Primates in Peril

The World's 25 Most Endangered Primates 2012-2014



Edited by Christoph Schwitzer, Russell A. Mittermeier, Anthony B. Rylands, Lucy A. Taylor, Federica Chiozza, Elizabeth A. Williamson, Janette Wallis and Fay E. Clark

2014















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Illustrations by Stephen D. Nash

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The 2012–2014 iteration of the World's 25 Most Endangered Primates was drawn up during an open meeting held during the XXIV Congress of the International Primatological Society (IPS), Cancún, 14 August 2012, and was published as a series of Species Fact Sheets (Mittermeier *et al.* 2012).

Here, we present an extended version of the 2012–2014 list, with more comprehensive information about the threats facing these primates and with bibliographic references cited in the text. We have updated the species profiles from the 2008–2010 edition for those species remaining on the list, and added additional profiles for newly listed species.

This publication is a joint effort of the IUCN SSC Primate Specialist Group, the International Primatological Society, Conservation International, and the Bristol Zoological Society.

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The World's 25 Most Endangered Primates: 2012-2014

Here we report on the seventh iteration of the biennial listing of a consensus of the 25 primate species considered to be among the most endangered worldwide and the most in need of conservation measures.

The 2012–2014 list of the world's 25 most endangered primates has five species from Africa, six from Madagascar, nine from Asia, and five from the Neotropics (Table 1). Madagascar tops the list with six species. Vietnam has five, Indonesia three, Brazil two, and China, Colombia, Côte d'Ivoire, the Democratic Republic of Congo, Ecuador, Equatorial Guinea, Ghana, Kenya, Peru, Sri Lanka, Tanzania, and Venezuela each have one.

The changes made in this list compared to the previous iteration (2010–2012) were not because the situation of the nine species that were dropped (Table 2) has improved. In some cases, for example, *Varecia variegata*, the situation has in fact worsened. By making these changes we intend rather to highlight other, closely-related species enduring equally bleak prospects for their survival. An exception may be the greater bamboo lemur, *Prolemur simus*, for which recent studies have confirmed a considerably larger distribution range and larger estimated population size than previously assumed. However, severe threats to this species in eastern Madagascar remain.

Nine of the primates were not on the previous (2010–2012) list (Table 3). Seven of them are listed as among the world's most endangered primates for the first time. The Tana River red colobus and the Ecuadorian brownheaded spider monkey had already been included in previous iterations, but were subsequently removed in favour of other highly threatened species of the same genera. The 2012–2014 list now contains two members each of these genera, thus particularly highlighting the severe threats they are facing.

During the discussion of the 2012–2014 list at the XXIV Congress of IPS in Cancún in 2012, a number of other highly threatened primate species were considered for inclusion (Table 4). For all of these, the situation in the wild is as precarious as it is for those that eventually made it on the list.

 $\textbf{Table 1}. \ \textbf{The World's 25 Most Endangered Primates 2012-2014}$

Africa		
Galagoides rondoensis	Rondo dwarf galago	Tanzania
Cercopithecus roloway	Roloway monkey	Côte d'Ivoire, Ghana
Piliocolobus pennantii pennantii	Bioko red colobus	Equatorial Guinea (Bioko Is.)
Piliocolobus rufomitratus	Tana River red colobus	Kenya
Gorilla beringei graueri	Grauer's gorilla	DRC
Madagascar		
Microcebus berthae	Madame Berthe's mouse lemur	Madagascar
Eulemur flavifrons	Sclater's black lemur	Madagascar
Varecia rubra	Red ruffed lemur	Madagascar
Lepilemur septentrionalis	Northern sportive lemur	Madagascar
Propithecus candidus	Silky sifaka	Madagascar
Indri indri	Indri	Madagascar
Asia		
Tarsius pumilus	Pygmy tarsier	Indonesia (Sulawesi)
Nycticebus javanicus	Javan slow loris	Indonesia (Java)
Simias concolor*	Pig-tailed snub-nosed langur	Indonesia (Mentawai Is.)
Trachypithecus delacouri	Delacour's langur	Vietnam
Trachypithecus poliocephalus	Golden-headed or Cat Ba langur	Vietnam
Semnopithecus vetulus nestor	Western purple-faced langur	Sri Lanka
Pygathrix cinerea	Grey-shanked douc monkey	Vietnam
Rhinopithecus avunculus	Tonkin snub-nosed monkey	Vietnam
Nomascus nasutus	Cao-Vit or Eastern black-crested gibbon	China, Vietnam
Neotropics		
Ateles hybridus	Variegated spider monkey	Colombia, Venezuela
Ateles fusciceps fusciceps	Ecuadorian brown-headed spider monkey	Ecuador
Cebus kaapori	Ka'apor capuchin	Brazil
Callicebus oenanthe	San Martín titi monkey	Peru
Alouatta guariba guariba	Northern brown howler	Brazil

^{*} The pig-tailed snub-nosed langur Simias concolor had previously been classified as Nasalis concolor and referred to as such in the 2012–2014 Top 25 Fact sheets.

Table 2. Primate species included on the 2010–2012 list that were removed from the 2012–2014 list.

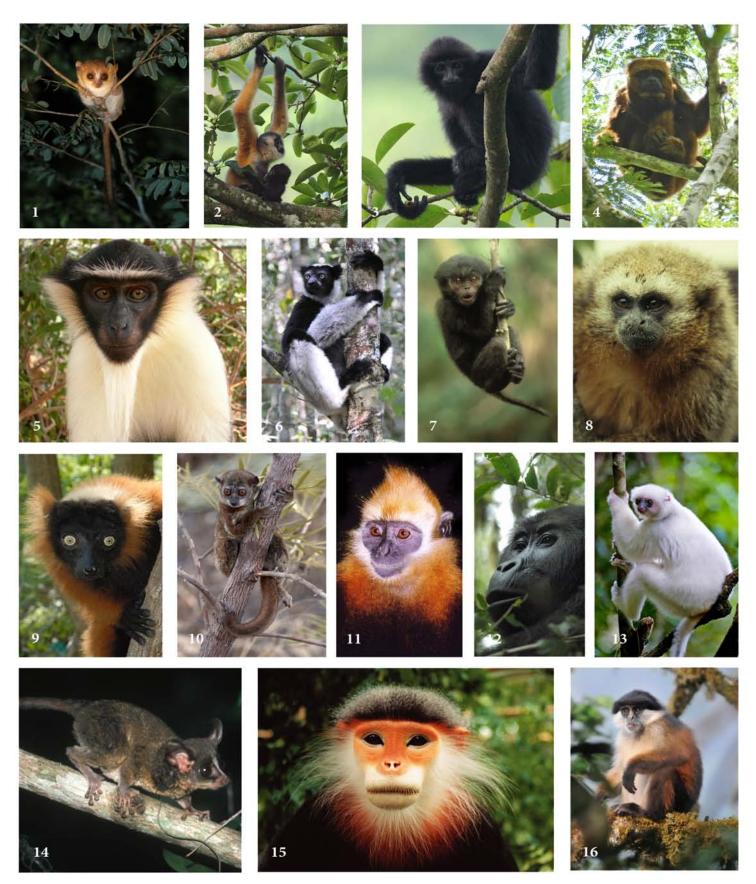
Africa		
Piliocolobus epieni	Niger Delta red colobus	Nigeria
Madagascar		
Prolemur simus	Greater bamboo lemur	Madagascar
Varecia variegata	Black-and-white ruffed lemur	Madagascar
Asia		
Tarsius tumpara	Siau Island tarsier	Indonesia (Siau Is.)
Macaca silenus	Lion-tailed macaque	India
Pongo pygmaeus pygmaeus	Northwest Bornean orangutan	Indonesia (West Kalimantan, Borneo), Malaysia (Sarawak)
Neotropics		
Cebus flavius	Blond capuchin	Brazil
Callicebus barbarabrownae	Barbara Brown's titi monkey	Brazil
Oreonax flavicauda	Peruvian yellow-tailed woolly monkey	Peru

Table 3. Primate species that were added to the 2012–2014 list. The Tana River red colobus and the Ecuadorian brown-headed spider monkey were added to the list after previously being removed. The other seven species are new to the list.

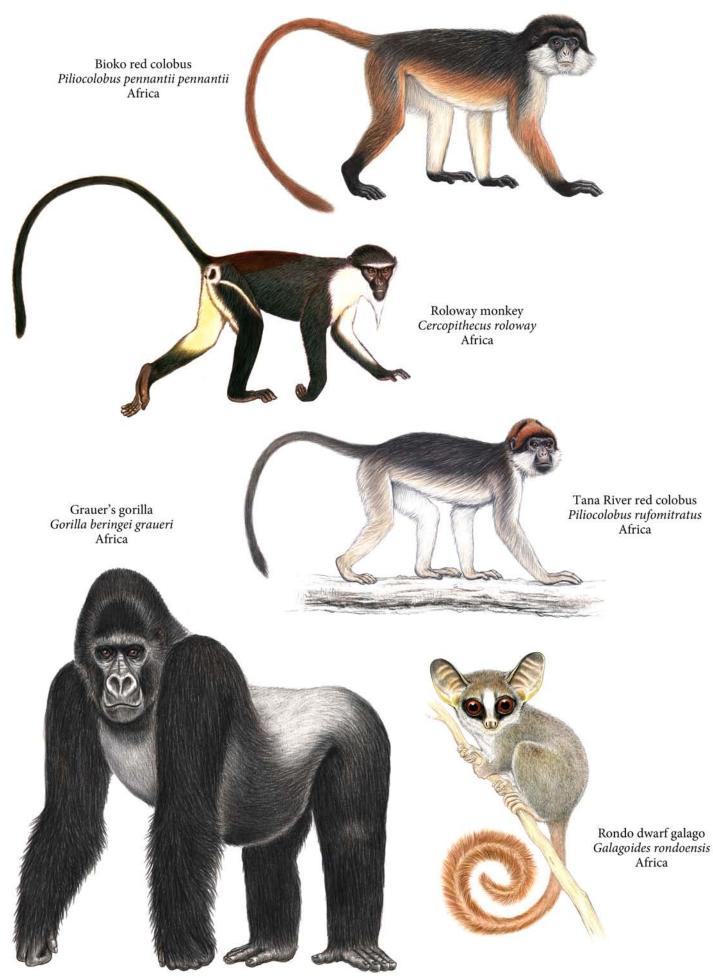
Africa		
Piliocolobus rufomitratus	Tana River red colobus	Kenya
Madagascar		
Microcebus berthae	Madame Berthe's mouse lemur	Madagascar
Varecia rubra	Red ruffed lemur	Madagascar
Indri indri	Indri	Madagascar
Asia		
Tarsius pumilus	Pygmy tarsier	Indonesia (Sulawesi)
Neotropics		
Ateles fusciceps fusciceps	Ecuadorian brown-headed spider monkey	Ecuador
Cebus kaapori	Ka'apor capuchin	Brazil
Callicebus oenanthe	San Martín titi monkey	Peru
Alouatta guariba guariba	Northern brown howler	Brazil

Table 4. Primate species considered during the discussion of the 2012–2014 list at the IPS Congress in Cancún that did not make it onto the list, but are also highly threatened.

Africa		
Piliocolobus preussi	Preuss's red colobus	Cameroon, Nigeria
Gorilla gorilla diehli	Cross River gorilla	Nigeria, Cameroon
Pan troglodytes ellioti	Nigeria-Cameroon chimpanzee	Nigeria, Cameroon
Madagascar		
Cheirogaleus sibreei	Sibree's dwarf lemur	Madagascar
Hapalemur alaotrensis	Lac Alaotra bamboo lemur	Madagascar
Eulemur cinereiceps	White-collared brown lemur	Madagascar
Propithecus perrieri	Perrier's sifaka	Madagascar
Asia		
Nasalis larvatus	Proboscis monkey	Indonesia (Borneo)
Presbytis comata	Grizzled leaf monkey	Indonesia
Rhinopithecus strykeri	Myanmar snub-nosed monkey	Myanmar, China
Nomascus hainanus	Hainan black-crested gibbon	China (Hainan)
Nomascus leucogenys	Northern white-cheeked black-crested gibbon	Laos, Vietnam, China
Neotropics		
Chiropotes satanas	Black bearded saki	Brazil
Leontopithecus caissara	Black-faced lion tamarin	Brazil
Saguinus bicolor	Pied tamarin	Brazil
Callicebus caquetensis	Caquetá titi monkey	Colombia



Photos of some of the Top 25 Most Endangered Primates. From top to bottom, left to right: 1. Microcebus berthae (photo by John R. Zaonarivelo); 2. Nomascus nasutus (female) (photo by Xu Yongbin); 3. Nomascus nasutus (male) (photo by Xu Yongbin); 4. Alouatta guariba guariba (photo by Leonardo Gomes Neves); 5. Cercopithecus roloway (photo by S. Wolters, WAPCA); 6. Indri indri (photo by Russell A. Mittermeier); 7. Simias concolor (juvenile) (photo by Richard Tenaza); 8. Callicebus oenanthe (photo by Russell A. Mittermeier); 9. Varecia rubra (photo by Russell A. Mittermeier); 10. Lepilemur septentrionalis (photo by Edward E. Louis, Jr.); 11. Trachypithecus poliocephalus (photo by Tilo Nadler); 12. Gorilla beringei graueri (photo by Russell A. Mittermeier); 13. Propithecus candidus (photo by Iñaki Relanzón); 14. Galagoides rondoensis (photo by Andrew Perkin); 15. Pygathrix cinerea (photo by Tilo Nadler); 16. Piliocolobus p. pennantii (photo by R. A. Bergl).

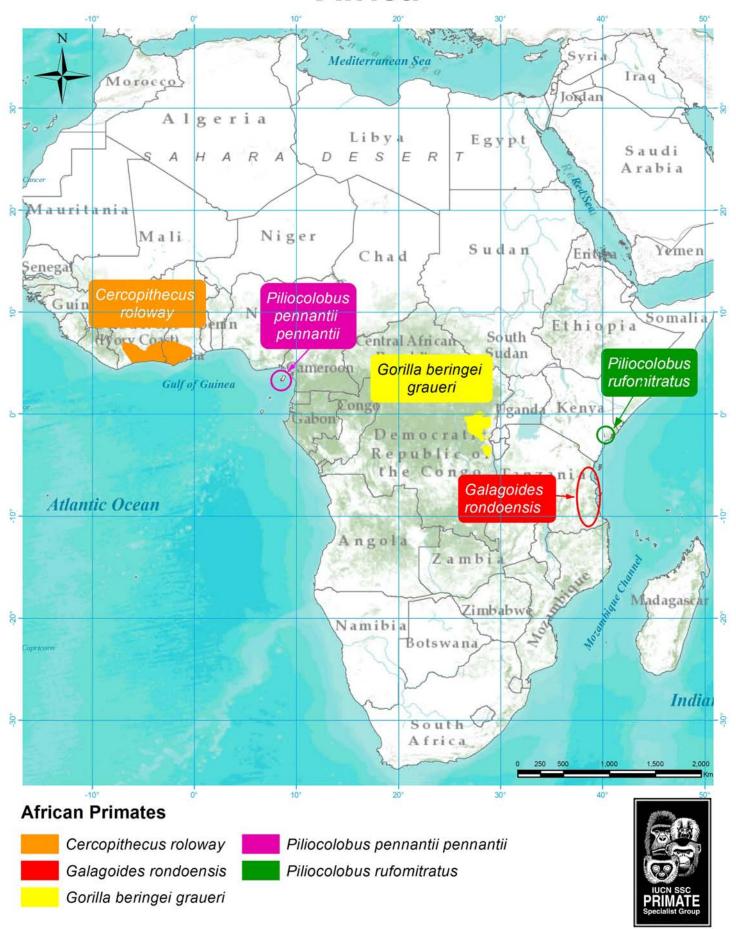








Africa



Rondo Dwarf Galago

Galagoides rondoensis Honess in Kingdon, 1997 Tanzania (2012)

Andrew Perkin

Weighing approximately 60 g, this is the smallest of all galago species (Perkin et al. 2013). It is distinct from other dwarf galagos in its diminutive size, a bottlebrush-shaped tail, its reproductive anatomy, and its distinctive "double unit rolling call" (Perkin and Honess 2013). Current knowledge indicates that this species occurs in two distinct areas, one in southwest Tanzania near the coastal towns of Lindi and Mtwara, the other approximately 400 km further north, above the Rufiji River, in pockets of forest around Dar es Salaam. One further population occurs in Sadaani National Park, approximately 100 km north of Dar es Salaam. Rondo dwarf galagos have a mixed diet of insects and fruit, often feed close to the ground, and move by vertical clinging and leaping in the shrubby understorey. They build daytime sleeping nests, which are often in the canopy (Bearder et al. 2003). As with many small primates, G. rondoensis is probably subject to predation by owls and other nocturnal predators. Among these, genets, palm civets and snakes invoke intense episodes of alarm calling (Perkin and Honess 2013).

Over the last decade, the status of G. rondoensis has changed from Endangered in 2000 to Critically Endangered in 2008 on the IUCN Red List (Perkin et al. 2008). It has an extremely limited and fragmented range in a number of remnant patches of Eastern African Coastal Dry Forest (sensu Burgess and Clarke 2000; p.18) in Tanzania, namely those at Zaraninge forest (06°08'S, 38°38'E) in Sadaani National Park (Perkin 2000), Pande Game Reserve (GR) (06°42'S, 39°05'E), Pugu/Kazimzumbwi (06°54'S, 39°05'E) (Perkin 2003, 2004), Rondo (NR) (10°08'S, 39°12'E), Litipo (10°02'S, 39°29'E) and Ziwani (10°20'S, 40°18'E) forest reserves (FR) (Honess 1996; Honess and Bearder 1996). New sub-populations were identified in 2007 near Lindi town in Chitoa FR (09°57'S, 39°27'E) and Ruawa FR (09°44'S, 39°33'E), and in 2011 in Noto Village Forest Reserve (09°53'S, 39°25'E) (Perkin et al. 2011, 2013.) and in the northern population at Ruvu South Forest Reserve (06°58'S, 38°52'E). Specimens of G. rondoensis, originally described as Galagoides demidovii phasma,



Rondo dwarf galago (*Galagoides rondoensis*) (Illustration: Stephen D. Nash)

were collected by Ionides from Rondo Plateau in 1955, and Lumsden from Nambunga, near Kitangari, (approximately 10°40'S, 39°25'E) on the Makonde Plateau in Newala District in 1953. Doubts surround the persistence of this species on the Makonde Plateau, which has been extensively cleared for agriculture. Surveys there in 1992 failed to detect any extant populations (Honess 1996).

No detailed surveys have been conducted to assess population sizes of *G. rondoensis*. Distribution surveys have been conducted, however, in the southern (Honess 1996, Perkin et al. in prep.) and northern coastal forests of Tanzania (29 surveyed) and Kenya (seven surveyed) (Perkin 2000, 2003, 2004; Perkin et al., 2013). Absolute population sizes remain undetermined but recent surveys have provided estimates of density (3-6/ha at Pande Game Reserve [Perkin 2003] and 8/ha at Pugu Forest Reserve [Perkin 2004]) and relative abundance from encounter rates (3-10/hr at Pande Game Reserve and Pugu/Kazimzumbwi Forest Reserve [Perkin 2003, 2004]) and 3.94/hr at Rondo Forest Reserve (Honess 1996). There is a clear and urgent need for further surveys to determine population sizes in these dwindling forest patches.

In 2008, it was reported that the total area of forest in which G. rondoensis is currently known to occur does not exceed 101.6 km² (Pande GR: 2.4 km², Rondo FR: 25 km², Ziwani FR: 7.7 km², Pugu/Kazimzumbwi FR: 33.5 km², Litipo FR: 4 km², Zaraninge forest: 20 km², Chitoa FR: 5 km², and Ruawa FR 4 km²) (Minimum area data source: Burgess and Clarke 2000; Doggart 2003; Perkin et al. in prep.). New data on forest area change indicates that while two new sub-populations have been discovered; the overall area of occupancy hovers around 100 km². 2008 and 2014 forest-area estimations are as follows: Zaraninge 2008: 20 km², 2014: 15 km²; Pande 2008: 2.4 km², 2014: 2.4 km²; Pugu/Kazimzumbwe 2008: 33.5 km², 2014: 9 km²; Ruvu South 2008: 20 km², 2014: 10 km²; Ruawa 2008: 4 km², 2014: 4 km²; Litipo 2008: 4 km², 2014: 3 km²; Chitoa 2008: 4 km², 2014: 5 km²; Noto 2008: 21 km², 2014: 20 km²; Rondo 2008: 25 km², 2014: 25 km²; Ziwani 2008: 7.7 km², 2014: 1 km². The total forest area estimates are as follows - 2008: 101.6 km², 2014: 94.4 km².

The major threat facing this species is loss of habitat. All sites are subject to some level of agricultural encroachment, charcoal manufacture and/or logging. All sites, except Pande (Game Reserve), Zaraninge (within Saadani National Park) and Rondo (Nature Reserve), are national or local authority forest reserves and as such nominally, but in practice minimally, protected. Since 2008, there have been changes resulting in the increase in protection of two forests. The Noto plateau forest, formerly open village land, is part of a newly created village forest reserve, and the Rondo Forest Reserve has now been declared a new Nature Reserve, both are important for Rondo galago

conservation given their relatively large size. Given current trends in charcoal production for nearby Dar es Salaam, the forest reserves of Pugu and Kazimzumbwi were predicted to disappear over the next 10-15 years (Ahrends 2005). Pugu/Kazimzumbwe as well as Ruvu South have seen continued and predicted losses to the rampant charcoal trade since Ahrends (2005) study. Pande, as a Game Reserve, is perhaps more secure, and Zareninge forest, being in a National Park, is the most protected part of the range of *G. rondoensis*. In the south, the Noto, Chitoa and Rondo populations are the most secure, as they are buffered by tracts of woodland. The type population at Rondo is buffered by woodland and Pinus plantations managed by the Rondo Forestry Project, and is now a Nature Reserve. Litipo, and Ruawa FRs are under threat from bordering village lands. Ziwani is now mostly degraded scrub forest, thicket and grassland.

