Echo (MP-RAGE) were obtained on a Siemens Trio 3T scanner.
Radiosynthesis of $[^{18}\text{F}]$altanserin was performed with a radiochemical purity of $>99\%$. Controls were infused 229 ± 7 MBq with a mean specific radioactivity of $\geq 125 \pm 83 \text{ GBq/\mu mol}$; patient A.M. was infused 241 MBq with a mean specific radioactivity of 374 GBq/\mu mol. $[^{18}\text{F}]$Altanserin was infused as a 2 min bolus followed by continuous infusion with a bolus/infusion ratio of $\text{Kbol} = 2.1 \text{ h}$. PET measurements were performed in 3D mode on a Siemens ECAT EXACT HR+ scanner (Siemens-CTI, Knoxville, TN, USA). A 10-min transmission mode on a Siemens ECAT EXACT HR+ scanner was the radioactivity concentration attributable to gray and from white matter; $f_{\text{WM}}$ and $f_{\text{GM}}$ are the fractions of white and gray matter within the voxel, respectively. Partial volume correction was performed on a voxel-wise basis. Both $f_{\text{WM}}$ and $f_{\text{GM}}$ were calculated from gray and white matter masks created by SPM5 segmentation. Missegmented voxels were manually removed using PMOD v.2.85. Masks were cut off at 0.5 and smoothed with a $5 \times 5 \times 5 \text{ mm}^3$ Gaussian filter kernel implemented into the PVE2 routine. Regression start for interpolation of the ideal white matter value at 1.00 was set to 0.95. Subsequently, the partial volume-corrected images were parametrized according to equation (1). Parametric partial volume-corrected PET datasets of controls displaying 5-HT$_{2A}$ receptor BP$_P$ referenced to plasma were warped onto the Montreal Neurological Institute (MNI)/International Consortium for Brain Mapping (ICBM) 152 T1 template as supplied with SPM5, with individual 2 mm$^3$ voxel size magnetic resonance imagings (MRIs) as source images and averaged using SPM5. For purposes of visualization, one of the individual MRI datasets was warped onto the same template. Both PET and MR images of patient A.M. are shown in native space (Figure 2B). The 3D surface rendering was performed with an algorithm developed inhouse.

RESULTS

In the present study, we used $[^{18}\text{F}]$altanserin PET referenced to radioligand plasma levels and corrected for partial volume effects to quantify the spatial distribution of 5-HT$_{2A}$R BP$_P$ in a rare patient (A.M.) with Urbach–Wiethe disease and selective bilateral amygdala calcification damage relative to 10 healthy control subjects. We found a large global reduction to a presently unreported extent overall reduction, 70% ± 4\% (Z = 4.1); Table 1; Figure 2B and C) of 5-HT$_{2A}$R BP$_P$ in cortical and subcortical regions of A.M. compared with controls. This demonstrates that brain abnormalities in A.M. are not restricted to the amygdala damage per se, but extend to a significantly decreased 5-HT$_{2A}$R expression throughout the brain.

DISCUSSION

The capacity to experience fear is presumed to be critical to promoting survival across species. As a consequence of LP and selective bilateral calcification damage to the BLA, patient A.M. lacks this capacity, a finding which is in keeping with the neuropsychological profile reported for other LP patients (Adolphs et al., 1994, 1998, 2005; Tranel et al., 2006). Consistent with our a priori hypothesis (Figure 1), amygdala pathology in A.M. is associated with a decrease in 5-HT$_{2A}$R-dependent signaling. Importantly, this decrease is global and not restricted to the amygdala. In humans, bilateral amygdala damage is associated with impaired fear recognition from facial expressions, reduced fearfulness in social contexts and a failure to acquire conditioned fear responses (Adolphs et al., 1994, 1998, 2005). In contrast, one of the most notable findings from functional
magnetic resonance imaging (fMRI) studies of anxiety disordered patients is an abnormal hyperresponsiveness of the amygdala to fear signals (Etkin and Wager, 2007). This suggests that the behavioral expression of anxiety substantially varies as a function of amygdala reactivity, ranging from absent amygdala responses and nonexpression of anxiety in amygdala-lesioned patients to exaggerated amygdala responses and overexpression of anxiety in patients with anxiety disorders.