Conservation action is urgently needed by: monitoring rates of habitat loss, surveying new areas for remnant populations, estimating population size, reassessing the phylogenetic relationships of the sub-populations and increasing awareness. There is emerging data (vocal and penile morphological) that the northern and southern populations may be phylogenetically distinct with important taxonomic implications. As such the conservation of all sub-populations is important.

Across its known range, the Rondo galago can be found in sympatry with a number of other galagos, including two much larger species in the genus *Otolemur*: Garnett's galago *O. garnettii* (Least Concern, Butynski *et al.* 2008a), and the thick-tailed galago, *O. crassicaudatus* (Least Concern, Bearder 2008). The Rondo galago is sympatric with the Zanzibar galago, *Galagoides zanzibaricus* (Least Concern, Butynski *et al.* 2008b), in the northern parts of its range (for example, in Zaraninge forest, Pugu/Kazimzumbwi FR and Pande GR). In the southern parts of its range (for example, in Rondo, Litipo and Noto), the Rondo galago is sympatric with Grant's galago, *Galagoides granti* (Least Concern, Honess *et al.* 2008).

A new project to address these conservation and research issues is being implemented this year. Targeted conservation initiatives are taking place in Ruvu South FR, Chitoa FR and Noto VFR.

References

Ahrends, A. 2005. Pugu Forest: going, going. *Arc Journal* 17: 23.

Bearder, S.K. 2008. *Otolemur crassicaudatus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. www.iucnredlist.org. Accessed 16 March 2014.

Bearder, S. K., L. Ambrose, C. Harcourt, P. E. Honess, A. Perkin, S. Pullen, E. Pimley and N. Svoboda. 2003. Species-typical patterns of infant care, sleeping site use and social cohesion among nocturnal primates in Africa. *Folia Primatologica* 74: 337–354.

Burgess, N. D. and G. P. Clarke. 2000. *Coastal Forests of Eastern Africa*. IUCN – The World Conservation Union, Gland, Switzerland, and Cambridge, UK.

Butynski, T. M., S. K. Bearder, S. and Y. de Jong. 2008a. *Otolemur garnettii*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 16 March 2014.

Butynski, T. M., Y de Jong, A. Perkin, S. K. Bearder and P. Honess. 2008b. *Galagoides zanzibaricus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. www.iucnredlist.org. Accessed 16 March 2014.

Doggart, N. (ed.). 2003. *Pande Game Reserve: A Biodiversity Survey*. Tanzania Forest Conservation Group, Technical Paper 7. Dar es Salaam.

Honess, P. E. 1996. Speciation among galagos (Primates, Galagidae) in Tanzanian forests. Doctoral thesis, Oxford Brookes University, Oxford, UK.

Honess, P. E. and S. K. Bearder. 1996. Descriptions of the dwarf galago species of Tanzania. *African Primates* 2: 75–79.

Honess, P. E., A. Perkin, S. K. Bearder, T. M. Butynski and Y. de Jong. 2008. *Galagoides granti*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed on 16 March 2014.

Kingdon, J. 1997. *The Kingdon Field Guide to African Mammals*. Academic Press, London.

Perkin, A. 2000. A Field Study of the Conservation Status and Diversity of Galagos in Zaraninge Forest, Coast Region, Tanzania. Report of WWF-Tanzania, Dar-es-Salaam.

Perkin, A. 2003. Mammals. In: *Pande Game Reserve: A Biodiversity Survey*, N. Doggart (ed.), 95pp. Tanzania Forest Conservation Group, Technical Paper 7. Dar es Salaam.

Perkin, A. 2004. *Galagos of the Coastal Forests and Eastern Arc Mtns. of Tanzania – Notes and Records.* Tanzania Forest Conservation Group, Technical Paper 8. Dar es Salaam, Tanzania.

Perkin, A., S. K. Bearder, P. Honess and T. M. Butynski. 2008. *Galagoides rondoensis*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 16 March 2014.

Perkin, A., S. K. Bearder and J. Karlsson. In prep. Galago surveys in Rondo, Litipo, Chitoa, Ruawa, Ndimba and Namatimbili forests, Lindi Region, southeastern Tanzania.

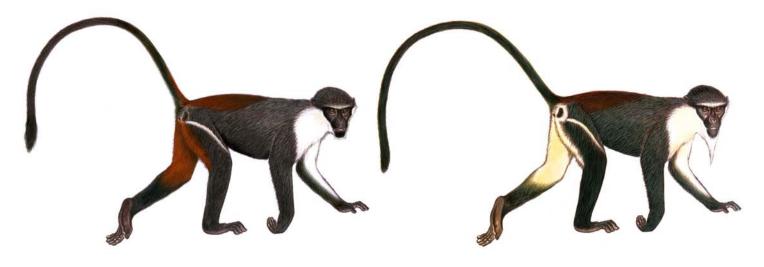
Perkin, A., B. Samwel and J. Gwegime. 2011. Going for gold in the Noto Plateau, SE Tanzania. *Arc Journal* 26: 14–16.

Perkin, A.W., P. E. Honess and T. M. Butynski. 2013. Mountain dwarf galago *Galagoides orinus*. In: *Mammals of Africa: Volume II: Primates*, T. Butynski, J. Kingdon and J. Kalin (eds.), pp. 452–454. Bloomsbury Publishing, London.

Roloway Monkey

Cercopithecus diana roloway (Schreber, 1774) Ghana, Côte d'Ivoire (2002, 2006, 2008, 2010, 2012)

W. Scott McGraw & John F. Oates



Roloway monkey (right) (Cercopithecus diana roloway) and Diana monkey (left) (Cercopithecus diana diana) (Illustrations: Stephen D. Nash)

There are two subspecies of *Cercopithecus diana*, both highly attractive, arboreal monkeys that inhabit the upper Guinean forests of West Africa (Grubb *et al.* 2003). Groves (2001) considers the two subspecies to be sufficiently distinct to be regarded as full species. Of the two forms, the Roloway (*C. d. roloway*) which is known from Ghana and central and eastern Côte d'Ivoire, is more seriously threatened with extinction; it is classified as Endangered (Oates *et al.* 2008), but its status should be upgraded to Critically Endangered.

The *roloway* subspecies is distinguished by its broad white brow line, long white beard and yellow thighs. Roloway monkeys are upper-canopy specialists that prefer undisturbed forest. Destruction and degradation of their habitat and relentless hunting for the bushmeat trade have reduced their population to small, isolated pockets. Miss Waldron's red colobus (*Procolobus badius waldroni*) once inhabited many of the same forest areas as the Roloway, but is now almost certainly extinct (Oates 2011). Unless more effective conservation action is taken, there is a strong possibility that the Roloway monkey will also disappear in the near future.

Over the last 40 years Roloway monkeys have been steadily extirpated in Ghana. Several recent surveys

have failed to confirm the presence of these monkeys in any reserves in western Ghana, including Bia National Park, Krokosua Hills Forest Reserve, Subri River Forest Reserve and Dadieso Forest Reserve (Oates 2006; Gatti 2010; Buzzard and Parker 2012; Wiafe 2013), although it is possible that the Ankasa Conservation Area still contains a few individuals (Magnuson 2003; Gatti 2010). The Kwabre forest in the far southwestern corner of the country is the only site in Ghana at which any Roloways have been reported as seen by scientists or conservationists in the last decade; surveys at this site were made by West African Primate Conservation Action in 2011 and 2012 (WAPCA 2012). Kwabre consists of fragments of swamp forest along the lower Tano River, adjacent to the Tanoé forest in Côte d'Ivoire; WAPCA has launched a community-based conservation project with villages around Kwabre, and collaboration with conservation efforts in Tanoé. Meanwhile, further efforts should be made to ascertain whether any Roloway monkeys still survive in the Ankasa, because this site has significant conservation potential and Roloways have been reported there in the relatively recent past.

In neighbouring Côte d'Ivoire, the Roloway's status is equally dire. Less than ten years ago Roloways were

known or strongly suspected to exist in three forests: the Yaya Forest Reserve, the Tanoé forest adjacent to the Ehy Lagoon, and Parc National des Iles Ehotilé (McGraw 1998, 2005; Koné and Akpatou 2005). Surveys of eighteen areas between 2004 and 2008 (Gonedelé Bi et al. 2008, 2012) confirmed the presence of Roloways only in the Tanoé forest suggesting that the Roloway monkey may have been eliminated from at least two forest areas (Parc National des Iles Ehotilé, Yaya Forest Reserve) within the last decade. Subsequent surveys carried out in southern Côte d'Ivoire suggest a handful of Roloways may still survive in two forest reserves along the country's coast. On 21 June 2012, Gonedelé bi Sery observed one Roloway individual in the Dassioko Sud Forest Reserve; however, Roloways have not been located in this forest reserve since, despite regular patrols there (Bitty et al. 2013; Gonedelé Bi et al. in review). In 2012, Gonedelé Bi and A. E. Bitty observed Roloways in Port Gauthier Forest Reserve, and in October 2013, Gonedelé Bi obtained photographs of monkeys poached inside this reserve, including an image purported to be a Roloway. The beard on this individual appears short for a Roloway, raising the possibility that surviving individuals in this portion of the interfluvial region may in fact be hybrids. The Dassioko Sud and Port Gauthier Forest Reserves are described as coastal evergreen forests and both are heavily degraded due to a large influx of farmers and hunters from the northern portion of the country (Bitty et al. 2013). Gonedelé Bi and colleagues, in cooperation with SODEFOR (Société de Développement des Forêts) and local communities, have organized regular forest patrols aimed at removing illegal farmers and hunters from both reserves. Nevertheless, the most recent surveys have failed to locate living Roloways in either reserve (Gonedelé Bi and Bitty 2013) meaning that the only forest in Côte d'Ivoire where Roloways are confirmed to exist is the Tanoé forest adjacent to the Ehy Lagoon. This wet forest also harbours one of the few remaining populations of white-naped mangabeys in Côte d'Ivoire. Efforts led by I. Koné and involving several organizations (CEPA, WAPCA) helped stop a large palm oil company from further habitat degradation and a community-based conservation effort has helped slow poaching within this forest (Koné 2008). Unfortunately, hunting still occurs in Tanoé and the primate populations within it are undoubtedly decreasing (Gonedelé Bi et al. 2013). As the potential last refuge for Roloways and White-naped mangabeys, the protection of the Tanoé forest should be the highest conservation priority. By any measure, the Roloway monkey must be considered one of the most critically endangered monkeys in Africa and appears to be on the verge of extinction (Oates 2011).

References

Bitty, E. A., S. Gonedelé Bi and W. S. McGraw. 2013. Accelerating deforestation and hunting in protected reserves jeopardize primates in southern Côte d'Ivoire. *American Journal of Physical Anthropology* Supp. 56: 81–82.

Buzzard, P. J. and A. J. A. Parker. 2012. Surveys from the Subri River Forest Reserve, Ghana. *African Primates* 7: 175–183.

Gatti, S. 2010. Status of Primate Populations in Protected Areas Targeted by the Community Forest Biodiversity Project. Unpublished report, West African Primate Conservation Action (WAPCA), Accra, Ghana.

Gonedelé Bi, S. and A. E. Bitty. 2013. Conservation of threatened primates of Dassioko Sud and Port Gauthier forest reserves in coastal Côte d'Ivoire. Final Report to Primate Conservation Inc., Charlestown, RI.

Gonedelé Bi, S., I. Koné, J.-C. K. Béné, A. E. Bitty, B. K. Akpatou, Z. Goné Bi, K. Ouattara and D. A. Koffi. 2008. Tanoé forest, south-eastern Côte d'Ivoire identified as a high priority site for the conservation of critically endangered primates in West Africa. *Tropical Conservation Science* 1: 265–278.

Gonedelé Bi, S., J.-C. K. Béné, A. E. Bitty, A. N'Guessan, A. D. Koffi, B. Akptatou and I. Koné. 2013. Roloway guenon (*Cercopithecus diana roloway*) and white-naped mangabey (*Cercocebus atys lunulatus*) prefer mangrove habitats in Tanoé Forest, southeastern Ivory Coast. *Ecosystems and Ecography* 3: 126.

Gonedelé Bi, S, I. Koné, A. E. Bitty, J.-C. K. Béné, B. Akptatou and D. Zinner. 2012. Distribution and conservation status of catarrhine primates in Côte d'Ivoire (West Africa). *Folia Primatologica* 83: 11–23.

Gonedelé Bi S., E. A. Bitty and W. S. McGraw. In review. Conservation of threatened primates in Dassioko Sud and Port Gauthier forest reserves: use of field patrols to assess primate abundance and illegal human activities.

Groves, C. P. 2001. *Primate Taxonomy*. Smithsonian Institution Press, Washington, DC.

Grubb, P., T. M. Butynski, J. F. Oates, S. K. Bearder, T. R. Disotell, C. P. Groves and T. T. Struhsaker. 2003. An assessment of the diversity of African primates. *International Journal of Primatology* 24: 1301–1357.

Koné, I. 2008. The Tanoé Swamp Forest, a poorly known high conservation value forest in jeopardy in southeastern Côte d'Ivoire. Unpublished Report.

Koné, I. and K. B. Akpatou. 2005. Recherche en Côte d'Ivoire de trois singes gravement menaces d'extinction. *CEPA Magazine* 12: 11–13.

Magnuson, L. 2003. Final Brief: Ecology and Conservation of the Roloway Monkey in Ghana. Unpublished report, Wildlife Division of Ghana, Forestry Commission, Ghana.

McGraw, W. S. 1998. Surveys of endangered primates in the forest reserves of eastern Côte d'Ivoire. *African Primates* 3: 22–25.

McGraw, W. S. 2005. Update on the search for Miss Waldron's red colobus monkey (*Procolobus badius waldroni*). *International Journal of Primatology* 26: 605–619.

Oates, J. F. 2006. Primate Conservation in the Forests of Western Ghana: Field Survey Results, 2005–2006. Report to the Wildlife Division, Forestry Commission, Ghana.

Oates, J. F. 2011. *Primates of West Africa: A Field Guide and Natural History*. Conservation International, Arlington, VA.

Oates, J. F., S. Gippoliti and C. P. Groves. 2008. *Cercopithecus diana* ssp. *roloway*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 16 March 2014.

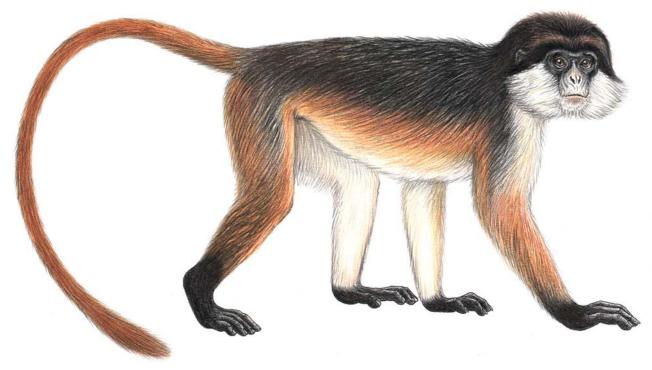
WAPCA. 2012. Annual Report. West African Primate Conservation Action, Accra, Ghana.

Wiafe, E. 2013. Status of the Critically Endangered Roloway monkey (*Cercopithecus diana roloway*) in the Dadieso Forest Reserve, Ghana. *African Primates* 8: 9–15.

Bioko Red Colobus

Piliocolobus pennantii pennantii (Waterhouse, 1838) Bioko Island, Equatorial Guinea (2004, 2006, 2010, 2012)

Drew T. Cronin, Gail W. Hearn & John F. Oates



Bioko red colobus (*Piliocolobus pennantii pennantii*) (Illustration: Stephen D. Nash)

Pennant's red colobus monkey *Piliocolobus pennantii* is presently regarded by the IUCN Red List as comprising three subspecies: *P. pennantii pennantii* of Bioko, *P. p. epieni* of the Niger Delta, and *P. p. bouvieri* of the Congo Republic. Some accounts give full species status to all three of these monkeys (Groves 2007; Oates 2011; Groves and Ting 2013). *P. p. pennantii* is currently classified as Endangered (Oates and Struhsaker 2008).

Piliocolobus pennantii pennantii may once have occurred over most of Bioko, but it is now probably limited to an area of less than 300 km² within the Gran Caldera and a 510 km² range in the Southern Highlands Scientific Reserve (GCSH) (Cronin et al. 2013). Low numbers of P. p. pennantii may have persisted through the 1980s in Pico Basile National Park (330 km²) (Gonzalez Kirchner 1994), but there have been no confirmed historical or current sightings in the area. Another isolated population was believed to exist in the southeastern extent of the GCSH; however, recent surveys did not uncover any evidence of this monkey and it is probably extirpated in that area (Cronin 2013).

P. p. pennantii is threatened by bushmeat hunting, most notably since the early 1980s when a commercial bushmeat market appeared in the town of Malabo (Butynski and Koster 1994). Following the discovery of offshore oil in 1996, and the subsequent expansion of Equatorial Guinea's economy, rising urban demand led to increased numbers of primate carcasses in the bushmeat market (Morra et al. 2009; Cronin 2013). In November 2007, a primate hunting ban was enacted on Bioko, but it lacked any realistic enforcement and contributed to a spike in the numbers of monkeys in the market. Between October 1997 and September 2010, a total of 1,754 P. p. pennantii were observed for sale in the market (Cronin 2013). The rate of occurrence of P. p. pennantii carcasses in the market though, has been consistently less than more common primates on Bioko, suggesting that its restricted range is passively protecting the remaining population from significant hunting.

The average price paid in the Malabo market for an adult *P. p. pennantii* in 2008 was about US\$50 (D. T.

Cronin, unpubl. data). This is well over twice the cost of the readily available, high quality whole chicken and beef at the same market. Similar high prices are paid on Bioko for all seven species of monkeys and for both species of duikers. Mainland carcasses are now also regularly shipped to Malabo for sale suggesting that transport costs are covered by the high profits relative to those in Nigeria, Cameroon, or Rio Muni (Morra et al. 2009). Bushmeat on Bioko is, obviously, now a 'luxury food' (Hearn et al. 2006). The continued high flow of primates, duikers and other wildlife into the Malabo bushmeat market indicates that neither of the protected areas is receiving adequate management and that existing hunting laws lack enforcement from the government of Equatorial Guinea.

Of the other two subspecies of *P. pennantii*, Bouvier's red colobus *P. p. bouvieri* of east-central Republic of Congo has not been observed alive by scientists for at least 25 years, raising concerns that it may be extinct (Oates 1996; Struhsaker 2005). The habitat of the Niger Delta red colobus *P. p. epieni* in southern Nigeria has been severely degraded by logging, the surviving monkeys face ever-increasing hunting pressure, and there is no protected area within its range (Oates 2011).

Red colobus monkeys are probably more threatened than any other taxonomic group of primates in Africa (Oates 1996; Struhsaker 2005, 2011), and the status of the western African forms is especially precarious. Preuss's red colobus P. preussi of western Cameroon and southeastern Nigeria is Critically Endangered (Oates et al. 2008) as a result of relentless hunting, and Miss Waldron's red colobus *P. badius waldroni* of eastern Côte d'Ivoire and western Ghana is now almost certainly extinct (Oates 2011). All remaining West African red colobus populations and their habitats therefore require rigorous protection. Such protection would also greatly assist the conservation of many sympatric threatened primate taxa. On Bioko this would include the Bioko Preuss's monkey Cercopithecus preussi insularis, the Bioko red-eared monkey C. erythrotis erythrotis, the Golden-bellied crowned monkey C. pogonias pogonias, the Bioko greater white-nosed monkey C. nictitans martini, the Bioko black colobus C. satanas satanas, and the Bioko drill Mandrillus leucophaeus poensis. Protection of *P. pennantii epieni* and *P. preussi* and their habitats on the mainland would benefit populations of Nigeria-Cameroon chimpanzees Pan troglodyes ellioti, Ebo Forest gorillas Gorilla gorilla subsp., Cameroon Preuss's monkey *Cercopithecus preussi preussi*, Nigerian white-throated guenon *Cercopithecus erythrogaster pococki*, Mainland drill *Mandrillus leucophaeus poensis* and Red-capped mangabey *Cercocebus torquatus*.

References

Butynski, T. M. and S. H. Koster. 1994. Distribution and conservation status of primates in Bioko Island, Equatorial Guinea. *Biodiversity and Conservation* 3: 893–909.

Cronin, D. T. 2013. The Impact of Bushmeat Hunting on the Primates of Bioko Island, Equatorial Guinea. Department of Biology. Drexel University, Philadelphia. PA

Cronin, D. T., C. Riaco and G. W. Hearn. 2013. Survey of threatened monkeys in the Iladyi River Valley Region, Southeastern Bioko Island, Equatorial Guinea. *African Primates* 8: 1–8.

Gonzàlez Kirchner, J. P. 1994. *Ecología y Conservación de los Primates de Guinea Ecuatorial*. Ceiba Ediciones, Cantabria, Spain.

Groves, C. P. 2007. The taxonomic diversity of the Colobinae of Africa. *Journal of Anthropological Sciences* 85: 7–34.

Groves, C. P. and N. Ting. 2013. Pennant's red colobus *Piliocolobus pennantii*. In: *Handbook of the Mammals of the World. Volume 3. Primates*, R. A. Mittermeier, A. B. Rylands and D. E. Wilson (eds.), pp.707–708. Lynx Edicions, Barcelona.

Hearn, G., W. A. Morra and T. M. Butynski. 2006. Monkeys in Trouble: The Rapidly Deteriorating Conservation Status of the Monkeys on Bioko Island, Equatorial Guinea. Report, Bioko Biodiversity Protection Program, Glenside, Pennsylvania.

Morra, W., G. Hearn, and A. J. Buck. 2009. The market for bushmeat: *Colobus satanas* on Bioko Island. *Ecological Economics* 68: 2619–2626.

Oates, J. F. 2011. *Primates of West Africa: A Field Guide and Natural History*. Conservation International, Arlington, VA.

Oates, J. F. 1996. African Primates: Status Survey and Conservation Action Plan. Revised edition. IUCN, Gland, Switzerland.

Oates, J. F. and T. T. Struhsaker. 2008. *Procolobus pennantii* ssp. *pennantii*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed on 17 March 2014.

Oates, J. F., T. T. Struhsaker, B. Morgan, J. Linder and N. Ting. 2008. *Procolobus preussi*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 17 March 2014.

Struhsaker, T. T. 2005. The conservation of red colobus and their habitats. *International Journal of Primatology* 26: 525–538.

Struhsaker, T. T. 2011. The Red Colobus Monkeys: Variation in Demography, Behavior, and Ecology of Endangered Species. Oxford University Press, Oxford.

Tana River Red Colobus

Piliocolobus rufomitratus (Peters, 1879) Kenya (2002, 2004, 2006, 2008, 2012)

David N. M. Mbora & Thomas M. Butynski



Tana River red colobus (*Piliocolobus rufomitratus*) (Illustration: Stephen D. Nash)

Gallery forests along the lower Tana River, Kenya, are part of the East African Coastal Forests Biodiversity Hotspot. The forests are the only habitat for two endemic primates: the Tana River red colobus, Piliocolobus rufomitratus, and the Tana River mangabey, Cercocebus galeritus Peters, 1879. Piliocolobus rufomitratus is classified as one of four subspecies of Procolobus rufomitratus on the IUCN Red List of 2008, which is still current. The other three are Procolobus r. oustaleti (Trouessart, 1906), Procolobus r. tephrosceles (Elliot, 1907), and Procolobus r. tholloni (Milne-Edwards, 1886). Here, we follow Groves (2005, 2007; Groves and Ting 2013) in placing all red colobus monkeys in the genus Piliocolobus, and rufomitratus and the other subspecies mentioned above as full species. Piliocolobus rufomitratus is currently classified as Endangered on the IUCN Red List (Butynski et al. 2008). Both the Tana River red colobus and the Tana River mangabey inhabit forest fragments (size range, about 1 ha to 500 ha) along a 60-km stretch from Nkanjonja to Mitapani (01°55'S, 40°05'E) (Butynski and Mwangi 1995; Mbora and Meikle 2004). There are another six sympatric primates in the area, but only the red colobus and mangabey are endemic and entirely forest dependent. The current population of the Tana River red colobus is less than 1,000 individuals and declining, and while the population of the mangabey is a little larger, it too is declining. Indeed, recent genetic analyses have shown that the effective population sizes of the two species are less than 100 individuals (Mbora and McPeek, in revision).

Several factors render the long-term survival of the Tana River red colobus and mangabey bleak and precarious. First, forest is increasingly being cleared for agricultural expansion, and the remaining patches used as a source of building materials and a variety of non-timber forest products. Second, in January 2007, the High Court of Kenya ordered the annulment of the Tana River Primate National Reserve (TRPNR) because, the court

found, that the reserve had not been established in accordance with the law. About half of the remaining forest was legally protected in TRPNR, and therefore no habitat of the Tana River red colobus and mangabey is legally protected at the present time. Third, habitat loss outside the TRPNR was exacerbated by the failure of the Tana Delta Irrigation Project (TDIP). TDIP was a rice-growing scheme managed by the Tana and Athi Rivers Development Authority that had protected forest patches on their land.

Despite the troubles highlighted here, there is reason for hope for the Tana River forests and the endemic monkeys. One of us (Mbora) has maintained a research project in the area over the years. In 2011, Mbora collaborated with Lara Allen to ascertain how and why local people exploited the Tana forests, in order to identify opportunities and constraints for possible conservation action. The study found that the Pokomo people of Tana have a comprehensive traditional system of natural resource use, conservation and management, and a strong desire to preserve the flora and fauna of the forests as part of their heritage. They strongly support community development initiatives related to the conservation of natural resources as these deliver tangible benefits to the people and their environment.

Partly galvanized by the participatory nature of the research, an organization called the Ndera Community Conservancy has now been established in Tana. The sole mission of this formally registered communitybased organization is to protect and conserve about half of the forest patches formerly within the TRPNR, and improve the viability of particular forest patches outside the Reserve. The Ndera Community Conservancy is working with government conservation initiatives and is making some progress. However, in order for the community to make significant progress in enhancing the viability of the habitat of the Tana River red colobus, the support of international conservation agencies is needed. With community structures, government, and the international conservation community working together, the prospects for the long-term viability of Tana River primates can be greatly improved.

References

Butynski, T. M. and G. Mwangi. 1995. Census of Kenya's endangered red colobus and crested mangabey. *African Primates* 1: 8–10.

Butynski, T. M., T. T. Struhsaker and Y. de Jong. 2008. *Procolobus rufomitratus* ssp. *rufomitratus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 17 March 2014.

Groves, C. P. 2005. Order Primates. In: *Mammal Species of the World*, D. E. Wilson and D. M. Reeder (eds), pp.111–184. The Johns Hopkins University Press, Baltimore, MD.

Groves, C. P. 2007. The taxonomic diversity of the Colobinae of Africa. *Journal of Anthropological Sciences* 85: 7–34.

Groves, C. P. and N. Ting. 2013. Tana River red colobus *Piliocolobus rufomitratus*. In: *Handbook of the Mammals of the World. Volume 3. Primates*, R. A. Mittermeier, A. B. Rylands and D. E. Wilson (eds.), p.709. Lynx Edicions, Barcelona.

Mbora, D. N. M. and D. B. Meikle. 2004. Forest fragmentation and the distribution, abundance and conservation of the Tana River red colobus (*Procolobus rufomitratus*). *Biological Conservation* 118: 67–77.

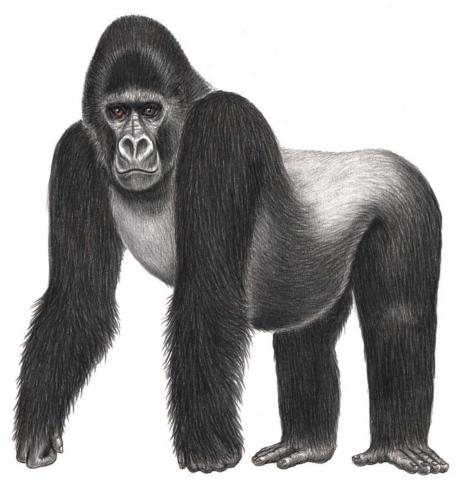
Mbora, D. N. M. and L. Allen. 2011. The Tana Forests 'People for Conservation and Conservation for People' Initiative (PCCP): Preserving the Habitat of the Tana River Red Colobus (*Procolobus rufomitratus*) and the Tana River Mangabey (*Cercocebus galeritus*) Through Community Conservation and Development in Tana River District, Kenya. Final Report on Phase 1, Mohamed bin Zayed Species Conservation Fund, Abu Dhabi.

Mbora, D. N. M and M. A. McPeek. In revision. How monkeys see a forest: population genetic structure in two forest monkeys. *Conservation Genetics*.

Grauer's Gorilla

Gorilla beringei graueri Matschie, 1914 Democratic Republic of Congo 2010, 2012

Stuart Nixon & Elizabeth A. Williamson



Grauer's gorilla (Gorilla beringei graueri) (Illustration: Stephen D. Nash)

The largest of the four subspecies of Gorilla, Grauer's gorilla (Gorilla beringei graueri), is listed on CITES Appendix I and classified as Endangered on the IUCN Red List (Robbins et al. 2008). Grauer's gorillas inhabit mixed lowland forest and the montane forests of the Albertine Rift escarpment in the eastern Democratic Republic of Congo (DRC). Formerly known as the eastern lowland gorilla, the name is misleading as this taxon ranges between approximately 600 m and 2,900 m above sea level. While relatively little is known about the ecology and behaviour of Grauer's gorilla, their diet is rich in herbs, leaves, bark, lianas and vines, seasonallyavailable fruit, bamboo (at the higher altitudes), and invertebrates (e.g., Schaller 1963; Yamagiwa et al. 2005). These gorillas opportunistically raid fields to feed on crops and are often found in regenerating forest

associated with abandoned agricultural clearings, mines and villages (e.g., Schaller 1963; Nixon *et al.* 2006).

Since the 1950s, habitat conversion has been widespread, while a proliferation of 12-gauge shotguns has facilitated the hunting of large mammals, resulting in the local extinction of gorillas in many areas (Emlen and Schaller 1960; P. Anderson, pers. comm.). Threats to their survival were exacerbated throughout the 1990s and early 2000s with persistent conflict in the Great Lakes region of Africa. Refugees, internally displaced people and armed groups settled throughout eastern DRC, putting enormous pressure on natural resources, including in the national parks. Destruction of high-altitude forest for timber and charcoal production continue to threaten the isolated gorilla populations

that persist in the North Kivu highlands, while poaching to feed artisanal miners and associated armed groups presents the most serious and immediate threat. Large numbers of military personnel stationed in rural areas and numerous rebel groups active throughout the region have been heavily implicated in illegal mining activities and facilitate access to the firearms and ammunition that fuel the ongoing civil conflict (United Nations 2010). Kahuzi-Biega National Park (KBNP) is a centre of illegal resource extraction, largely under the control of rebel militia. Meanwhile, DRC's protected area authority (the Congolese Institute for Nature Conservation – ICCN) remains chronically underfinanced and its staff poorly equipped. ICCN faces conflicts not only with local communities but also with armed groups, and highly dedicated ICCN personnel have been killed in the line of duty.

NGOs are working with the government authorities to support protected area rehabilitation and develop conservation programmes, and in 2012 IUCN published a conservation strategy with clear priorities for Grauer's gorillas (Maldonado et al. 2012). The establishment of accurate baselines on the abundance and distribution of this subspecies and the threats to their survival is urgently needed to guide the future implementation of the action plan. Four, broadly-defined population centres are recognized: Maïko-Tayna-Usala (including Maïko National Park and adjacent forests, Tayna Nature Reserve, Kisimba-Ikoba Nature Reserve and the Usala forest), Kahuzi-Kasese (including the lowland sector of KBNP and adjacent forests), and the Itombwe Massif. Additional isolated populations are found in Masisi, the highland sector of KBNP and Mt. Tshiaberimu in Virunga National Park. In the 1990s, Hall et al. (1998) estimated that the total population numbered 8,660-25,500 individuals, despite substantial habitat loss and several localized extinctions. The largest known population, in the lowland sector of KBNP, has since undergone a catastrophic 80% decline (Amsini et al. 2008). Elsewhere, 50% reductions have been documented (Wildlife Conservation Society, unpubl. data) and a recent analysis of ape habitat across Africa estimates that suitable environmental conditions for Grauer's gorillas have declined by 52% since the 1990s (Junker et al. 2012). Acknowledging gaps in our knowledge, data collated during the past 14 years indicate that Grauer's gorilla numbers are likely to been reduced to 2,000-10,000 individuals (Nixon et al. 2012). In collaboration with ICCN and the Max Planck Institute for Evolutionary Anthropology, Leipzig, a

consortium of international NGOs has initiated a twoyear project to assess the status of Grauer's gorilla across its range.

In the face of ongoing political and economic instability in eastern DRC, the threats are likely to remain intense for the foreseeable future, and concerted action to protect Grauer's gorilla is needed. In its favour, a highly localized distribution in discrete populations enables efficient prioritization of valuable resources, and a recent increase in the KBNP highland population (WCS unpublished data) is evidence that highly-targeted conservation efforts can be successful even in the face of acute pressures.

References

Amsini, F., O. Ilambu, I. Liengola, D. Kujirakwinja, J. Hart, F. Grossman and A. J. Plumptre. 2008. *The Impact of Civil War on the Kahuzi-Biega National Park: Results of Surveys between 2000–2008*. Wildlife Conservation Society, Institut Congolais pour la Conservation de la Nature, Kinshasa, DRC.

Emlen, J. T. and G. B. Schaller. 1960. Distribution and status of the mountain gorilla (*Gorilla gorilla beringei*). *Zoologica* 45: 41–52.

Hall, J. S., K. Saltonstall, B.-I. Inogwabini and I. Omari. 1998. Distribution, abundance and conservation status of Grauer's gorilla. *Oryx* 32: 122–130.

Junker, J., S. Blake, C. Boesch, G. Campbell, L. du Toit, C. Duvall, A. Ekobo, G. Etoga, A. Galat-Luong, J. Gamys, J. Ganas-Swaray, S. Gatti, A. Ghiurghi, N. Granier, J. Hart, J. Head, I. Herbinger, T. C. Hicks, B. Huijbregts, I. S. Imong, N. Kumpel, S. Lahm, J. Lindsell, F. Maisels, M. McLennan, L. Martinez, B. Morgan, D. Morgan, F. Mulindahabi, R. Mundry, K. P. N'Goran, E. Normand, A. Ntongho, D. T. Okon. C. A. Petre, A. Plumptre, H. Rainey, S. Regnaut, C. Sanz, E. Stokes, A. Tondossama, S. Tranquilli, J. Sunderland-Groves, P. Walsh, Y. Warren, E. A. Williamson and H. S. Kuehl. 2012. Recent decline in suitable environmental conditions for African great apes. *Diversity and Distributions* 18: 1077–1091.

Maldonado, O., C. Aveling, D. Cox, S. Nixon, R. Nishuli, D. Merlo, L. Pintea and E. A. Williamson. 2012. *Grauer's Gorillas and Chimpanzees in Eastern Democratic Republic of Congo (Kahuzi-Biega, Maïko, Tayna and Itombwe Landscape): Conservation Action Plan 2012–*

2022. IUCN/SSC Primate Specialist Group, Ministry of Environment, Nature Conservation and Tourism, Institut Congolais pour la Conservation de la Nature, and Jane Goodall Institute, Gland, Switzerland.

Nixon, S., E. Emmanuel, K. Mufabule, F. Nixon, D. Bolamba and P. Mehlman. 2006. The Post-conflict Status of Grauer's Eastern Gorilla (*Gorilla beringei graueri*) and Other Wildlife in the Maïko National Park Southern Sector and Adjacent Forests, Eastern Democratic Republic of Congo. Unpublished report, Institut Congolais pour la Conservation de la Nature and Dian Fossey Gorilla Fund International, Goma, DRC.

Nixon. S., A. J. Plumptre, L. Pintea, J. A. Hart, F. Amsini, E. Bahati, Delattre, C. K. Kaghoma, D. Kujirakwinja, J. C. Kyungu, K. Mufabule, R. Nishuli and P. Ngobobo. 2012. The forgotten gorilla; historical perspectives and future challenges for conserving Grauer's gorilla. Abstract #641. XXIV Congress of the International Primatological Society, Cancún, Mexico.

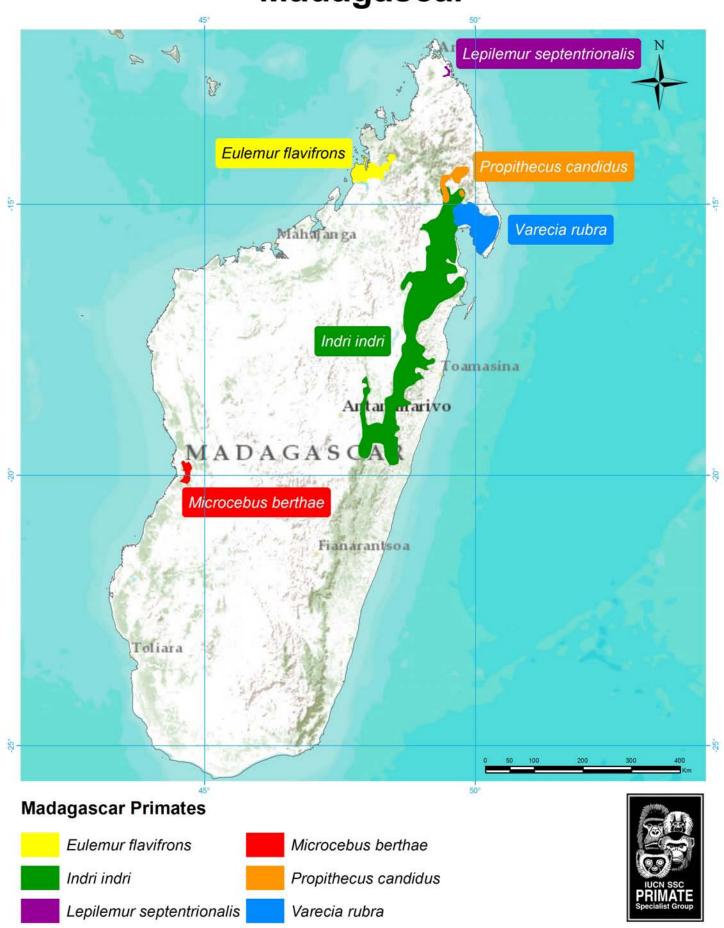
Robbins, M., J. A. Hart, F. Maisels, P. Mehlman, S. Nixon and E. A. Williamson. 2008. *Gorilla beringei* ssp. *graueri*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 16 March 2014.

Schaller, G. B. 1963. *The Mountain Gorilla: Ecology and Behavior*. University of Chicago Press, Chicago, IL.

United Nations. 2010. Final report of the Group of Experts on the Democratic Republic of the Congo. United Nations Security Council, New York.

Yamagiwa, J., A. K. Basabose, K. Kaleme and T. Yumoto. 2005. Diet of Grauer's gorillas in the montane forest of Kahuzi, Democratic Republic of Congo. *International Journal of Primatology* 26: 1345–1373.

Madagascar



Madame Berthe's Mouse Lemur

Microcebus berthae Rasoloarison et al., 2000 Madagascar (2012)

Christoph Schwitzer, Livia Schäffler, Russell A. Mittermeier, Edward E. Louis Jr. & Matthew Richardson



Microcebus berthae, with a body mass of 31 g and a head-body length of 9.0–9.5 cm, is the smallest of the mouse lemurs (and very likely the world's smallest primate; Rasoloarison et al. 2000). It was discovered in the Kirindy Forest in 1992 and was originally thought to be Microcebus myoxinus (Mittermeier et al. 2010). It is found in the central Menabe region of western Madagascar south of the Tsiribihina and north of the

Morondava River (Schmid and Kappeler 1994; Schwab and Ganzhorn 2004). There, it is known to occur in Ambadira Forest and in the Kirindy Classified Forest, as well as in the narrow corridor connecting the two regions (part of the Menabe-Antimena Protected Area). It was formerly found in the Andranomena Special Reserve as well, but has likely been extirpated there.

The species occurs in dry deciduous lowland forest (from sea level to 150 m). It feeds on fruits and gums, and relies heavily on sugary insect excretions and animal matter during the harsh dry season (Dammhahn and Kappeler 2008a). Madame Berthe's mouse lemur has a promiscuous mating system based on testis size, the presence of sperm plugs in females' vaginas, and size dimorphism (Schwab 2000). While both sexes of M. berthae engage in daily periods of torpor, decreasing their metabolism and body temperature to reduce energy expenditure, they do not enter prolonged torpor during the dry season (Ortmann et al. 1997; Schmid et al. 2000). The most common diurnal resting sites of this species are tangles of thin branches surrounded by leaves, but they also use old nests of Mirza, tree holes, and rolled bark found in trees. Sleeping sites are located from 2.5 to 12 m above ground. Males seem to distribute their sleeping sites over a larger area than females, and females reuse the same sleeping site more often than males.

Madame Berthe's mouse lemur appears to be entirely solitary. Individuals do not form sleeping groups and, with the exception of females with young, usually sleep alone. During the night, males and females forage separately. Home ranges, however, are extensively overlapping, with those of the males (4.9 ha) being much larger than those of the females (2.5 ha) (Dammhahn and Kappeler 2005). The nightly path averages 4470 m for males and 3190 m for females. *Microcebus berthae* is sympatric with *M. murinus* in the Kirindy Classified Forest; the two seem to avoid interspecific competition by means of spatial segregation, thereby making the distribution of both rather patchy (Schwab and

Ganzhorn 2004; Dammhahn and Kappeler 2008b). In general, *M. berthae* appears to be more localized than *M. murinus*. Population densities have been estimated at 30–180 individuals/km² in forest patches where it occurs (suggesting high localized densities), but the overall generalized density is about 80 individuals/km² (Schäffler 2012; Schäffler and Kappeler, in press). Population densities in Ambadira Forest tend to be higher than in Kirindy Forest, and the population is largely confined to the most suitable core areas in the interior of the range, far from the range boundary (Schäffler 2012; Schäffler and Kappeler, in press).

Microcebus berthae is classified as Endangered (Andrainarivo et al. 2011). The extent of occurrence covers less than the remnant forest cover of 710 km² (Zinner et al. 2013), and the area of occupancy is considerably smaller than previously assumed based on geographic range borders (Schäffler 2012; Schäffler and Kappeler, in press). The geographic range is severely fragmented, and its extent of occurrence, area of occupancy, and the quality of its habitat are all declining. It is threatened mainly by slash-andburn agriculture and logging, and is particularly sensitive to anthropogenic disturbances. Sensitivity to fragmentation is evident as the species is only found in core areas of extensive forests, and the regional distribution pattern reveals susceptibility to habitat degradation and spatial avoidance of human environments (Schäffler 2012; Schäffler and Kappeler, in press). In 2005 the total population of this species was estimated at no more than 8000 adult individuals living in a handful of forests (Schwab and Ganzhorn 2004), most of which are at higher risk of destruction and fragmentation now than they were 10 years ago. Schäffler and Kappeler (in press) gave a higher estimate of 40,000 individuals, but did not discriminate between adults and juveniles. Pressure on the forests of the central Menabe is strong, and deforestation continues on a large scale. To quantify recent forest loss, Zinner et al. (2013) used a series of satellite images (1973–2010) for estimating annual deforestation rates. The overall rate was 0.67%, but it accelerated to over 1.5% during certain periods, with a maximum of 2.55% per year between 2008 and 2010. Not all areas in the forest block of the central Menabe were affected similarly. Areas surrounding existing clearings showed the highest losses of largely undisturbed forest. If deforestation continues at the same rate as during the last years, 50% of the 1973 forest cover will be gone within the next 11-37 years (Zinner et al. 2013). Madame Berthe's mouse lemur is not being kept in captivity (ISIS 2014).