However, the amygdala is not an island; to perceive, assess, control and adequately respond to fear signals, various brain regions must interact on electrophysiological and neurochemical levels. Quantitative analyses of amygdala connectivity have demonstrated that the amygdala is one of the most highly interconnected regions of the brain (Young et al., 1994; Pessoa, 2008). Given this empirical background, network dysfunctions, especially abnormal amygdala-prefrontal coupling, have been hypothesized as underlying anxiety disorders. These hypotheses state that due to a lack of top-down control, there is deficient inhibitory tone in the amygdala, leading to exaggerated amygdala responses to fear signals and overexpression of conditioned fear in anxiety disordered patients (Quirk and Gehlert, 2003). In addition, the amygdala receives via reciprocal projections, bottom-up modulatory input from the brainstem including the raphe nuclei, which provide the principle source of 5-HT innervation of the brain (Hornung, 2003; Hensler, 2006; Michelsen et al., 2007). Therefore, we propose that early bilateral amygdala calcification damage in patient A.M. might have resulted in a reduced formation of projections from and to the raphe nuclei, perhaps paving the way for a global underexpression of 5-HT3AR. Notwithstanding

Table 1 Synopsis of global and regional 5-HT2AR BP values as determined in LP patient A.M. and 10 healthy control subjects

<table>
<thead>
<tr>
<th>Region</th>
<th>Controls</th>
<th>Patient A.M.</th>
<th>Z score</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sub- and archicortex (global)</td>
<td>0.47 (0.1)</td>
<td>0.15</td>
<td>-2.8</td>
</tr>
<tr>
<td>Left caudate nucleus</td>
<td>0.39 (0.2)</td>
<td>0.18</td>
<td>-1.2</td>
</tr>
<tr>
<td>Right caudate nucleus</td>
<td>0.41 (0.2)</td>
<td>0.11</td>
<td>-1.5</td>
</tr>
<tr>
<td>Left amygdala</td>
<td>0.43 (0.2)</td>
<td>0.15</td>
<td>-1.5</td>
</tr>
<tr>
<td>Right amygdala</td>
<td>0.43 (0.1)</td>
<td>0.19</td>
<td>-1.6</td>
</tr>
<tr>
<td>Left putamen</td>
<td>0.69 (0.2)</td>
<td>0.12</td>
<td>-2.3</td>
</tr>
<tr>
<td>Right putamen</td>
<td>0.49 (0.3)</td>
<td>0.07</td>
<td>-1.6</td>
</tr>
<tr>
<td>Left hippocampus</td>
<td>0.57 (0.2)</td>
<td>0.16</td>
<td>-2.0</td>
</tr>
<tr>
<td>Right hippocampus</td>
<td>0.64 (0.3)</td>
<td>0.24</td>
<td>-1.4</td>
</tr>
<tr>
<td>Neocortex (global)</td>
<td>2.65 (0.5)</td>
<td>0.78</td>
<td>-4.1</td>
</tr>
<tr>
<td>Left insular cortex</td>
<td>2.02 (0.4)</td>
<td>0.55</td>
<td>-3.7</td>
</tr>
<tr>
<td>Right insular cortex</td>
<td>2.03 (0.4)</td>
<td>0.57</td>
<td>-3.8</td>
</tr>
<tr>
<td>Left anterior cingulate cortex</td>
<td>2.16 (0.5)</td>
<td>0.60</td>
<td>-3.6</td>
</tr>
<tr>
<td>Right anterior cingulate cortex</td>
<td>2.16 (0.4)</td>
<td>0.55</td>
<td>-3.2</td>
</tr>
<tr>
<td>Left orbital prefrontal cortex</td>
<td>2.54 (0.4)</td>
<td>0.55</td>
<td>-3.4</td>
</tr>
<tr>
<td>Right orbital prefrontal cortex</td>
<td>2.46 (0.4)</td>
<td>0.56</td>
<td>-3.0</td>
</tr>
<tr>
<td>Left tempopolar cortex</td>
<td>2.49 (0.4)</td>
<td>0.67</td>
<td>-3.2</td>
</tr>
<tr>
<td>Right tempopolar cortex</td>
<td>2.51 (0.4)</td>
<td>0.59</td>
<td>-3.3</td>
</tr>
<tr>
<td>Left dorsolateral prefrontal cortex</td>
<td>2.67 (0.5)</td>
<td>0.84</td>
<td>-3.7</td>
</tr>
<tr>
<td>Right dorsolateral prefrontal cortex</td>
<td>2.64 (0.5)</td>
<td>0.83</td>
<td>-3.4</td>
</tr>
<tr>
<td>Left lateral temporal cortex</td>
<td>2.57 (0.5)</td>
<td>0.79</td>
<td>-3.9</td>
</tr>
<tr>
<td>Right lateral temporal cortex</td>
<td>2.61 (0.5)</td>
<td>0.83</td>
<td>-3.8</td>
</tr>
<tr>
<td>Left occipital cortex</td>
<td>2.72 (0.4)</td>
<td>0.89</td>
<td>-4.2</td>
</tr>
<tr>
<td>Right occipital cortex</td>
<td>2.69 (0.4)</td>
<td>0.86</td>
<td>-4.8</td>
</tr>
<tr>
<td>Left lateral parietal cortex</td>
<td>2.82 (0.5)</td>
<td>0.86</td>
<td>-3.9</td>
</tr>
<tr>
<td>Right lateral parietal cortex</td>
<td>2.82 (0.5)</td>
<td>0.80</td>
<td>-4.4</td>
</tr>
</tbody>
</table>

Data are given as means (±SD). Abbreviations: LP, lipoid proteinosis of Urbach-Wiethe; SD, standard deviation; Z, Z score, i.e. the quotient of the difference and SD.
the exact etiology of this aberrance, patient A.M. had neither notable neuropsychological impairments nor notable psychopathology on conventional testing when undergoing PET scanning. This suggests that the early onset of the disease during childhood and the slowly progressive amygdala degeneration with disease duration has triggered plastic adaptations compensating potential deficits (Hurleman et al., 2007). This hypothesis definitely merits further investigation in future studies.

The present study extends the network perspective of described neurobiological models of anxiety behaviors to the neurochemical level. While Weisstaub et al. (2006) applied a genetic knock-out strategy to demonstrate that 5-HT2A nonexpression eliminates anxiety-like responses in mice, we used a lesion strategy to show that a homologous behavioral phenotype in humans is associated with reduced 5-HT2AR availability. Together, these studies converge on suggesting that the expression of anxiety behaviors requires integrity of a 5-HT2AR- and amygdala-dependent mechanism.

In contrast to patient A.M., anxiety disordered patients suffer from pathological fear that no longer serves adaptive functions. Traditionally, the conceptualization of the neurobiology underlying anxiety disorders has focussed on 5-HT and y-aminobutyric acid (GABA) neurotransmission (Nutt, 2005). Indeed the hitherto most successful therapeutic approaches are based on pharmacological interventions at the 5-HT transporter (5-HTT) and the GABA A receptor (GABA A R) complex, the molecular target of the benzodiazepine class of anxiolytic drugs (Millan, 2003). Even in healthy people, multimodal imaging has shown that 5-HTT availability predicts amygdala reactivity (Rhodes et al., 2007).

Imaging genetics provides consistent evidence for a association between amygdala reactivity and the 5-HTT linked polymorphic region (5-HTTLPR), with relatively heightened amygdala responses to fear signals in S allele carriers (Munafo et al., 2008). Evidence from pharmacological imaging indicates that amygdala responses to fear signals decrease after prolonged administration of the 5-HTT antagonists citalopram (Harnett et al., 2006) and escitalopram (Arce et al., 2008) as well as after single-dose administration of the potent GABA A R antagonist lorazepam (Paulus et al., 2005).

The proposed conceptualization of anxiety disorders as resulting from both neuroreceptor and network dysfunction might lead to the development of more focused treatment strategies that modulate 5-HT signaling via 5-HT2A R. There is accumulating support for this therapeutic rationale in observations of clinical populations: the 5-HT2A/2CR antagonists ritanserin and mianserin are anxiolytic in patients and effectively block the anxiogenic effects of m-chlorophenylpiperazine (Culemans et al., 1985; Pigott et al., 1991). The antidepressant nefazodone possesses anxiolytic activity at 5-HT2A R (along with 5-HT and norepinephrine reuptake inhibition properties) and is more effective than imipramine in the therapy of anxiety disorders (Bystritsky et al., 1999). Mirtazapine is an antidepressant with anxiolytic activity; among its many effects is the ability to block 5-HT2A R (Ribeiro et al., 2001). Selective 5-HT2A R antagonists and drugs that reduce 5-HT2A R activity may have anxiolytic potential. In line with this reasoning, atypical antipsychotics with prominent 5-HT2A R blockade are being studied for their efficacy in anxiety disordered patients (Pillay and Stein, 2007; but see Griebel et al., 1997).

CONFLICT OF INTEREST
None declared.

REFERENCES