A conservation action plan for Kirindy-Ambadira (Central Menabe) was published recently (Markolf *et al.* 2013) as part of the IUCN Lemur Conservation Strategy 2013–2016 (Schwitzer *et al.* 2013a). The conservation objectives for this area as laid out in the action plan are additional ecological research and threat analyses of the endemic fauna; improved environmental education; and immediate-term threat mitigation actions such as the introduction of short-cycle chicken farming. Madame Berthe's mouse lemur is also a priority species for *ex situ* conservation measures (Schwitzer *et al.* 2013b).

References

Andrainarivo, C., V. N. Andriaholinirina, A. T. C. Feistner, T. Felix, J. U. Ganzhorn, N. Garbutt, C. Golden, W. B. Konstant, E. E. Louis Jr., D. M. Meyers, R. A. Mittermeier, A. Perieras, F. Princee, J. C. Rabarivola, B. Rakotosamimanana, Rasamimanana, H., J. Ratsimbazafy, G. Raveloarinoro, A. Razafimanantsoa, Y. Rumpler, C. Schwitzer, R. Sussman, U. Thalmann, L. Wilmé and P. C. Wright. 2011. *Microcebus berthae*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 17 March 2014.

Dammhahn, M. and P. M. Kappeler. 2005. Social system of *Microcebus berthae*, the world's smallest primate. *International Journal of Primatology* 26: 407–435.

Dammhahn, M. and P. M. Kappeler. 2008a. Comparative feeding ecology of sympatric *Microcebus berthae* and *M. murinus. International Journal of Primatology* 29: 1567–1589.

Dammhahn, M. and P. M. Kappeler. 2008b. Small-scale coexistence of two mouse lemur species (*Microcebus berthae* and *M. murinus*) within a homogeneous competitive environment. *Oecologia* 157: 473–483.

ISIS. 2014. Zoological Information Management System (ZIMS) version 1.7, released 27 January 2014. ISIS, Apple Valley, MN.

Markolf M., P. M. Kappeler, R. Lewis and I. A. Y. Jacky. 2013. Kirindy–Ambadira (Central Menabe). In: *Lemurs of Madagascar: A Strategy for Their Conservation 2013–2016*, C. Schwitzer *et al.* (eds.), pp.113–115. IUCN SSC Primate Specialist Group, Bristol Conservation and Science Foundation, and Conservation International, Bristol, UK.

Mittermeier, R. A., E. E. Louis Jr., M. Richardson, C. Schwitzer, O. Langrand, A. B. Rylands, F. Hawkins, S. Rajaobelina, J. Ratsimbazafy, R. Rasoloarison, C. Roos, P. M. Kappeler and J. MacKinnon. 2010. *Lemurs of Madagascar*. 3rd edition. Conservation International Tropical Field Guide Series, Conservation International, Arlington, VA.

Ortmann, S., G. Heldmaier, J. Schmid and J. U. Ganzhorn. 1997. Spontaneous daily torpor in Malagasy mouse lemurs. *Naturwissenschaften* 84: 28–32.

Rasoloarison, R. M., S. M. Goodman and J. U. Ganzhorn. 2000. Taxonomic revision of mouse lemurs (*Microcebus*) in the western portions of Madagascar. *International Journal of Primatology* 21: 963–1019.

Schäffler, L. 2012. Determinants of Population Structure in the World's Smallest Primate, *Microcebus berthae*, Across its Global Range in Menabe Central, Western Madagascar. PhD dissertation, Universität Göttingen, Göttingen, Germany.

Schäffler, L. and P. M. Kappeler. In press. Distribution and abundance of the world's smallest primate, *Microcebus berthae*, in Central Western Madagascar. *International Journal of Primatology* 35: DOI: 10.1007/s10764-014-9768-2.

Schmid, J. and P. M. Kappeler. 1994. Sympatric mouse lemurs (*Microcebus* spp.) in western Madagascar. *Folia Primatologica* 63: 162–170.

Schmid, J., T. Ruf and G. Heldmaier. 2000. Metabolism and temperature regulation during daily torpor in the smallest primate, the pygmy mouse lemur (*Microcebus myoxinus*) in Madagascar. *Journal of Comparative Physiology* B 170: 59–68.

Schwab, D. 2000. A preliminary study of spatial distribution and mating system of pygmy mouse lemurs (*Microcebus* cf *myoxinus*). *American Journal of Primatology* 51: 41–60.

Schwab, D. and J. U. Ganzhorn. 2004. Distribution, population structure and habitat use of *Microcebus berthae* compared to those of other sympatric cheirogaleids. *International Journal of Primatology* 25: 307–330.

Schwitzer, C., R. A. Mittermeier, N. Davies, S. Johnson, J. Ratsimbazafy, J. Razafindramanana, E. E. Louis Jr. and S. Rajaobelina (eds). 2013a. *Lemurs of Madagascar: A Strategy for Their Conservation 2013–2016*. IUCN SSC Primate Specialist Group, Bristol Conservation and Science Foundation, and Conservation International, Bristol, UK. 185 pp.

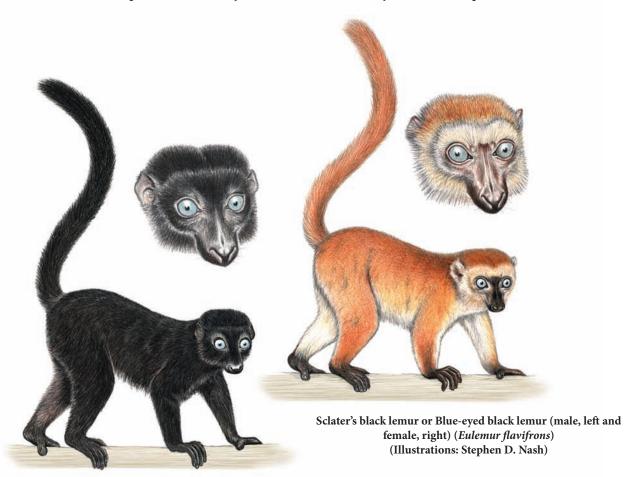
Schwitzer C., T. King, E. Robsomanitrandrasana, C. Chamberlan and T. Rasolofoharivelo. 2013b. Integrating *ex situ* and *in situ* conservation of lemurs. In: *Lemurs of Madagascar: A Strategy for Their Conservation 2013–2016*, C. Schwitzer *et al.* (eds.), pp. 146–152. IUCN SSC Primate Specialist Group, Bristol Conservation and Science Foundation, and Conservation International, Bristol, UK.

Zinner, D., C. Wygoda, L. Razafimanantsoa, R. Rasoloarison, H. T. Andrianandrasana, J. U. Ganzhorn and F. Torkler. 2013. Analysis of deforestation patterns in the central Menabe, Madagascar, between 1973 and 2010. *Regional Environmental Change* 14: 157–166.

Sclater's Black Lemur or Blue-eyed Black Lemur

Eulemur flavifrons (Gray, 1867) Madagascar (2008, 2010, 2012)

Christoph Schwitzer, Guy H. Randriatahina & Sylviane Volampeno



The Blue-eyed black lemur or Sclater's black lemur was rediscovered by science only in 1983 after more than a century of uncertainty about its existence (Koenders et al. 1985; Meier et al. 1996). Its taxonomic validity was thereafter confirmed independently by Rabarivola (1998) as well as Pastorini (2000). The species was until recently regarded as a subspecies of Eulemur macaco, but was elevated to full species status on the basis of the consistency of the morphological differences between the Black lemur and the Blue-eyed black lemur and the pairwise genetic distances between macaco and flavifrons of 68-72 bp (which are in the same range as between the former *E. fulvus* subspecies, i.e., 29–90 bp, according to Pastorini 2000). Furthermore, the fact that the hybrid zone between the two taxa is restricted to just the northeastern part of the distribution of *E. flavifrons* (Andrianjakarivelo 2004; Schwitzer et al. 2005, 2006; Mittermeier *et al.* 2008) favours this new taxonomy.

Eulemur flavifrons occurs only in northwestern Madagascar in a very small area of about 2,700 km² south of the Andranomalaza, north of the Maevarano, and west of the Sandrakota rivers, where it inhabits primary and secondary forest fragments (Koenders et al. 1985; Meyers et al. 1989; Rabarivola et al. 1991). The area of repartition of Eulemur flavifrons lies within a transition zone between the humid Sambirano region in the north and the western dry deciduous forest region in the south, harbouring semi-humid forests with tree heights of up to 30 m on ferruginous alkalescent and alkaline soils based on sandstone, basalt or clay (IRNT 1991a). Average annual precipitation is around 1,600 mm (IRNT 1991b).

There is only a small population of *Eulemur flavifrons* remaining, the majority living in forest fragments on and

adjacent to the Sahamalaza Peninsula (Mouton 1999; Randriatahina and Rabarivola 2004). Rakotondratsima (1999) estimated the population of the Sahamalaza Peninsula to be about 450-2,300 individuals and to have declined by about 35.3% in three years (see also Andriamanandratra 1996). Andrianjakarivelo (2004) found the mean density of E. flavifrons in eight inventoried forest fragments to be 24 individuals per km² (range: 4–85 ind./km²). A total count in two fragments of the Ankarafa Forest on the Sahamalaza Peninsula yielded a density of 60 individuals per km² (Schwitzer et al. 2005, 2007a). Volampeno et al. (2011a) calculated a density of 97 individuals per km² in Ankarafa. However, the density of the species in Ankarafa seems to be higher than in any other forest in the range of E. flavifrons (Randriatahina and Rabarivola 2004). Extrapolating the two density estimates of Andrianjakarivelo (2004) and Schwitzer et al. (2005, 2007a) to the total surface of the terrestrial core zones of the Sahamalaza-Iles Radama National Park (115.8 km²) yields a remaining, severely fragmented population of 2,780-6,950 Blueeyed black lemurs. Eulemur flavifrons was assessed as Critically Endangered (CR A4cd) at the most recent lemur Red List assessment in July 2012, based on a suspected ongoing decline in the area of occupancy and quality of habitat of at least 80% during a 24-year period spanning the past and future (Andrainarivo et al. 2011; Schwitzer et al. 2013). The principal threats to its survival are forest destruction and fragmentation due to slash-and-burn agriculture and selective logging, and continued hunting and trapping, especially in the eastern (mainland) part of its distribution (Gerson 1995; Rakotondratsima 1999; Seiler et al. 2010, 2011/12, 2013). Andrianjakarivelo (2004) found a density of up to 570 traps/km² in certain areas where E. flavifrons occurs.

The Blue-eyed black lemur's home range size and use differs between primary and secondary forest fragments, indicating that it is somewhat able to adapt to different types of habitat. Larger home ranges and lower densities of *E. flavifrons* in secondary forest as compared to primary forest, however, suggest that the former is less suitable habitat for the species (Schwitzer *et al.* 2007a). During a 12-month study, *E. flavifrons* consumed parts of 72 different plant species from 35 families. 52.3% of these were fruits, and 47.7% were leaves. The animals also fed on flowers, insects, insect exudates and fungi (Polowinsky and Schwitzer 2009). *Eulemur flavifrons* exhibits a bimodal activity pattern, which peaks during the morning and evening twilight.

It shows activity bouts during the day and night year-round. Nocturnal illumination and the proportion of illuminated lunar disc are positively associated with the amount of nocturnal activity. Total daily activity, as well as nocturnal activity, is higher in secondary forest than in primary forest (Schwitzer *et al.* 2007b).

Blue-eyed black lemur groups are multi-male multi-female, ranging in size from 6 to 11 individuals, including 3 to 7 adults (Randriatahina and Roeder 2013). Both sexes disperse, but only males have been seen moving into a foreign social group. The sex ratio at birth varies strongly between years and could be male-biased (Randriatahina and Roeder 2013). Births occur between late August and October, at the end of the dry season. During two successive birth seasons, infant mortality was 22.7%. Infants start to become independent at around ten weeks of age (Volampeno *et al.* 2011b).

Parts of the range of Sclater's black lemur officially received protected area status in June 2007 (Parc National Sahamalaza - Iles Radama), including the Sahamalaza Peninsula and some mainland forests to the north and east (Moisson et al. 1999; Lernould 2002; Schwitzer and Lork 2004; Schwitzer et al. 2006). The Sahamalaza Peninsula is also a UNESCO Biosphere Reserve. The Association Européenne pour l'Etude et la Conservation des Lémuriens (AEECL) is a consortium of European zoos that have joined forces to conserve Madagascar's lemurs. AEECL implemented a community-based conservation program in Sahamalaza in December 2000 in order to protect the remaining lemur habitat and to improve the living standards of the local human population. AEECL also maintains a field station in Sahamalaza, which serves as a base for studying the conservation ecology of E. flavifrons and of other lemur species in the area. In 2011, AEECL and Madagascar National Parks started a community-based ecotourism program on the periphery of the protected area.

As of 2014, there were 31 Blue-eyed black lemurs living in European and 30 in North American zoos (ISIS, 2014). The European captive population of the species is being managed in a European Endangered Species Programme (EEP) coordinated by Mulhouse Zoo.

References

Andrainarivo, C., V. N. Andriaholinirina, A. T. C. Feistner, T. Felix, J. U. Ganzhorn, N. Garbutt, C. Golden, W. B. Konstant, E. E. Louis Jr., D. M. Meyers, R. A. Mittermeier, A. Perieras, F. Princee, J. C. Rabarivola, B. Rakotosamimanana, Rasamimanana, H., J. Ratsimbazafy, G. Raveloarinoro, A. Razafimanantsoa, Y. Rumpler, C. Schwitzer, R. Sussman, U. Thalmann, L. Wilmé and P. C. Wright. 2011. *Eulemur flavifrons*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 16 March 2014.

Andriamanandratra, A. N. 1996. Proposition pour un nouveau parc national dans la région du Nord-Ouest de Madagascar: un commencement intégratif. Report to AEECL. Mulhouse: Zoo de Mulhouse.

Andrianjakarivelo, V. 2004. Exploration de la zone en dehors de la peninsule Sahamalaza pour l'évaluation rapide de la population d'*E. m. flavifrons*. Report, WCS Madagascar, Antananarivo.

Gerson, J. S. 1995. The status of *Eulemur macaco flavifrons* at two localities in northwestern Madagascar. *American Journal of Physical Anthropology* Suppl. 20: 98. Abstract.

ISIS. 2014. Zoological Information Management System (ZIMS) version 1.7, released January 27, 2014. ISIS, Apple Valley, MN, USA.

Koenders, L., Y. Rumpler, J. Ratsirarson and A. Peyrieras. 1985. *Lemur macaco flavifrons* (Gray, 1867): a rediscovered subspecies of primate. *Folia Primatologica* 44: 210–215.

Lernould, J.-M. 2002. Un programme international de recherche et de conservation pour le lémur aux yeux turquoise (*Eulemur macaco flavifrons*). *Lemur News* 7: 30–33.

Madagascar, IRNT. 1991a. Carte des ressources en sols. Feuille SC 38 L. Antananarivo: FTM. Projet Inventaire des Ressources Naturelles Terrestres de Madagasikara.

Madagascar, IRNT. 1991b. Carte des ressources en eaux. Feuille SC 38 L. Antananarivo: FTM. Projet Inventaire des Ressources Naturelles Terrestres de Madagasikara.

Meier, B., A. Lonina and T. Hahn. 1996. Expeditionsbericht Sommer 1995 – Schaffung eines neuen Nationalparks in Madagaskar. *Zeitschrift des Kölner Zoo* 39: 61–72.

Meyers, D. M., C. Rabarivola and Y. Rumpler. 1989. Distribution and conservation of Sclater's lemur: implications of a morphological cline. *Primate Conservation* (10): 77–81.

Mittermeier, R. A., J. U. Ganzhorn, W. R. Konstant, K. Glander, I. Tattersall, C. P. Groves, A. B. Rylands, A. Hapke, J. Ratsimbazafy, M. I. Mayor, E. E. Louis Jr., Y. Rumpler, C. Schwitzer and R. M. Rasoloarison. 2008. Lemur diversity in Madagascar. *International Journal of Primatology* 29: 1607–1656.

Moisson, P., Y. Rumpler, J.-M. Lernould and G. Nogge. 1999. Creation of a natural reserve for *Eulemur macaco flavifrons* in the north-west of Madagascar: an update. *Folia Primatologica* 70: 201. Abstract.

Mouton, E. 1999. Mission de terrain sur la presqu'île de Sahamalaza (Nord-ouest Madagascar). Rapport préliminaire pour la création d'une aire protégée. Parc Zoologique et Botanique, Mulhouse. 23pp.

Pastorini, J. 2000. Molecular Systematics of Lemurs. PhD dissertation, Universität Zürich, Zürich. 183pp.

Polowinsky, S. Y. and C. Schwitzer. 2009. Nutritional ecology of the blue-eyed black lemur (*Eulemur flavifrons*): integrating *in situ* and *ex situ* research to assist the conservation of a critically endangered species. In: *Zoo Animal Nutrition Vol. IV*, M. Clauss *et al.* (eds.), pp.169–178. Filander Verlag, Fuerth.

Rabarivola, C. 1998. Etude génétique comparative de populations insulaires et "continentales" de *Eulemur macaco*. Utilisation simultanée des dermatoglyphes, de marqueurs sanguins et de l'ADN (RAPD) pour étudier la différenciation de *E. macaco* en deux sous-espèces: *E. m. macaco* et *E.m. flavifrons*. Doctoral thesis, Université d'Antananarivo, Antananarivo.

Rabarivola, C., D. Meyers and Y. Rumpler. 1991. Distribution and morphological characters of intermediate forms between the black lemur (*Eulemur macaco macaco*) and Sclater's lemur (*Eulemur macaco flavifrons*). *Primates* 32: 269–73.

Rakotondratsima, M. 1999. Etude quantitative de *Eulemur macaco flavifrons* dans la presqu'île Radama. In: *Wildlife Conservation Society Madagascar Country Program: Evaluation de l'état de l'environnement naturel terrestre de la presqu'île Radama*, pp.15–29. Wildlife Conservation Society (WCS), Antananarivo.

Randriatahina, G. H. and J. C. Rabarivola. 2004. Inventaire des lémuriens dans la partie nord-ouest de Madagascar et distribution d'*Eulemur macaco flavifrons*. *Lemur News* 9: 7–9.

Randriatahina, G. H. and J. J. Roeder. 2013. Group size, composition and stability in a wild population of blue-eyed black lemurs (*Eulemur flavifrons*) at Ankarafa, Sahamalaza National Park. In: *Leaping Ahead*, J. Masters, M. Gamba and F. Génin (eds.), pp.127–136. Springer, New York.

Schwitzer, C. and A. Lork. 2004. "Projet Sahamalaza–Iles Radama" Ein internationales Schutzprojekt für den Sclater's Maki (*Eulemur macaco flavifrons* Gray, 1867). *Zeitschrift des Kölner Zoo* 47: 75–84.

Schwitzer, C., R. A. Mittermeier, N. Davies, S. Johnson, J. Ratsimbazafy, J. Razafindramanana, E. E. Louis Jr. and S. Rajaobelina (eds). 2013. *Lemurs of Madagascar: A Strategy for Their Conservation 2013–2016*. IUCN SSC Primate Specialist Group, Bristol Conservation and Science Foundation, and Conservation International, Bristol, UK.

Schwitzer. C., N. Schwitzer, G. H. Randriatahina, C. Rabarivola and W. Kaumanns. 2005. Inventory of the *Eulemur macaco flavifrons* population in the Sahamalaza protected area, northwestern Madagascar, with notes on an unusual colour variant of *E. macaco. Primate Report* Special Issue 72: 39–40. Abstract.

Schwitzer, C., N. Schwitzer, G. H. Randriatahina, C. Rabarivola and W. Kaumanns. 2006. "Programme Sahamalaza": New perspectives for the *in situ* and *ex situ* study and conservation of the blue-eyed black lemur (*Eulemur macaco flavifrons*) in a fragmented habitat. In: *Proceedings of the German-Malagasy Research Cooperation in Life and Earth Sciences*, C. Schwitzer, S. Brandt, O. Ramilijaona, M. Rakotomalala Razanahoera, D. Ackermand, T. Razakamanana and J. U. Ganzhorn (eds.), pp.135–149. Concept Verlag, Berlin.

Schwitzer, N., G. H. Randriatahina, W. Kaumanns, D. Hoffmeister and C. Schwitzer. 2007a. Habitat utilization of blue-eyed black lemurs, *Eulemur macaco flavifrons* (Gray, 1867), in primary and altered forest fragments. *Primate Conservation* (22): 79–87.

Schwitzer, N., W. Kaumanns, P. C. Seitz and C. Schwitzer C. 2007b. Cathemeral activity patterns of the blue-eyed black lemur *Eulemur macaco flavifrons* in intact and degraded forest fragments. *Endangered Species Research* 3: 239–247.

Seiler, M., G. H. Randriatahina and C. Schwitzer. 2010. Ongoing threats to lemurs and their habitat inside the Sahamalaza–Iles Radama National Park. *Lemur News* 15: 7–9.

Seiler, M., G. H. Randriatahina and C. Schwitzer. 2011/12. Rapid boost of forest destruction and poaching of lemurs inside the Sahamalaza–Iles Radama National Park since 2009. *Lemur News* 16: 28–30.

Seiler, M., G. Randriatahina, S. Volampeno and C. Schwitzer. 2013. Sahamalaza–Iles Radama National Park. In: *Lemurs of Madagascar: A Strategy for Their Conservation 2013–2016*, C. Schwitzer *et al.* (eds.), pp.132–134. IUCN SSC Primate Specialist Group, Bristol Conservation and Science Foundation, and Conservation International, Bristol, UK.

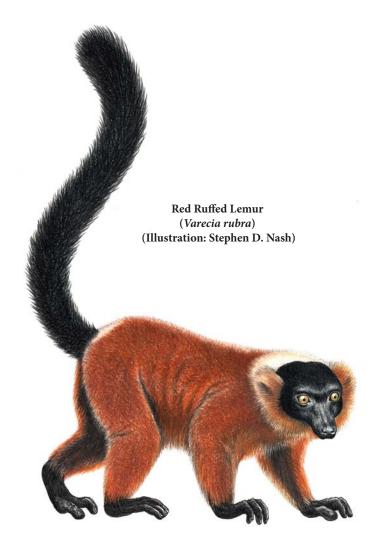
Volampeno, M. N., J. C. Masters and C. T. Downs. 2011a. A population estimate of blue-eyed black lemurs in Ankarafa forest, Sahamalaza–Iles Radama National Park, Madagascar. *Folia Primatologica* 81: 305–314.

Volampeno, M. S. N., J. C. Masters and C. T. Downs. 2011b. Life history traits, maternal behavior and infant development of blue-eyed black lemurs (*Eulemur flavifrons*). *American Journal of Primatology* 73: 474–484.

Red Ruffed Lemur

Varecia rubra É. Geoffroy, 1812 Madagascar (2012)

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The Red ruffed lemur is confined to the Masoala Peninsula and the region immediately north of the Bay of Antongil in northeastern Madagascar (Petter and Petter-Rousseaux 1979; Tattersall 1982). It may have occurred as far north as Antalaha in the past, but this is not certain (Tattersall 1977). The Antainambalana River appears to separate it from *V. variegata subcincta*, and recent surveys have shown that the westernmost distribution of *V. rubra* is near the confluence of the Antainambalana and Sahantaha rivers (Hekkala *et al.* 2007). Variations in color pattern are well known in this species, but have not been attributed to clear geographic regions. It may intergrade with *V. variegata subcincta*; the confluence of the Vohimara and Antainambalana

rivers has been investigated as a possible contact or hybrid zone between the two, but without conclusive results (Tattersall 1982; Lindsay and Simons 1986; Vasey and Tattersall 2002; Hekkala *et al.* 2007).

With a head-body-length of 50–55 cm and a body mass of 3.0-3.6 kg (Vasey 2003), Varecia rubra is a large member of the Lemuridae. It inhabits primary and some secondary moist lowland forest (up to 1200 m above sea level) and prefers tall forest, where it is often observed in the crowns of large feeding trees. The species feeds mainly on fruit, supplemented with flowers, nectar, and leaves. In one study conducted between May and November (Rigamonti 1993), Red ruffed lemurs fed on ripe fruits for 73.9% of their feeding time, flowers for 5.3%, and leaves for 20.9% (18.3% of these mature). Only a few plant species were used as food resources: 72.5% of the observed feeding bouts occurred in only seven tree species. The animals fed on 42 plant species altogether, compared to 106 species that would have been available to them in their home range area. The composition of the diet varied from month to month, but fruits were consistently the main item, even when they were hard to find. The core areas used within their territories always correlated with large, fruit-bearing trees. In the cold-wet season, when few fruits are available, the study group split up into subgroups to use different core areas. Females are reported to eat more low-fibre, high-protein items (young leaves and flowers) prior to giving birth and during lactation, presumably to meet the higher energy demands of reproduction (Vasey 2000a, 2002). At Andranobe, 132 different plant species from 36 families were eaten over the course of a year (Vasey 2000b).

This species has been studied in the forests of Ambatonakolahy (Rigamonti 1993) and Andranobe (Vasey 1997a) on the Masoala Peninsula. Social organization is described as fission/fusion, and communities are usually multimale-multifemale and number 5–31 individuals. Home ranges cover 23–58 ha and appear to be defended (Rigamonti 1993; Vasey

2006). In one study at Andranobe, *V. rubra* spent 28% of its time feeding, 53% resting, and 19% traveling. Females fed more and rested less than males (Vasey 2005). The species is most active during the hot rainy season. Mating occurs in early July, and infants are born in October and fully weaned by February (Vasey 2007).

The Red ruffed lemur is classified as Critically Endangered (Andrainarivo et al. 2011) based on a suspected population reduction of ≥80% over a 3-generation time period of 24 years in the future. The principal threats to the species are habitat loss and hunting (Simons and Lindsay 1987; Rigamonti 1996; Vasey 1996, 1997b). Because of their large size and evident need for tall primary forest, these animals are particularly susceptible to human encroachment, and hunting and trapping for food still takes place. Furthermore, remaining populations are concentrated on the Masoala Peninsula, and they may be threatened by the frequent cyclones that hit this part of Madagascar. The only protected area where Varecia rubra is known to occur is Masoala National Park (Kremen 1998). Masoala was the national park most affected by the very rapid upsurge of illegal logging after the political events of early 2009, and this logging has continued well into 2010. Population density has been variously estimated at 6 individuals/km² (Rakotondratsima and Kremen 2001), 21-23 individuals/km² in Ambatonakolahy (Rigamonti 1993), and 31-54 individuals/km² in Andranobe (Vasey 1997b).

The IUCN lemur conservation strategy 2013–2016 (Schwitzer *et al.* 2013) proposes a suite of conservation measures for Masoala National Park to ensure the conservation of the Red ruffed lemur: further patrols and surveillance; campaigns of environmental education and awareness; and support for small-scale husbandry of domestic animals as a source of protein. As of 2014, there were 590 Red ruffed lemurs reported in captivity worldwide (ISIS 2014). Such populations in American and European zoos represent a safeguard against extinction, but they are unfortunately very limited in their genetic diversity (Schwitzer 2003).

References

Andrainarivo, C., V. N. Andriaholinirina, A. T. C. Feistner, T. Felix, J. U. Ganzhorn, N. Garbutt, C. Golden, W. B. Konstant, E. E. Louis Jr., D. M. Meyers, R. A. Mittermeier, A. Perieras, F. Princee, J. C. Rabarivola, B. Rakotosamimanana, Rasamimanana, H.,

J. Ratsimbazafy, G. Raveloarinoro, A. Razafimanantsoa, Y. Rumpler, C. Schwitzer, R. Sussman, U. Thalmann, L. Wilmé and P. C. Wright. 2011. *Varecia rubra*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. www.iucnredlist.org. Accessed 17 March 2014.

Hekkala, E. R., M. Rakotondratsima and N. Vasey. 2007. Habitat and distribution of the ruffed lemur, *Varecia*, north of the Bay of Antongil in north-eastern Madagascar. *Primate Conservation* (22): 89–95.

ISIS. 2014. Zoological Information Management System (ZIMS) version 1.7, released 27 January 2014. ISIS, Apple Valley, MN.

Kremen, C. 1998. Madagascar creates its largest protected area on the Masoala Peninsula. *Lemur News* 3: 1–3.

Lindsay, N. B. D. and H. J. Simons. 1986. Notes on *Varecia* in the northern limits of its range. *Dodo* 23: 29–24.

Petter, J.-J. and A. Petter-Rousseaux. 1979. Classification of the prosimians. In: *The Study of Prosimian Behavior*, G. A. Doyle and R. D. Martin (eds.), pp.359–409. Academic Press, New York.

Rakotondratsima, M. and C. Kremen. 2001. Suivi écologique de deux espèces de lémuriens diurnes *Varecia variegata rubra* et *Eulemur fulvus albifrons* dans la presqu'île de Masoala (1993–1998). *Lemur News* 6: 31–35.

Rigamonti, M. M. 1993. Home range and diet in red ruffed lemurs (*Varecia varigata rubra*) on the Masoala Peninsula, Madagascar. In: *Lemur Social Systems and their Ecological Basis*, P. M. Kappeler and J. U. Ganzhorn (eds.), pp.25–39. Plenum Press, New York.

Rigamonti, M. M. 1996. Red ruffed lemur (*Varecia variegata rubra*): a rare species from the Masoala rain forests. *Lemur News* 2: 9–11.

Schwitzer, C. 2003. Energy Intake and Obesity in Captive Lemurs (Primates, Lemuridae). Doctoral thesis, Universität zu Köln. Schüling Verlag, Münster, Germany.

Schwitzer, C., R. A. Mittermeier, N. Davies, S. Johnson, J. Ratsimbazafy, J. Razafindramanana, E. E. Louis Jr. and S. Rajaobelina (eds.). 2013. *Lemurs of Madagascar: A Strategy for Their Conservation 2013–2016*. IUCN SSC Primate Specialist Group, Bristol Conservation and Science Foundation, and Conservation International, Bristol, UK. 185pp.

Simons, H. J. and N. B. D. Lindsay. 1987. Survey work on ruffed lemurs (*Varecia variegata*) and other primates in the northeastern rain forests of Madagascar. *Primate Conservation* (8): 88–91.

Tattersall, I. 1977. Distribution of the Malagasy lemurs part 1: The lemurs of northern Madagascar. *Annals of the New York Academy of Sciences* 293: 160–169.

Tattersall, I. 1982. *The Primates of Madagascar*. Columbia University Press, New York.

Vasey, N. 1996. Clinging to life: *Varecia variegata rubra* and the Masoala coastal forests. *Lemur News* 2: 7–9.

Vasey, N. 1997a. Community Ecology and Behavior of *Varecia variegata rubra* and *Lemur fulvus albifrons* on the Masoala Peninsula, Madagascar. PhD thesis, Washington University, St. Louis, MO.

Vasey, N. 1997b. How many red ruffed lemurs are left? *International Journal of Primatology* 18: 207–216.

Vasey, N. 2000a. Niche separation in *Varecia variegata rubra* and *Eulemur fulvus albifrons*: I. Interspecific patterns. *American Journal of Physical Anthropology* 112: 411–431.

Vasey, N. 2000b. Plant species composition of diet in two sympatric lemurs: *Varecia variegata rubra* and *Eulemur fulvus albifrons*. *American Journal of Physical Anthropology* 30 (suppl.): 309–310.

Vasey, N. 2002. Niche separation in *Varecia variegata rubra* and *Eulemur fulvus albifrons*: II. Intraspecific patterns. *American Journal of Physical Anthropology* 118: 169–183.

Vasey, N. 2003. *Varecia*, ruffed lemurs. In: S. M. Goodman and J. P. Benstead (eds), *The Natural History of Madagascar*, pp.1332–1336. Aldine de Gruyter, New York.

Vasey, N. 2005. Activity budgets and activity rhythms in red ruffed lemurs (*Varecia rubra*) on the Masoala Peninsula, Madagascar: seasonality and reproductive energetics. *American Journal of Primatology* 66: 23–44.

Vasey, N. 2006. Impact of seasonality and reproduction on social structure, ranging patterns, and fission-fusion social organization in red ruffed lemurs. In: L. Gould and M. A. Sauther (eds), *Lemurs: Ecology and Adaptation*, pp.275–304. Springer, New York.

Vasey, N. 2007. The breeding system of wild red ruffed lemurs (*Varecia rubra*): a preliminary report. *Primates* 48: 41–54.

Vasey, N. and I. Tattersall. 2002. Do ruffed lemurs form a hybrid zone? Distribution and discovery of *Varecia*, with systematic and conservation implications. *American Museum Novitates* (3376): 1–26.

Northern Sportive Lemur

Lepilemur septentrionalis Rumpler and Albignac, 1975 Madagascar (2008, 2010, 2012)

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Originally described based on cytogenetic and morphometric characteristics (Rumpler and Albignac 1975), the taxonomic status of the Northern sportive lemur (Lepilemur septentrionalis) has since been supported by more detailed cytogenetic, morphogenetic and especially molecular data (Ravoarimanana et al. 2004; Andriaholinirina et al. 2006; Louis et al. 2006), and subsequently accepted in recent taxonomic revisions of primates (Groves 2001, 2005) and lemurs (Mittermeier et al. 2008, 2010). With the taxonomic revision confirming L. septentrionalis and L. ankaranensis as distinct species, the perceived range of the Northern sportive lemur was drastically reduced; limited to a few degraded patches of dry forest in the Sahafary region just south of Antsiranana. The number of animals observed during surveys has decreased dramatically over a six-year period. The first survey was performed in 2001 by I. Ravoarimanana and the second in 2007 by A. Zaramody in the Andrahona, Ankarakataova, and Sahafary regions. The population was estimated at about 120 individuals; mainly in the Sahafary area.

Expeditions by Omaha's Henry Doorly Zoo and Aquarium (OHDZA) and the Madagascar Biodiversity Partnership (MBP) in 2010 and 2011 verified the continued existence of the Northern sportive lemur but with a tremendous decline in the Sahafary classified forest, and not a single animal was detected in the Analalava forest where it had been seen in 2005. One individual was reported, however, when Analalava was revisited in July 2012 (Ranaivoarisoa et al. 2013). Fortunately, Ranaivoarisoa et al. (2013) confirmed the presence of the Northern sportive lemur in Montagne des Français (MDF) in 2010, but could only identify 19 individuals across its range based on capture and direct visual observations. Further surveys of the Montagne des Français region in 2012–2013 by OHDZA and MBP that included the previously known habitats of Sahafary and Analalava classified forests through to its northern extent in MDF, documented only 52 L. septentrionalis individuals, with 95% of these lemurs located in MDF. The most recent population estimates based on only



Northern sportive lemur (Lepilemur septentrionalis) (Illustration: Stephen D. Nash)

capture surveys in 2013 in the Montagne des Français area provided the following population estimates:
1) Abatoire - 7 individuals; 2) Andranonakomba - 2 individuals; 3) Ampamakiampafana - 11 individuals; 4) Ambatobe - 2 individuals; and Berambo - 5 individuals for a total of 27 individuals; this species was documented at Anketrakala and Ampitsinjozatsambo in 2012, which were not recently surveyed.

In 2008, the Service d'Appui à la Gestion de l'Environnement promoted the designation of Montagne des Français as a newly protected area, and supported the development of a Vondron'Olona Ifototra (VOI) in Andavakoera, the primary village of this mountain forest. However, sustained human encroachment from the city of Antsiranana continues to finance the production of charcoal and collection of sand, activities that are threatening this last remaining northern sportive lemur population (Ranaivoarisoa et al. 2013). Thus, habitat loss from uncontrolled long-term slashand-burn practices and the conversion of the remaining endemic forest are the primary challenges to overcome. The Northern sportive lemur is nocturnal, spending the day sleeping in tree holes, and very little is known about its ecology and behaviour. However, recent work has shown that its folivorous diet and predilection for new-growth leaves complicates any attempts or plans to maintain it in captivity. Currently, there is no record of any sportive lemur held in any zoological park, as all known attempts to maintain them in captivity have failed; on average within one week of capture. In situ conservation programmes and community-based interactions are, therefore, the only viable solutions. The combination of a very small range composed of rapidly deteriorating suitable habitat with high pressure from hunting puts the Critically Endangered Northern sportive lemur (Andrainarivo et al. 2011) on the cusp of extinction.

References

Andrainarivo, C., V. N. Andriaholinirina, A. T. C. Feistner, T. Felix, J. U. Ganzhorn, N. Garbutt, C. Golden, W. B. Konstant, E. E. Louis Jr., D. M. Meyers, R. A. Mittermeier, A. Perieras, F. Princee, J. C. Rabarivola, B. Rakotosamimanana, Rasamimanana, H., J. Ratsimbazafy, G. Raveloarinoro, A. Razafimanantsoa, Y. Rumpler, C. Schwitzer, R. Sussman, U. Thalmann, L. Wilmé and P. C. Wright. 2011. *Lepilemur septentrionalis*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 16 March 2014.

Andriaholinirina, N., J.-L. Fausser, C. Roos., D. Zinner, U. Thalmann, C. Rabarivola, I. Ravoarimanana, J. U. Ganzhorn, B. Meier, R. Hilgartner, L. Walter., A. Zaramody, C. Langer, T. Hahn, E. Zimmermann, U. Radespiel, M. Craul, J. Tomiuk, I. Tattersall and Y. Rumpler. 2006. Molecular phylogeny and taxonomic revision of the sportive lemurs (*Lepilemur*, Primates). *BMC Evolutionary Biology* 6: 17.

Groves, C. P. 2001. *Primate Taxonomy*. Smithsonian Institution Press, Washington, DC.

Groves, C. P. 2005. Order Primates. In: *Mammal Species of the World: A Taxonomic and Geographic Reference*, 3rd edition, D. E. Wilson and D. M. Reeder (eds.), pp.111–184. Johns Hopkins University Press, Baltimore, MD.

Louis Jr., E. E., S. E. Engberg, R. Lei, H. Geng, J. A. Sommer, R. Randriamampionona, J. C. Randriamanana, J. R. Zaonarivelo, R. Andriantompohavana, G. Randria, R. B. Prosper, G. Rakotoarisoa, A. Rooney and R. A. Brenneman. 2006. Molecular and morphological analyses of the sportive lemurs (Family Megaladapidae: genus *Lepilemur*) reveals 11 previously unrecognized species. *Texas Tech University Museum*, *Special Publications* (49): 1–47.

Mittermeier, R. A., J. U. Ganzhorn, W. R. Konstant, K. Glander, I. Tattersall, C. P. Groves, A. B. Rylands, A. Hapke, J. Ratsimbazafy, M. I. Mayor, E. E. Louis Jr., Y. Rumpler, C. Schwitzer and R. M. Rasoloarison. 2008. Lemur diversity in Madagascar. *International Journal of Primatology* 29: 1607–1656.

Mittermeier, R. A., E. E. Louis Jr., M. Richardson, C. Schwitzer, O. Langrand, A. B. Rylands, F. Hawkins, S. Rajaobelina, J. Ratsimbazafy, R. Rasoloarison, C. Roos, P. M. Kappeler and J. Mackinnon. 2010. *Lemurs of Madagascar*, 3rd edition. Conservation International Tropical Field Guide Series, Arlington, VA.

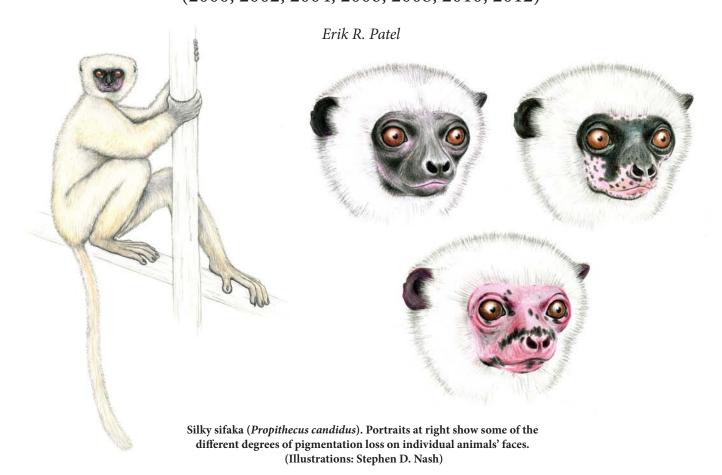
Ranaivoarisoa, J. F., J. R. Zaonarivelo, R. Lei, S. E. Johnson, T. M. Wyman, R. A. Mittermeier, and E. E. Louis Jr. 2013. Rapid survey and assessment of the northern sportive lemur, *Lepilemur septentrionalis*, in northern Madagascar. *Primate Conservation* (27): 23–31.

Ravoarimanana, I. B., R. Tiedemann, D. Montagnon and Y. Rumpler. 2004. Molecular and cytogenetic evidence for cryptic speciation within a rare endemic Malagasy lemur, the northern sportive lemur (*Lepilemur septentrionalis*). *Molecular Phylogenetics and Evolution* 31: 440–448.

Rumpler, Y. and R. Albignac. 1975. Intraspecific chromosome variability in a lemur from north of Madagascar: *Lepilemur septentrionalis*, species nova. *American Journal of Physical Anthropology* 42: 425–429.

Silky Sifaka

Propithecus candidus Grandidier, 1871 Madagascar (2000, 2002, 2004, 2006, 2008, 2010, 2012)



Propithecus candidus is a large white sifaka from northeastern Madagascar. Silky sifakas were recently raised to full species status (Groves and Helgen 2007; Mayor et al. 2002, 2004; Mittermeier et al. 2010), although debates remain about the number of sifaka species (reviewed in Tattersall 2007). Head-body length ranges from 48-54 cm, tail length 45-51 cm, total length 93-105 cm, and adult body weight from 5-6.9 kg (Lehman et al. 2005; Gordon et al. 2013). The pelage is long, silky and white, which gives this species its common name. In some individuals, silver-gray or black tints may appear on the crown, back and limbs, and the pygal region (at the base of the tail) is sometimes yellow. The muzzle and face are bare, and tips of the naked black ears protrude just beyond the white fur of the head and cheeks. Although all infants are born with black skin, all individuals lose pigmentation to varying degrees as they get older, resulting in skin color which is a mix of pink and black, with some individuals having all pink or all black faces. The extent of pigment loss is more extreme than in any other lemur, which led early explorers (e.g., Milne-Edwards and Grandidier 1875) to believe that it was an albino subspecies of *P. diadema*. The lack of red eyes or photophobia and the presence of some black fur, however, confirm that silky sifakas are not albino, but are a leucistic species which may suffer from a heritable vitiligo-like skin disorder (K. Linder, pers. comm.; Cousins 2007).

Unlike *Propithecus perrieri* and *P. edwardsi*, where adult males and females are difficult to distinguish, adult male and female *P. candidus* can be readily distinguished from one another by the pelage coloration of the upper chest. Adult males have a large brown "chest patch" that results from chest scent-marking with the sternal gular gland. As rates of male chest scent marking increase during the mating season, male chest patches become larger and can cover the entire front torso to the abdomen (Patel 2006a).

Propithecus candidus has been classified as Critically Endangered on the IUCN Red List since 1996

(Andrainarivo et al. 2011). This is one of the rarest and most endangered lemurs. Global population size is currently estimated to be between 300 and 2000. Line-transect surveys underway in 2014 will provide a more population estimate, even though surveying is challenging because the Silky sifaka is patchily distributed and rare; sightings are few. Encounter rates have been obtained from several sites in Marojejy and are presented below. Silky sifakas are hunted throughout their range as there is no local taboo, or fady, against eating them. Sadly, hunting of lemurs has increased in recent years across Madagascar. Even species once thought to be protected by taboo, such as *Indri*, are frequently hunted in some regions (Jenkins et al. 2011; Nielson and Patel 2008). Habitat disturbance, such as slash-and-burn agriculture (tavy), logging of precious woods (for example, rosewood) cutting trees for firewood and construction also occurs in and adjacent to the protected areas where they are found (Patel 2007a; Patel et al. 2005). Illegal mining of crystal (quartz) and amethyst has increased recently in Anjanaharibe-Sud (primarily) and Marojejy, leading to a two-month closure of Anjanaharibe-Sud to tourism and research in 2012. Of greatest concern is the recent resurgence of illegal rosewood logging (with associated bushmeat hunting) in Marojejy by armed gangs of loggers which, after a several year reprieve, began again in December 2013. Similar park regions that have been impacted in the past are now again being logged, including the Andratamarina, Andrahanjo, and Mandena tourist regions. A new conservation project by the Duke Lemur Center, called 'SAVA Conservation', which began in January 2012, is working to ameliorate some of these conservation threats. Based in Sambava, the project works with Madagascar National Parks, and follows a multi-faceted community-based approach to biodiversity conservation, similar to that used by the Madagascar Fauna Group (MFG).

The Silky sifaka has a restricted range in northeastern Madagascar that includes the humid forest belt extending from Maroantsetra to the Andapa Basin and the Marojejy and Anjanaharibe Massifs, including the Marojejy Anjanaharibe-Sud Tsaratanana corridor (COMATSA). The Marojejy National Park marks the northern limit of its distribution, although remarkably WWF and SAHALA, a Malagasy environmental association, have recently discovered several groups at the Andrakengy site in the Tsaratanana Corridor approximately 25 km northwest (300°) of Doany, just south of the Androranga River (Sandrine Rasarimanana, pers. comm.). There are

also unconfirmed reports of silky sifakas in the Anjiabe region (just south of Andrakengy) in the Tsaratanana Corridor and at several sites in the Betaolana Corridor, including Antsahabe, Befamatra, Ambodihasina, Antanambe, Ambodimandresy, and Ambodivohitra (Fara and Andriamarasolo 2010; Andriamarasolo, pers. comm.; BioDev/WWF 2011).

The Antainambalana River, in the Makira Natural Park, is the southern range limit for the species. As of 2014, two small groups of Silky sifakas (a group of two and the other of three) have been found in an unusually low elevation forest (235 m to 565 m above sea level) at the new WCS-managed Silky sifaka site at Andaparaty (Rabeson) just north of the Antainambalana River. Recent infrastructure developments at that site should encourage more tourism and research. Silky sifakas may occur in northeastern Makira (Besariaka, Amparihibe, Bezavona), although they have yet to be observed there (Milne-Edwards and Grandidier 1875; Tattersall 1982; Wilme and Callmander 2006; Rasolofoson et al. 2007; Patel and Andrianandrasana 2008). Surveys in recent years by SIMPONA (the main NGO in the Marojejy National Park in charge of conservation, research and social development) have revealed the presence of several groups of Silky sifakas just outside Makira in the disturbed forests of Maherivaratra (near Ambavala town) and about 10 km to the northwest in the Antohakalava forest (near Anivorano village) (pers. obs.) which is a slight enlargement of their known geographic range. Hunting and slash-andburn agriculture are widespread in Antohakalava and Maherivaratra, placing the remaining silkies there under heavy anthropogenic pressure.

The majority of the remaining population of *P. candidus* is found in just two protected areas managed by Madagascar National Parks (Andapa): Marojejy National Park and Anjanaharibe-Sud Special Reserve. Only perhaps a dozen groups have been found in the (WCS-managed) Makira Natural Park at a few widely separated sites: Andaparaty (noted above), Manandriana (near the western edge of Anjanaharibe-Sud), and Soavera (south of Manandriana) (Rasolofoson *et al.* 2007; Ratelolahy and Raivoarisoa 2007). At least a dozen or more groups are found in the COMATSA forests, managed by WCS, which are expected to be official protected areas soon.

More published results from formal line-transect surveys are clearly needed, particularly in Anjanaharibe-Sud

where there have been no quantitative lemur surveys since 1994 (Schmid and Smolker 1998). Moreover, very little is known about the newly protected western extension of Anjanaharibe-Sud. In Marojejy, recent intensive, line-transect surveys in central-eastern ("tourist zone") and northwestern Marojejy have returned group encounter rates of 0.0357 groups/km and 0.0238 groups/km, respectively (Moore and Patel 2013). A similarly low encounter rate of 0.0137 groups/ km was obtained during a three-month line-transect survey in northwestern Marojejy in 2007. At least a dozen groups were found in far western Marojejy near the village of Antsahaberaoka in 2007. The classic and highly cited Marojejy lemur survey by Sterling and McFadden (2000) found Silky sifakas as high as 1875 m above sea level. Only the Andaparaty group in Makira has been found inhabiting forests below 700 m above sea level. The Marojejy expedition undertaken by Duckworth et al. (1995) found Silky sifakas in the northwest and southeast sectors of the park, but not in the lower elevation southwest. Goodman and colleagues from WWF have also documented the presence of P. candidus in COMATSA (Goodman et al. 2003; Fara and Andriamarasolo 2010).

A number of studies have examined the behavioural biology, communication, and feeding ecology of Silky sifakas in Marojejy National Park and the Makira Natural Park. Silky sifakas show the greatest elevational range of any of the sifakas; as low as 235 m above sea level in the Makira (Andaparaty) and as high as 1,875 m in Marojejy. Thus, they inhabit several types of elevation-specific habitats including primary montane rainforest, sclerophyllous forest, and even low ericoid bush at their highest elevations. Their large size (as one of the two largest sifaka species with *P. diadema*) and thick pelage may be adaptations for cold climates at high elevations (Lehman *et al.* 2005).

The social structure of *P. candidus* is variable. They are mostly found in male-female pairs and one-male groups, but occasionally in multi-male/multi-female groups. Groups range in size from 2 to 9 and are cohesive, with inter-individual distances seldom exceeding 25 m. Home ranges vary considerably by site, and are smaller for the Marojejy Camp 2 group inhabiting undisturbed primary forest (100% MCP = 57.2 ha and 95% Kernal = 41.4 ha) than the main group at Andaparaty in Makira in a disturbed forest and unusually low elevation forest (100% MCP = 98.6 ha and 95% Kernal = 67.8 ha). Remarkably, the Camp 2 group ranges from 700 m to

1200 m above sea level, thereby exhibiting a 500-m elevational range within their home range, exceeding that of any other known sifaka group in Madagascar.

Approximately 25% of the day is spent feeding, 44% resting, and the remainder is devoted to social behaviour (16.8%), travelling, and sleeping. Long bouts of terrestrial play involving adults are not uncommon. Rates of aggression are low, and occur mainly during feeding. Females have feeding priority over males. A recent 12-month study at Marojejy Camp 2 documented feeding from more than 100 types of trees, vines, epiphytic ferns, epiphytic hemi-parasitic plants, a few terrestrial parasitic plants, as well as soil. Plant samples were collected, dried, and identified by a botanist. The top ten foods in order of percentage of feeding time were: 10.8% Hazinina (Symphonia sp.), 10.1% Lalona (Weinmannia sp.), 6.8% Vahindrobanga (Landolphia sp.), 6.2% Volomborona (Albizia polyphylla), 6.0% Soretry (Plagioscyphus sp.), 5.1% Rotro (Eugenia sp.), 3.9% Vahivy (Dichapetalium madagascariense), 3.8% Taintsitsihy (Backerella clavata), 3.3% Fotsidity (Ficus polita), 3.3% Nanto (Mimusops sp.). Plant part percentages confirm that this species is a folivore/seed predator, with most of their diet comprised of leaves (47.7%) and seeds (31.4%). Fruits (10.6%), flowers (9.8%), and stems (0.5%) were also regularly consumed. By contrast, the most commonly consumed food by the Silky sifakas at the low elevation (235 m to 565m above sea level) Andaparaty (Makira) site was Mampay (Fabaceae, 22.3%), which has never been observed bring eaten by Silky sifakas in Marojejy (Rajaonarison et al. 2012).

Mating has been observed and occurs on a single day each year in December or January. Infants are born in June or July. Females generally give birth to a single offspring every two years, although births in consecutive years have been observed (Patel 2006b). Infants initially grasp the fur on their mother's belly, and only about four weeks later begin to ride "jockey style" on their mothers back. As is typical of *Propithecus*, all group members interact affiliatively with infants. Grooming is the most frequent form of non-maternal infant care, followed by playing, occasional carrying, as well as nursing in a few remarkable instances (Patel 2007b). Dispersal is bisexual and has been observed on three occasions.

Other than humans, only the fossa (*Cryptoprocta ferox*) has been documented as a predator of the Silky sifaka (Patel 2005). No aerial predation attempts by raptors have

ever been observed, although these sifakas sometimes stare skyward and emit loud "aerial disturbance" roars in the presence of the large Madagascar buzzard (Buteo brachypterus), which does not, however, eat lemurs, only small birds. Loud sneeze-like "zzuss!" vocalizations are their second type of alarm call, and are emitted in response to terrestrial disturbances and to lost calls by other group members, as well as after receiving aggression. A detailed acoustic analysis (160 calls from nine adults in three groups) has revealed sex and individual differences in the acoustic structure of the silky sifaka "zzuss" vocalization. Male and female zzuss calls differed most in F0- and amplitude related features, characteristics that are relatively unconstrained by overall body size. All measures differed among individual callers, with F0-related variables again playing the largest role. Based on usage, these calls most likely function both as generalized alarm and groupcoordination signals.

As in all prosimians, olfactory communication is well developed. Eastern sifakas have several specialized scent-marking glands that include a sebaceous chest gland only found in males and mixed apocrinesebaceous genital glands in both sexes (Schilling 1979). Sifakas do not allomark, as in Eulemur, by directly scent-marking conspecifics. Females scent-mark trees by rubbing their genital glands in a rhythmic vertical motion. Males scent-mark trees by rubbing them with their chest gland or genital glands, or a combination of the two. Males routinely bite or gouge trees with their toothcombs just prior to chest-marking, which leaves long-lasting visible marks. Silky sifakas do not eat bark or gum, so such non-nutritive male bark-biting is likely communicative in function. A recent study in Marojejy found that most of the 102 gouged tree, vine, and epiphyte species were food species (61.8%), and many were known to be sleeping trees (38.2%). Multiple regression analysis revealed that the number of gouges per tree species was predicted by the percentile rank of those species as food tree species and sleeping tree species. As first described by Powzyk (1997), barkbiting likely promotes scent longevity, attracts the visual attention of conspecifics, and in some cases remove the scent marks of conspecifics. Male bark-biting may also result in the deposition of saliva cues and may be considered an honest signal of male status (Patel 2012a; Patel and Girard-Buttoz 2008).

A recent six-month scent marking study at Camp 2 of Marojejy examined marking rates and the territorial scent mark function. Mean focal scent mark rate for adult silky sifakas was 1.64 marks/hr, and the mean adult male mark rate (3.6 marks/hr) was more than five times higher than the adult females (0.7 marks/hr). For the adult male, combined chest-genital marking accounted for 40.1% of his scent marks, followed by genital marks at 35.3% and chest marks at 24.6%. Some evidence for territorial scent marking was found. GPS points were recorded for 1549 focal scent marks over six months; although the effect was moderate, more marks were deposited near the periphery of the home range than near the core, particularly for the adult male (Patel 2013). A one-year study of conspecific responsiveness to intragroup scent marks found that only 17% of male P. candidus marks are responded to by other group members but 71% of female marks received a response, on average within 61 seconds (Patel 2006a). In both P. edwardsi and P. candidus, male overmarking of a female's mark is the most common response, followed by males overmarking the scent marks of other males. Male eastern sifakas preferentially use one type of scent marking, combined chest-ano-genital marking, when depositing an overmark (Andrianandrasana et al. 2007).

Recent preliminary work has examined Silky sifaka parasites. Over 100 fecal samples from the Silky sifakas in the Marojejy Camp 2 group (mainly) and Andaparaty (Makira) group were examined for endoparasites by several experienced labs. Ectoparasites were also noted during biomedical field exams. One of the Makira sifakas was infected with Lemurstrongylus sp. and all harboured the ectoparasite Listrophriodes sp. From the larger Marojejy sample, eggs of two species of nematodes were recovered (Lemurstrongylus sp. and Lemuricola sp.), a tapeworm (Bertiella sp.), an unknown oocyst, and two species of ectoparasites (Gaudalges and Listrophriodes sp.). ELISAs did not detect Giardia sp. or Cryptosporidium sp. antigens among the Marojejy or Makira sifakas, despite the fact that captive sifaka populations frequently harbour such infections (Loudon et al. 2013; Patel 2012b).

References

Andrainarivo, C., V. N. Andriaholinirina, A. T. C. Feistner, T. Felix, J. U. Ganzhorn, N. Garbutt, C. Golden, W. B. Konstant, E. E. Louis Jr., D. M. Meyers, R. A. Mittermeier, A. Perieras, F. Princee, J. C. Rabarivola, B. Rakotosamimanana, Rasamimanana, H., J. Ratsimbazafy, G. Raveloarinoro, A. Razafimanantsoa, Y. Rumpler, C. Schwitzer, R. Sussman, U. Thalmann, L. Wilmé and P. C. Wright. 2011. *Propithecus candidus*.

In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. www.iucnredlist.org>. Accessed 16 March 2014.

Andrianandrasana, L. H., E. R. Patel and P. C. Wright. 2007. A comparison of scent overmarking in two species of wild rainforest sifakas: silky sifakas (*Propithecus candidus*) and Milne-Edwards' sifakas (*Propithecus edwardsi*). Abstracts of the 2007 International Congress on Prosimians, pp.54–55. Ithala Game Reserve, Louwsberg, South Africa. Abstract.

BioDev/WWF 2011. Rapport Provisoire: Inventaire Biologique COMATSA. 176pp.

Cousins, D. 2007. Albinism and leucism in primates. *International Zoo News* 54: 134–145.

Duckworth, J. W., M. I. Evans, A. F. A. Hawkins, R. J. Safford and R. J. Wilkinson. 1995. The lemurs of Marojejy Strict Nature Reserve, Madagascar: a status overview with notes on ecology and threats. *International Journal of Primatology* 16: 545–559.

Fara, L.R. and I. Andriamarasolo. 2010. Conservation des lémuriens via la protection de leurs habitat et le développement communautaire dans les corridors de Betaolana et Tsaratanana-Betaolana, region de SAVA. *Lemur News* 15: 54–59.

Goodman, S. M., M. J. Raherilalao, D. Rakotomalala, A. Raselimanana, H. Schütz and V. Soarimalala. 2003. Les lémuriens. In: *Nouveaux résultats d'inventaires biologiques faisant référence à l'altitude dans la région des massifs montagneux de Marojejy et d'Anjanaharibe-Sud*, S. M. Goodman and L. Wilmé (eds.), pp.279–286. Centre d'Information et de Documentation Scientifique et Technique, Antananarivo.

Gordon, A. D., S. E. Johnson and E. E. Louis Jr. 2013. Females are the ecological sex: sex-specific body mass ecogeography in wild sifaka populations (*Propithecus* spp.). *American Journal of Physical Anthropology* 151: 77–87.

Groves, C. P. 2001. *Primate Taxonomy*. Smithsonian Institution Press, Washington, DC.

Groves, C. P. and K. M. Helgen. 2007. Craniodental characters in the taxonomy of *Propithecus*. *International Journal of Primatology* 28: 1363–1383.

Jenkins, R. K. B., A. Keane, A. R. Rakotoarivelo, V. Rakotomboavonjy, F. H. Randrianandrianina, H. J. Razafimanahaka, S. R. Ralaiarimalala and J. P. G. Jones. 2011. Analysis of patterns of bushmeat consumption reveals extensive exploitation of protected species in eastern Madagascar. *PLoS ONE* 6: 1–12.

Lehman, S. M., M. I. Mayor and P. C. Wright. 2005. Ecogeographic size variations in sifakas: a test of the resource seasonality and resource quality hypotheses. *American Journal of Physical Anthropology* 126: 318–328.

Loudon, J. E., E. R. Patel, C. Faulkner, B. Schopler, R. Kamer and C. V. Williams. 2013. An ethnoprimatological assessment of human impact on the parasite ecology of silky sifaka (*Propithecus candidus*). *American Journal of Physical Anthropology* 56: 182. Abstract.

Mayor, M. I., J. A. Sommer, R. M. Huebinger, R. C. Barbe and E. E. Louis Jr. 2002. Characterization of seven microsatellite marker loci in a genus of Malagasy lemurs (*Propithecus*). *Molecular Ecology Notes* 2: 385–388.

Mayor, M. I., J. A. Sommer, M. L. Houck, J. R. Zaonarivelo, P. C. Wright, C. Ingram, S. R. Engel and E. E. Louis Jr. 2004. Specific status of *Propithecus* spp. *International Journal of Primatology* 25: 875–900.

Milne-Edwards, A. and A. Grandidier. 1875. *Histoire physique, naturelle et politique de Madagascar. Vol. VI. Histoire naturelle des mammifères*, Tome I, Texte I. Imprimerie Nationale, Paris.

Mittermeier, R. A., E. E. Louis Jr., M. Richardson, C. Schwitzer, O. Langrand, A. B. Rylands, F. Hawkins, S. Rajaobelina, J. Ratsimbazafy, R. Rasoloarison, C. Roos, P. M. Kappeler and J. Mackinnon. 2010. *Lemurs of Madagascar*. 3rd edition. Conservation International Tropical Field Guide Series, Arlington, VA.

Moore, J. and Patel, E.R. 2012. Lemur population surveys and remote sensing in Marojejy National Park. Prosimian Congress Conference Abstract. Valbio, Madagascar. Abstract.

Nielson, M. and E. R. Patel. 2008. The role of taste preference and wealth in bushmeat hunting in villages adjacent to Marojejy National Park, Madagascar. In: XXII Congress of the International Primatological Society, Primate Eye Special Issue, pp.222–223. Abstract.

Patel, E. R. 2005. Silky sifaka predation (*Propithecus candidus*) by a fossa (*Cryptoprocta ferox*). *Lemur News* 10: 25–27.

Patel, E. R. 2006a. Scent-marking in wild silky sifakas (*Propithecus candidus*) in Madagascar: sex differences and seasonal effects in usage and response across multiple scent-mark types. *International Journal of Primatology* 27: Abstract #496.

Patel, E. R. 2006b. Activity budget, ranging, and group size in silky sifakas (*Propithecus candidus*). *Lemur News* 11: 42–45.

Patel, E. R. 2007a. Logging of rare rosewood and palisandre (*Dalbergia* spp.) within Marojejy National Park, Madagascar. *Madagascar Conservation and Development* 2: 11–16.

Patel, E. R. 2007b. Non-maternal infant care in wild silky sifakas (*Propithecus candidus*). *Lemur News* 12: 39–42.

Patel, E. R. 2012a. Acoustic and Olfactory Communication in Eastern Sifakas (*Propithecus* sp.) and Rhesus Macaques (*Macaca mullata*). Doctoral dissertation, Cornell University, Ithaca, NY.

Patel, E. R. 2012b. Parasites found in wild silky sifakas. *Duke Lemur Center SAVA Conservation Newsletter* 2: 9–10.

Patel, E. R. 2013. Territorial scent marking in wild silky sifakas. *Duke Lemur Center SAVA Conservation Newsletter* 2: 12–14.

Patel, E. R. and L. H. Andrianandrasana. 2008. Low elevation silky sifakas (*Propithecus candidus*) in the Makira Conservation Site at Andaparaty-Rabeson: ranging, demography, and possible sympatry with red ruffed lemurs (*Varecia rubra*). *Lemur News* 13: 18–22.

Patel, E. R. and C. Girard-Buttoz. 2008. Non-nutritive tree gouging in wild Milne-Edwards' sifakas (*Propithecus edwardsi*): Description and potential communicative functions. *XXII Congress of the International Primatological Society, Primate Eye Special Issue*: Abstract #110.

Patel, E. R., J. J. Marshall and H. Parathian. 2005. Silky sifaka (*Propithecus candidus*) conservation education in northeastern Madagascar. *Laboratory Primate Newsletter* 44: 8–11.

Powzyk, J. A. 1997. The Socio-ecology of Two Sympatric Indrids: *Propithecus diadema* and *Indri indri*: A Comparison of Feeding Strategies and Their Possible Repercussions on Species-specific Behaviors. Doctoral Dissertation, Duke University, Durham, NC.

Rajaonarison, M. F., E. R. Patel and E. Razafimahatratra. 2012. Diet, ranging, and activity budget of wild silky sifakas in Makira. Prosimian Congress Conference Abstract. Valbio, Madagascar. Abstract.

Rasolofoson, D., G. Rakotondratsimba, O. Rakotonirainy, L. M. A. Rakotozafy, J. H. Ratsimbazafy, L. Rabetafika and R. M. Randrianarison. 2007. Influence of human pressure on lemur groups on the Makira Plateau, Maroantsetra, Madagascar. *Madagascar Conservation and Development* 2: 21–27.

Ratelolahy, F. J. and F. M. J. Raivoarisoa. 2007. Distribution et statut de population de Propithèque Soyeux (*Propithecus candidus*) dans la forêt de Makira, région d'Anjanaharibe, Nord Est de Madagascar. Report, Wildlife Conservation Society, Antananarivo.

Schilling, A. 1979. Olfactory communication in prosimians. In: *The Study of Prosimian Behavior*, G. A. Doyle and R. D. Martin (ed.), pp.461–542. Academic Press, New York.

Schmid, J. and R. Smolker. 1998. Lemurs of the Reserve Speciale d'Anjanaharibe-Sud, Madagascar. *Fieldiana Zoology* 90: 227–240.

Sterling, E. and K. McFadden. 2000. Rapid census of lemur populations in the Parc National de Marojejy, Madagascar. *Fieldiana Zoology* 97: 265–274.

Tattersall, I. 1982. *The Primates of Madagascar*. Columbia University Press, New York.

Tattersall, I. 2007. Madagascar's lemurs: cryptic diversity or taxonomic inflation? *Evolutionary Anthropology* 16: 12–23.

Wilmé, L. and M. W. Callmander. 2006. Relic populations of primates: sifakas. *Lemur News* 11: 24–31.

Indri

Indri indri (Gmelin, 1788) Madagascar (2012)

Christoph Schwitzer, Russell A. Mittermeier, Edward E. Louis Jr. & Matthew Richardson



Some color forms of Indri (Indri indri) (Illustrations: Stephen D. Nash)

The Indri occurs in northeastern and central eastern Madagascar, roughly from the Anosibe an'ala Classified Forest in the south to the Anjanaharibe-Sud Special Reserve in the north (Petter *et al.* 1977; Tattersall 1982; Powzyk and Thalmann 2003). There appears to be a regional trend regarding the amount of white and black fur on the coat, and as a consequence two distinct subspecies were formerly recognized. These are now believed to constitute a cline, with darker individuals to be found in the north of the species' range and lighter ones to the south.

With a head-body length of 64–72 cm and a body mass of 5.8–7.1 kg (and some individuals weighing up to 9 kg), the Indri is the largest of the living lemurs (Glander and Powzyk 1998; Powzyk 1997; E. E. Louis Jr., pers. obs.). In the Analamazaotra Special Reserve and the Anjozorobe-Angavo protected area, males are slightly larger than females, and there is some slight dimorphism in colour pattern. Whether these differences hold true in other parts of the range remains to be determined. The species inhabits primary and secondary moist lowland and montane forest, as well as some disturbed habitats,

from sea level to 1800 m (Goodman and Ganzhorn 2004a, 2004b). It is often found in mountainous habitats or steep terrain with numerous ridges and valleys. All levels of the canopy are used, although during October-December the animals tend to stay in the lower levels to avoid biting insects. Population densities typically range from 9 to 16 individuals/km², but are thought to be as low as 5.2 individuals/km² in some areas (Powzyk and Thalmann 2003; Glessner and Britt 2005). The indri is reported to reach quite high densities (22.9 individuals/km²) if not hunted by local people (Powzyk and Thalmann 2003).

Indris feed mainly on immature leaves, although fruits, seeds, flowers, buds and bark are also taken, the latter varying in proportion according to the season (Powzyk and Mowry 2007). Individuals descend to the ground to eat soil as well (Powzyk 1997; Britt *et al.* 2002; Powzyk and Thalmann 2003). The Indri has been studied in the forests of Analamazaotra (Pollock 1975a, 1975b, 1977, 1979a, 1979b) and in nearby Mantadia National Park (Powzyk 1996, 1997; Powzyk and Mowry 2003). There it lives in small groups of 2–6 individuals,

normally consisting of a monogamous adult pair and their offspring (Pollock 1979; Powzyk 1997). Although groups in fragmented habitat tend to be larger than those in more extensive, undisturbed areas (Pollock 1979a, 1979b; Powzyk 1997), this is not always the case (V. Sorrentino, pers. comm.). Changes in the composition of larger groups are quite frequent (V. Sorrentino, pers. comm.). Home ranges average 18 ha in the fragmented forests of Analamazaotra, but can be as large as 40 ha in the more pristine forests of Mantadia, where day ranges of 300–800 m are common. In the low altitude forest of Betampona, home ranges average 27 ha (Glessner and Britt 2005). A large central part of each ranging area constitutes a defended territory, from which other groups are excluded.

Mating takes place between December and March. The female produces a single young about every two or three years—a very slow reproductive rate for a prosimian. Births usually occur in May or June (but can be as late as August), the gestation period being between 135 and 145 days (range 130–150). Females do not reach full sexual maturity until 7–9 years of age (Pollock 1977).

The Indri is Critically Endangered (Andrainarivo et al. 2011) based on a predicted population reduction of ≥80% in the future over a 3-generation time period (36 years) due to a continuing and projected decline in area, extent and quality of habitat, in addition to continuing and projected exploitation through unsustainable hunting pressure. The principal threat to this species is habitat destruction for slash-and-burn agriculture, and logging and firewood gathering, even in protected areas. Contrary to what was believed in the past, illegal hunting is also a major problem for the Indri in certain areas (Jenkins et al. 2011). Although long thought to be protected by local "fady" (traditional taboos), these do not appear to be universal and the animals are now hunted even in places where such tribal taboos do exist. In many areas these taboos are breaking down with cultural erosion and immigration, and local people often find ways to circumvent taboos even if they are still in place. For example, a person for whom eating the indri is forbidden may still hunt the animals for sale to others, while those who may be forbidden to kill Indris can purchase them for food. Recent studies of villages in the Makira Forest indicate that Indris have also been hunted in the past for their skins (worn as clothing), that Indri meat is prized and fetches a premium price, and that current levels of hunting are unsustainable (Golden 2005, 2009; Jenkins et al.

2011; R. Dolch, pers. comm.). This species occurs in three national parks (Mananara-Nord, Mantadia, and Zahamena), two strict nature reserves (Betampona and Zahamena), and five special reserves (Ambatovaky, Analamazaotra, Anjanaharibe-Sud, Mangerivola, and Marotandrano) (Nicoll and Langrand 1989; Powzyk 1997; Schmid and Smolker 1998; Britt et al. 1999; CBSG 2002). It is found as well in the Anjozorobe-Angavo Protected Area and in the forests of Makira, which are currently under temporary government protection (though hunting pressure in the latter appears to be especially heavy). The corridor between Mantadia and Zahamena has been proposed as a new conservation site, and the Anosibe an'ala Classified Forest should be considered for the creation of a new park or reserve as well. No population figures are available, but a reasonable estimate would be 1,000-10,000. The Indri does not occur on the Masoala Peninsula or in Marojejy National Park, despite the latter area being connected to forest less than 40 km away where the species is present. Before wholesale deforestation occurred it was much more widely distributed, with a separate group said to occupy almost every ridge of the island's eastern forests. Subfossil evidence indicates that they once occurred well into the interior of Madagascar at least as far west as the Itasy Massif, southwest to Ampoza-Ankazoabo (Tattersall 1982; Godfrey et al. 1999) and north to the Ankarana Massif (Jungers et al. 1995).

The IUCN lemur conservation strategy 2013–2016 details conservation measures that will benefit the Indri in seven priority sites: Marojejy National Park and Anjanaharibe-Sud Special Reserve; Makira; Mananara Nord National Park; Ankeniheny-Zahamena Corridor (CAZ); Betampona Natural Reserve; Anjozorobe-Angavo and Tsinjoarivo (Schwitzer *et al.* 2013).

References

Andrainarivo, C., V. N. Andriaholinirina, A. T. C. Feistner, T. Felix, J. U. Ganzhorn, N. Garbutt, C. Golden, W. B. Konstant, E. E. Louis Jr., D. M. Meyers, R. A. Mittermeier, A. Perieras, F. Princee, J. C. Rabarivola, B. Rakotosamimanana, Rasamimanana, H., J. Ratsimbazafy, G. Raveloarinoro, A. Razafimanantsoa, Y. Rumpler, C. Schwitzer, R. Sussman, U. Thalmann, L. Wilmé and P. C. Wright. 2011. *Indri indri*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. www.iucnredlist.org. Accessed 16 March 2014.

Britt, A., A. Axel and R. Young. 1999. Brief surveys of two classified forests in Toamasina Province, eastern Madagascar. *Lemur News* 4: 25–27.

Britt, A., N. J. Randriamandratonirina, K. D. Glasscock and B. R. Iambana. 2002. Diet and feeding behaviour of *Indri indri* in a low-altitude rain forest. *Folia Primatologica* 73: 225–239.

Conservation Breeding Specialist Group (CBSG). 2002. Evaluation et plans de gestion pour la conservation (CAMP) de la faune de Madagascar: Lémuriens, autres mammifères, reptiles et amphibiens, poissons d'eau douce et évaluation de la viabilité des populations et des habitats (PHVA) de *Hypogeomys antimena* (Vositse). Mantasoa, Madagascar 20–25 Mai 2001. Version finale Juillet 2002. Apple Valley, MN.

Glander, K. E. and J. A. Powzyk. 1998. Morphometrics of wild *Indri indri* and *Propithecus diadema diadema*. *Folia Primatologica* 69 (Suppl): 399. Abstract.

Glessner, K. D. and A. Britt. 2005. Population density and home range size of *Indri indri* in a protected low altitude rain forest. *International Journal of Primatology* 26: 855–872.

Godfrey, L. R., W. L. Jungers, E. L. Simons, P. S. Chatrath and B. Rakotosamimanana. 1999. Past and present distributions of lemurs in Madagascar. In: *New Directions in Lemur Studies*, B. Rakotosamimanana *et al.* (eds.), pp.19–53. Springer, New York.

Golden, C. D. 2005. Eaten to endangerment: Mammal hunting and the bushmeat trade in Madagascar's Makira Forest. Undergraduate thesis, Harvard University, Cambridge, MA.

Golden, C. D. 2009. Bushmeat hunting and use in the Makira Forest north-eastern Madagascar: a conservation and livelihoods issue. *Oryx* 43: 386–392.

Goodman, S. M. and J. U. Ganzhorn. 2004a. Elevational ranges of lemurs in the humid forests of Madagascar. *International Journal of Primatology* 25: 331–350.

Goodman, S. M. and J. U. Ganzhorn. 2004b. Biogeography of lemurs in the humid forests of Madagascar: the role of elevational distribution and rivers. *Journal of Biogeography* 31: 47–55.

Jenkins, R., A. Keane, A. Rakotoarivelo, V. Rakotomboavonjy, F. Randrianandrianina, H. Razafimanahaka, S. Ralaiarimalala and J. Jones. 2011. Analysis of patterns of bushmeat consumption reveals extensive exploitation of protected species in eastern Madagascar. *PLoS ONE* 6: e27570.

Jungers, W. L., L. R. Godfrey, E. L. Simons and P. S. Chatrath. 1995. Subfossil *Indri indri* from the Ankarana Massif of northern Madagascar. *American Journal of Physical Anthropology* 97: 357–366.

Nicoll, M. E. and O. Langrand. 1989. Madagascar: *Revue de la Conservation et des Aires Protégées*. WWF-Fonds Mondial pour la Nature, Madagascar.

Petter, J. J., R. Albignac and Y. Rumpler. 1977. Mammifères, Lémuriens (Primates, Prosimiens). *Faune de Madagascar* 44: 1–513. Orstom CNRS, Paris.

Pollock, J. I. 1975a. The Social Behaviour and Ecology of *Indri indri*. Doctoral Dissertation, University College, London.

Pollock, J. I. 1975b. Field observations on *Indri indri*: a preliminary report. In: *Lemur Biology*, I. Tattersall and R. W. Sussman (eds.), pp.287–311. Plenum Press, New York.

Pollock, J. I. 1977. The ecology and sociology of feeding in *Indri indri*. In: *Primate Ecology: Studies of Feeding and Ranging Behaviour in Lemurs, Monkeys and Apes*, T. H. Clutton-Brock (ed.), pp.37–69. Academic Press, London.

Pollock, J. I. 1979a. Female dominance in *Indri indri*. *Folia Primatologica* 31: 143–164.

Pollock, J. J. 1979b. Spatial distribution and ranging behavior in lemurs. In: *The Study of Prosimian Behavior*, G. A. Doyle and R. D. Martin (eds.), pp.359–409. Academic Press, London.

Powzyk, J. A. 1996. A comparison of feeding strategies between the sympatric *Indri indri* and *Propithecus diadema diadema* in primary rain forest. *American Journal of Physical Anthropology* 22 (suppl.): 190.

Powzyk, J. A. 1997. The socio-ecology of two sympatric indrids, *Propithecus diadema diadema* and *Indri indri*: a comparison of feeding strategies and their possible

repercussions on species-specific behaviors. Doctoral thesis, Duke University, Durham, NC.

Powzyk, J. A. and C. B. Mowry. 2003. Dietary and feeding differences between sympatric *Propithecus diadema diadema* and *Indri indri*. *International Journal of Primatology* 24: 1143–1162.

Powzyk, J. A. and C. B. Mowry. 2007. The feeding ecology and related adaptations of *Indri indri*. In: *Lemurs*, L. Gould and M. L. Sauther (eds.), pp.353–368. Springer, New York.

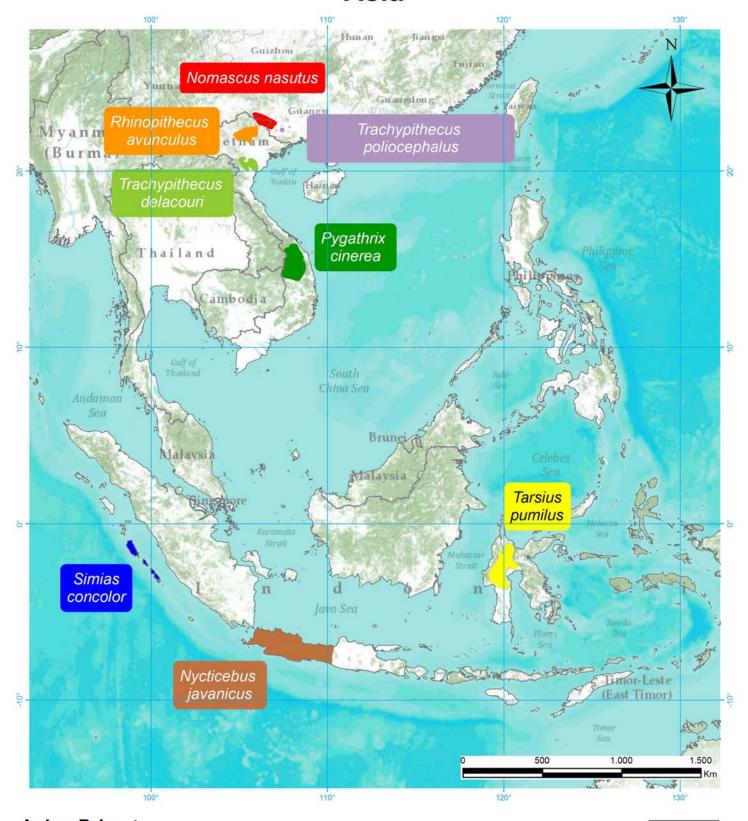
Powzyk, J. and U. Thalmann. 2003. *Indri indri*. In: *The Natural History of Madagascar*, S. M. Goodman and J. P. Benstead (eds.), pp.1342–1345. University of Chicago Press, Chicago, IL.

Schmid, J. and R. Smolker. 1998. Lemurs in the reserve special d'Anjanaharibe-Sud, Madagascar: with reference to elevational variation. *Fieldiana Zoology* 90: 227–240.

Schwitzer, C., R. A. Mittermeier, N. Davies, S. Johnson, J. Ratsimbazafy, J. Razafindramanana, E. E. Louis Jr. and S. Rajaobelina (eds). 2013. *Lemurs of Madagascar: A Strategy for Their Conservation 2013–2016*. IUCN SSC Primate Specialist Group, Bristol Conservation and Science Foundation, and Conservation International, Bristol, UK. 185pp.

Tattersall, I. 1982. *The Primates of Madagascar*. Columbia University Press, New York.

Asia



Asian Primates

Nomascus nasutus
Simias concolor
Nycticebus javanicus
Tarsius pumilus
Pygathrix cinerea
Trachypithecus delacouri
Rhinopithecus avunculus
Trachypithecus poliocephalus



Pygmy Tarsier

Tarsius pumilus Miller and Hollister 1921 Central Sulawesi, Indonesia (2012)

Nanda B. Grow & Sharon Gursky-Doyen



Pygmy Tarsier (Tarsius pumilus) (Illustration: Stephen D. Nash)

The Pygmy tarsier, *Tarsius pumilus*, was rediscovered in 2008 by Gursky and Grow (2009). It had not been observed for more than 90 years, and was even speculated to be extinct. Pygmy tarsiers are a high-altitude tarsier species endemic to the mountains of central Sulawesi, Indonesia. The species is distinguished from lowland tarsiers by its high-altitude habitat, extremely small size, and unusual behaviours (Shekelle 2008; Grow and Gursky-Doyen 2010). In particular, Pygmy tarsiers weigh an average of 55 g, while lowland tarsier species weigh 108–136 g (Grow 2013a). Further, unlike lowland Sulawesian tarsiers, Pygmy tarsiers have not been observed to produce scent marks or audible duet calls (Grow and Gursky-Doyen 2010), rendering the species extremely cryptic and difficult to locate.

Named by Miller and Hollister in 1921, Pygmy tarsiers were previously known only from two museum specimens. In 1917, the holotype (USNM 219454) was collected by H. C. Raven at 1800 m on Mt. Rano Rano (1°30'S, 120°28'E). The species was first described based on this specimen. Musser and Dagosto (1987) confirmed the species status of T. pumilus based on the holotype, and found that G. Heinrich had independently collected an adult female T. pumilus specimen at 2200 m on Mt. Latimojong (AMNH 196477; 3°30'S, 120°05'E) in the northern part of the southwestern peninsula of Sulawesi. In the intervening decades, there were numerous attempts to locate a living population of this elusive species, but all were unsuccessful until Gursky and Grow's surveys in 2008, at 2100 m on Mt. Rore Katimbu (01°16.8'S, 120°18.5'E) in Lore Lindu National Park, central Sulawesi (Grow and Gursky-Doyen 2010). These surveys concentrated efforts on the same mountain where a small mammal survey accidentally collected a third specimen in 2000, indicating the species still existed in the wild (Maryanto and Yani 2004).

All of the myriad threats affecting this species are compounded by the extremely limited distribution of this species. Pygmy tarsiers have only been observed at elevations above 2000 m at Mt. Rore Katimbo, despite surveys at lower altitudes (Grow 2013b). Gursky (unpubl. data 2009) conducted surveys at Latimojong in south Sulawesi but did not observe any Pygmy tarsiers. Surveys by Grow in 2010 and 2012 determined that the population density of *T. pumilus* at Rore Katimbo is approximately 92 individuals per 100 ha, with an estimated 6 groups per 100 ha (Grow *et al.* 2013). The amount of available habitat for Pygmy tarsiers is severely constrained; in Lore Lindu National Park, only 20% of the park consists of elevations higher than 1500 m, and high-altitude Pygmy tarsiers will occupy only a fraction of that area.

Human advancement into higher altitudes is a major threat given the limited distribution and low population densities of Pygmy tarsiers. Although the tarsiers are located in a protected area, there is tremendous deforestation and illegal encroachment of villages into Lore Lindu National Park. Residents of villages located in and near the park continue to extract resources and modify the landscape. Resources regularly collected from the protected forest include resin from Dipterocarpaceae trees, birds and mammals for the wildlife trade, and lumber (N. Grow, pers. obs.). The villages in Lore Lindu National Park are also undergoing massive population growth, creating a higher demand for agricultural land and firewood, increasingly at high altitudes in montane forest where resources are still available. As human populations expand in the future, Pygmy tarsier habitat is at risk of further clearing for agricultural usage. The IUCN Red List notes that this extremely limited population is currently decreasing, and the species is classified as Data Deficient (Shekelle and Salim 2008).

References

Grow, N. 2013a. Altitudinal Effects on The Behavior and Morphology of Pygmy Tarsiers (*Tarsius pumilus*) in Central Sulawesi, Indonesia. Doctoral dissertation, Texas A&M University, College Station, TX.

Grow, N. 2013b. Altitudinal distribution and ranging patterns of pygmy tarsiers (*Tarsius pumilus*). In: *High Altitude Primates*, N. B. Grow, S. Gursky-Doyen and A. Krzton (eds.), pp.43–59. Springer, New York.

Grow, N. and S. Gursky-Doyen. 2010. Preliminary data on the behavior, ecology, and morphology of pygmy tarsiers (Tarsius pumilus). International Journal of Primatology 31: 1174–1191.

Grow, N., S. Gursky and Y. Duma. 2013. Altitude and forest edges influence the density and distribution of pygmy tarsiers (*Tarsius pumilus*). *American Journal of Primatology* 75: 464–477.

Gursky-Doyen, S. and N. Grow. 2009. Elusive highland pygmy tarsier rediscovered in Sulawesi, Indonesia. *Oryx* 43: 173–174.

Maryanto, I. and M. Yani. 2004. The third record of pygmy tarsier (*Tarsius pumilus*) from Lore Lindu National Park, central Sulawesi, Indonesia. *Tropical Biodiversity* 8: 79–85.

Miller, G. S. and N. Hollister. 1921. Twenty new mammals collected by H. C. Raven in Celebes. *Proceedings of the Biological Society of Washington* 34: 93–104.

Musser, G. and M. Dagosto. 1987. The identity of *Tarsius pumilus*, a pygmy species endemic to the montane mossy forests of Central Sulawesi. *American Museum Novitates* (2867): 1–53.

Shekelle, M. 2008. The history and mystery of the mountain tarsier, *Tarsius pumilus*. *Primate Conservation* (23): 121–124.

Shekelle, M. and A. Salim. 2008. *Tarsius pumilus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 18 March 2014.

Javan Slow Loris

Nycticebus javanicus É. Geoffroy Saint-Hilaire, 1812 Indonesia (2008, 2010, 2012)

K. Anna I. Nekaris, E. Johanna Rode & Vincent Nijman

All Asian lorises are imperilled by the devastating loss of their habitat; indeed, this major threat resulted in Sri Lanka's Critically Endangered Horton Plains slender loris appearing rightfully in the last two incarnations of this list (Nekaris 2006; Nekaris and Perera 2007). An even greater immediate threat to Asian lorises, however, is their high demand in the rampant Asian pet and traditional medicine trades and their use as tourist photo props (Schulze and Groves 2004; Streicher 2004). Easy to catch due to their slow locomotion, numbers of lorises in animal markets far outstretch the ability of these slow-reproducing primates to recover their population numbers in the wild. Indeed, this threat raised international concern, resulting in the transfer of all members of the genus *Nycticebus* to CITES Appendix I in 2007 (Nekaris and Nijman 2007). Eight species of slow loris are now recognized: N. coucang (greater), N. pygmaeus (pygmy), N. bengalensis (Bengal), N. javanicus (Javan) and four Bornean: N. menagensis, N. bancanus, N. borneanus, and N. kayan (Roos 2003; Chen et al. 2007; Munds et al. 2013). All slow lorises suffer from trade throughout their range, but when combined with tremendous habitat loss, no other species has been harder hit than the Javan slow loris. There is a general public perception that slow lorises are 'cute' as demonstrated by a recent trend for videos of these animals being used as pets on You Tube and other social media outlets, which exacerbates their demand in the pet trade (Nekaris et al. 2013b).

Recognized by the IUCN as a species in 2006, and currently listed as Critically Endangered (Nekaris *et al.* 2013a), the Javan slow loris is distinguished easily from its congeners in several respects. Both morphologically and genetically, it is most similar to, yet still distinct from, the largest slow loris, *N. bengalensis* of mainland Asia (Roos 2003; Groves and Maryanto 2008). Weighing about 1 kg, the most distinctive feature of the Javan slow loris is its facial mask, comprised of bold fork marks leading from the eyes and ears to the crown of the head, revealing a white diamond pattern on the forehead



Javan slow loris (*Nycticebus javanicus*) (Illustration: Stephen D. Nash)

(Nekaris and Jaffe 2007). Despite being legally protected since 1973, with its creamy neck, bold dorsal stripe, and panda-like face, it is no wonder that Indonesian pet traders in the 1990s targeted Javan slow lorises above other endemic loris species. Since 2002, however, the numbers of Javan lorises in trade have decreased,

with a stark rise in numbers of Sumatran greater slow lorises (*N. coucang*), a species whose threat status must also be carefully monitored. Indeed, over one year of market surveys on Java in 2013, quadruple the number of Sumatran than Javan slow lorises were counted, with traders claiming that Javan slow lorises could no longer be found. In November 2013 alone, nearly 300 Sumatran lorises were confiscated in two raids. The smaller raid, yielding 76 individuals, was followed by the almost immediate death toll of 31 individuals, and by 12 infant lorises being born. All of these animals were confiscated before ever making it to markets, but show the dramatic extent of this trade.

Nycticebus javanicus is found only on the Indonesian island of Java. Java has a long history of cultivation and deforestation that already started c.1000 AD, but took off in 1830 when the Dutch colonial government imposed the so-called "cultuurstelsel". To support this agro-economic system, farmers were forced to grow export crops on communal grounds, which were often forest (Whitten et al. 1996). By the end of the 19th century the natural forest was severely fragmented, and at the beginning of the last century the remaining forest, especially in West and Central Java, showed a fragmentation pattern very similar to that seen today. Over the last few decades, the decrease in forest area has been slow. At present, less than 10% of the original forest remains, most of it covering the higher slopes of the central mountains.

GIS models made available by Thorn et al. in 2009 suggested that historic forest loss and continued degradation mean that less than 20% of habitat suitable for N. javanicus remains and that only 17% of the potential distribution of *N. javanicus* is currently within the protected area network of Java. Based largely on Thorn et al.'s recommendations, Voskamp et al. (2014) investigated eight of these areas along with an additional six unprotected areas. Their results concurred with those conducted by three separate research groups, with animals occurring at 0.02 to 0.20 ind./km², when they could be found at all; this means that 5-10 km must be walked to see a single loris (Nekaris and Nijman 2008; Winarti 2008). Roads and human disturbance have been shown to correlate negatively with Javan slow loris abundance (Collins 2007; Winarti 2008). Surprisingly however, during Voskamp et al.'s study, numbers of lorises were higher in agro-forest that is, in some cases, extremely disturbed by humans.

Also urgently required are programmes to mitigate trade in all species of slow loris. A number of studies have found that slow lorises are not always a targeted group, but that they do have economic value throughout their range. Rather than seeking a loris, villagers moving through the forest simply pick up a loris when they happen to see it (Starr et al. 2008). Similarly, when forests are clear cut (for agriculture or cash crops), villagers pick through the felled trees and collect the lorises; with a defence mechanism to cling to branches rather than to flee, and with their nocturnal senses stunned by bright daylight, lorises are an easy target (Ratjacsek 1998). Nijman and Nekaris (2014) showed that traditional beliefs about slow lorises may hinder people from hunting them, particularly beliefs regarding their being venomous or poisonous.

In Java itself, lorises are often specifically targeted for the trade (K. L. Sanchez, pers. obs.). Local villagers who find a loris take it to a distributor dealer who compiles a stock of lorises. These animals go to middlemen who then distribute them throughout the "bird" markets in the main towns in Java. The traders who ultimately sell the animals are aware that trading lorises is profitable, reaching a price in the market up to ten times or more the purchasing price at the stocker's level.

Once they arrive at a market, lorises face other threats. To avoid being bitten by slow lorises, which are one of the few venomous mammals, traders habitually cut or pull out an animal's lower front teeth. Most of these lorises die due to general infection, dental abscess or pneumonia. Those that do survive are no longer able to eat their preferred food (gum) (Wiens et al. 2006), or engage in the important behaviour of social grooming with the toothcomb, meaning that any confiscated animals are unlikely to survive if released to the wild. Reintroduction itself is a threat to the Javan loris; three major trade hubs, markets in Jakarta, Bandar Lampung and Palembang, receive lorises from throughout the region. The similar appearance of lorises to the untrained eye results in release of other loris species into Java, with potential for disastrous effects from hybridization or displacement by invasive species.

Only a single study has assessed the success of reintroduction of Javan slow lorises, finding up to a 90% death rate (Moore 2009). Illness, hypothermia and exhaustion were all implicated in the death of the lorises. Sadly, reintroductions were started before

anything was known about their behaviour, ecology or wild distribution. No habitat assessment could be made since it was not even known in what type of habitat the species occurred.

In 2011, the first long-term study of Javan slow loris behavioural ecology was instigated by the Little Fireface Project in Garut District, West Java, Indonesia (Nekaris et al. 2013b; Rode et al. 2014). This multi-disciplinary project has obtained the first data about slow loris behaviour in an agro-forest matrix, including home range size, social organization, infant dispersal, and feeding ecology. Some notable discoveries have been that both sexes disperse from their natal range at about 18 months old, dispersal distances are some 1-2 km from the natal range, home range sizes are large (5-10 ha), and the diet of lorises comprises mainly gum, supplemented with nectar and insects. Several initiatives have been put into place to conserve slow lorises in the area and in Java. National workshops have been held for law enforcement officers and rescue center employees to feed essential data into a national loris action plan. At the local level, lorises are totally dependent on local people for their protection, feeding on human planted tree species and residing in human farmlands. Thus a major conservation program, combining empowerment activities, conservation education and village events, has been launched, and it is hoped that it can be used as a model for other key loris sites in Indonesia.

For a long time, slow lorises were thought to be common throughout Indonesia, and the presence of animals in trade was believed to be an indicator of their abundance. We are only beginning to unravel the complexity of their taxonomy and distribution, leading to a bleak picture overall. While Java has an impressive and comprehensive protected area network, encompassing over 120 terrestrial conservation areas covering some 5,000 km², enforcement of environmental laws and active protection of forest is lacking in most of these parks. Besides curbing the illegal trade, it is paramount that these conservation areas, and indeed all other remaining forest areas on the island, be effectively protected.

References

Chen, J. H., D. Pan, C. P. Groves, Y. X. Wang, E. Narushima, H. Fitch-Snyder, P. Crow, V. N. Thanh, O. Ryder, H. W. Zhang, Y. X. Fu and Y. P. Zhang.

2006. Molecular phylogeny of *Nycticebus* inferred from mitochondrial genes. *International Journal of Primatology* 27: 1187–1200.

Collins, R. L. 2007. Behavioural Data of Captive Greater Slow Loris (*Nycticebus coucang*) and Javan Slow Loris (*N. javanicus*), and a Survey of Javan Slow Loris in Mt. Salak, West Java, Java. MSc dissertation, Oxford Brookes University, Oxford.

Groves, C. P. and I. Maryanto. 2008. Craniometry of slow lorises (genus *Nycticebus*) of insular Southeast Asia. In: *Primates of the Oriental Night*, M. Shekelle, C. P. Groves, I. Maryanto, H. Schulze and H. Fitch-Snyder (eds.), pp.115–122. Research Center for Biology, Indonesian Institute of Sciences and the Indonesian Biological Society, Bogor, Indonesia.

Moore, R. S. 2009. Ethics, Ecology and Evolution of Indonesian Slow Lorises (*Nycticebus* spp.) Rescued from the Pet Trade. PhD Thesis, Oxford Brookes University, Oxford.

Munds, R. A., K. A. I. Nekaris and S. M. Ford. 2013. Taxonomy of the Bornean slow loris, with new species *Nycticebus kayan* (Primates, Lorisidae). *American Journal of Primatology* 75: 46–56.

Nekaris, K. A. I. 2006. Horton Plains slender loris, Ceylon mountain slender loris *Loris tardigradus nycticeboides* Hill, 1942). In: *Primates in peril: the world's 25 most endangered primates 2004–2006*, R. A. Mittermeier, C. Valladares-Pádua, A. B. Rylands, A. A. Eudey, T. M. Butynski, J. U. Ganzhorn, R. Kormos, J. M. Aguiar and S. Walker (eds.), pp.10–11, 23. *Primate Conservation* (20): 1–28.

Nekaris, K. A. I. and S. Jaffe. 2007. Unexpected diversity within the Javan slow loris trade: implications for slow loris taxonomy. *Contributions to Zoology* 76: 187–196.

Nekaris, K. A. I. and N. Campbell. 2012. Media attention promotes conservation of threatened Asian slow lorises. *Oryx* 46: 169–170.

Nekaris, K. A. I. and V. Nijman. 2007. CITES proposal highlights rarity of Asian nocturnal primates (Lorisidae: *Nycticebus*). *Folia Primatologica* 78: 211–214.

Nekaris, K. A. I. and V. Nijman. 2008. Survey on the abundance and conservation of Sumatran slow lorises

(*Nycticebus coucang hilleri*) in Aceh, Northern Sumatra. *Folia Primatologica* 79: 365. Abstract.

Nekaris, K. A. I. and V. Perera. 2007. Horton Plains slender loris, Ceylon mountain slender loris *Loris tardigradus nycticeboides*. In: *Primates in Peril: the World's 25 Most Endangered Primates 2006–2008*, R. A. Mittermeier *et al.* (eds.), pp.12–13, 27. *Primate Conservation* (22): 1–40.

Nekaris, K. A. I., G. V. Blackham and V. Nijman. 2008. Implications of low encounter rates in five nocturnal species (*Nycticebus* spp). *Biodiversity and Conservation* 17: 733–747.

Nekaris, K. A. I., M. Shekelle, Wirdateti, E. J. Rode and V. Nijman. 2013a. *Nycticebus javanicus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 17 March 2014.

Nekaris, K. A. I., R. S. Moore, E. J. Rode and B. G. Fry. 2013b. Mad, bad, and dangerous to know: the only venomous primates, slow lorises. *BMC Journal of Venomous Animals and Toxins including Tropical Diseases*. 19: 21 (27 September 2013).

Nijman, V. and K. A. I. Nekaris. 2014. Traditions, taboos and trade in slow lorises in Sundanese communities in southern Java, Indonesia. *Endangered Species Research*. In press.

Ratajszczak, R. 1998. Taxonomy, distribution and status of the lesser slow loris *Nycticebus pygmaeus* and their implications for captive management. *Folia Primatologica* 69: 171–174.

Rode, E. J., V. Nijman, Wirdateti and K. A. I Nekaris. 2014. Ethology of the Critically Endangered Javan slow loris (*Nycticebus javanicus*) in West Java. *Asian Primates Journal*. In press.

Roos, C. 2003. Molekulare Phylogenie der Halbaffen, Schlankaffen, und Gibbons. Doctoral thesis, Technische Universität München, München, Germany.

Schulze, H. and C. P. Groves. 2004. Asian lorises: taxonomic problems caused by illegal trade. In: *Conservation of Primates in Vietnam*, T. Nadler, U. Streicher and Ha Thang Long (eds.), pp.33–36. Frankfurt Zoological Society, Frankfurt.

Starr, C. R., U. Streicher and K. A. I. Nekaris. 2008. The distribution and conservation of the pygmy loris (*Nycticebus pygmaeus*) in eastern Cambodia. *XXII Congress of the International Primatological Society, Edinburgh, UK. Primate Eye* 96: 116. Abstract.

Streicher, U. 2004. Aspects of the Ecology and Conservation of the Pygmy Loris *Nycticebus pygmaeus* in Vietnam. Dissertation, Ludwig-Maximilians Universität, München, Germany.

Thorn, J. S., V. Nijman, D. Smith and K. A. I. Nekaris. 2009. Ecological niche modelling as a technique for assessing threats and setting conservation priorities for Asian slow lorises (Primates: *Nycticebus*). *Diversity and Distributions* 15: 289–298.

Voskamp, A., E. J. Rode, C. N. Z. Coudrat, Wirdateti, A. Nawanto, R. Wilson and K. A. I. Nekaris. 2014. Habitat preferences and distribution of the Critically Endangered Javan slow loris (*Nycticebus javanicus*). *Endangered Species Research*. In press.

Whitten, A. J., R. E. Soeriaatmadja and S. A. Afiff. 1996. *The Ecology of Java and Bali*. The Ecology of Indonesia Series, Vol. II. Periplus Editions, Singapore.

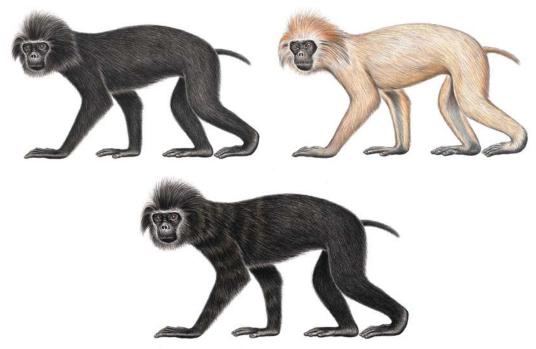
Wiens, F, A. Zitzmann and N. A. Hussein. 2006. Fast food for slow lorises: is low metabolism related to secondary compounds in high-energy plant diet? *Journal of Mammalogy* 87: 790–798.

Winarti, I. 2008. Field Research on Javan Slow Loris' Population in Sukakerta Ciamis and Kawungsari Tasikmalaya, West Java, Indonesia. Report, International Animal Rescue Indonesia (IARI), Ciapus, Bogor, Indonesia.

Pig-tailed Snub-nose Langur

Simias concolor Miller, 1903 Indonesia (2002, 2004, 2006, 2008, 2010, 2012)

Lisa M. Paciulli, Jatna Supriatna & Christian Roos



The two color forms of the Pig-tailed snub-nose langur (Simias concolor concolor) above, and the Siberut subspecies (Simias concolor siberu), below (Illustrations: Stephen D. Nash)

The Pig-tailed snub-nose langur (Simias concolor) is again serving as the flagship species for the six Mentawai Island primates. The other three species inhabiting the 7,000-km² archipelago located west of Sumatra are Kloss's gibbon (Hylobates klossii), the Pagai langur (Presybtis potenziani), the Siberut langur (P. siberu), the Pagai macaque (Macaca pagensis), and the Siberut macaque (M. siberu). Simias is a monotypic genus with two subspecies: Simias concolor concolor Miller, 1903 inhabits Sipora, North Pagai, and South Pagai islands and several small islets off of South Pagai; Simias c. siberu Chasen and Kloss, 1927 is restricted to Siberut Island (Zinner et al. 2013).

The first activity budget of habituated pig-tailed langurs described the activities of two groups living in the Betumonga region of southwestern North Pagai. The data show that they spend almost equal amounts of time resting (46%) and feeding (44%), and less time moving (7%) (Paciulli and Holmes 2008). New estimates of the amount of forest cover remaining on the Pagai Islands (about 826 km²) have been calculated using Google Earth Pro composite satellite imagery

(Paciulli and Viola 2009). The forest cover coupled with primate density data (Paciulli 2004) indicate that there are approximately 3,347 pig-tailed langurs, 1,049 Kloss's gibbons, 1,545 Pagai langurs, and 7,984 Pagai macaques on the Pagai Islands. All of the primate species seem to reach their highest known densities in the Peleonan Forest, site of the Siberut Conservation Project in northern Siberut (Waltert *et al.* 2008).

Simias concolor is classified as Critically Endangered on the IUCN Red List (Whittaker and Mittermeier 2008), threatened mainly by heavy hunting and commercial logging (Whittaker 2006). The Pagai island populations are threatened by forest conversion to oil palm plantations, and forest clearing and product extraction by local people (Whittaker 2006). Although hunting appears to be declining and opportunistic in many areas of the Pagais, where it still occurs it has devastating effects on *S. concolor*, the preferred game species (Mitchell and Tilson 1986; Fuentes 2002; Paciulli 2004). Tenaza (1987) estimated that twice as many individuals are killed by hunters each year as are born in the Pagai Islands. In a multi-population study, Erb *et al.* (2012) found that

hunting pressure reduced group size, resulting in the formation of male-female pairs, which is atypical for Asian colobines, which normally form small one-male groups with around five females.

The uncertainty of Indonesian government land-use means that land function and thus protection level on the Mentawai islands can change at any time with little notice, putting the species at further risk. There is only one main protected area for S. concolor: the 190,500-ha Siberut National Park, a UNESCO Biosphere Reserve, covers 47% of Siberut Island and serves as the main reserve for the Mentawai primates. The large majority of the other remaining natural habitat lies outside of officially protected areas. Simias concolor seems to be particularly sensitive to logging, having 5 individuals/ km² in unlogged Pagai forests to half that amount (2.5 individuals/km²) in Pagai forest patches logged 20 years earlier (Paciulli 2004). Drastic measures need to be taken to ensure that the Peleonan Forest on Siberut and areas on the Pagais are truly protected.

Whittaker (2006) suggested the following conservation actions for *S. concolor*: 1) increased protection for Siberut National Park, which currently lacks enforcement; 2) formal protection of the Peleonan forest in North Siberut, which is home to unusually high primate populations and is easily accessible; 3) protection of areas in the Pagai Islands by cooperating with a logging corporation that has practiced sustainable logging technique there since 1971; 4) conservation education, especially regarding hunting; and 5) the development of alternative economic models for the local people to reduce the likelihood of selling off their lands to logging companies.

References

Chasen, F. N. and C. B. Kloss. 1927. Spolia Mentawiensia-Mammals. *Proceedings of the Zoological Society of London* 53: 797–840.

Erb, W. M., C. Borries, N. S. Lestari and T. Ziegler. 2012. Demography of simakobu (*Simias concolor*) and the impact of human disturbance. *American Journal of Primatology* 74: 580–590.

Fuentes, A. 2002. Monkeys, humans and politics in the Mentawai Islands: no simple solutions in a complex world. In: *Primates Face to Face: Conservation Implications of Human-Nonhuman Primate Interactions*,

A. Fuentes and L. D. Wolfe (eds.), pp.187–207. Cambridge University Press, Cambridge, UK.

Miller, G. S. 1903. Seventy new Malayan mammals. *Smithsonionan Miscellaneous Collections* 45: 1–73.

Mitchell, A. H. and R. L. Tilson. 1986. Restoring the balance: traditional hunting and primate conservation in the Mentawai Islands, Indonesia. In: *Primate Ecology and Conservation*, J. G. Else and P. C. Lee (eds.), pp.249–260. Cambridge University Press, Cambridge, UK.

Paciulli, L. M. 2004. The Effects of Logging, Hunting, and Vegetation on the Densities of the Pagai, Mentawai Island Primates. Doctoral dissertation, State University of New York at Stony Brook, Stony Brook, NY.

Paciulli, L. M. and S. Holmes. 2008. Activity budget of simakobu monkeys (*Simias concolor*) inhabiting the Mentawai Islands, Indonesia. XXII Congress of the International Primatological Society, Edinburgh, UK, 3–8 August 2008. *Primate Eye* 96 Special Issue: 304. Abstract.

Paciulli, L. M. and J. Viola. 2009. Population estimates of Mentawai primates on the Pagai Islands, Mentawai, West Sumatra, Indonesia. *American Journal of Physical Anthropology* Suppl. 43. In press. Abstract.

Tenaza, R. 1987. The status of primates and their habitats in the Pagai Islands, Indonesia. *Primate Conservation* (8): 104–110.

Waltert, M., C. Abegg, T. Ziegler, S. Hadi, D. Priata and K. Hodges. 2008. Abundance and community structure of Mentawai primates in the Peleonan Forest, North Siberut, Indonesia. *Oryx* 42: 375–379.

Whittaker, D. J. 2006. A conservation action plan for the Mentawai primates. *Primate Conservation* (20): 95–105.

Whittaker, D. J. and R. A. Mittermeier. 2008. *Simias concolor.* In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 18 March 2014.

Zinner, D., G. H. Fickenscher and C. Roos. 2013. Family Cercopithecidae (Old World monkeys). In: *Handbook of the Mammals of the World, Volume 3 Primates*, R. A. Mittermeier, A. B. Rylands and D. E. Wilson (eds.), pp.550–627. Lynx Edicions, Barcelona.

Delacour's Langur

Trachypithecus delacouri (Osgood, 1932) Vietnam (2000, 2002, 2004, 2006, 2008, 2010, 2012)

Tilo Nadler



Delacour's langur is endemic to Vietnam, occurring in a very restricted area in the north of the country that comprises about 5,000 km² between 20°–20.30°N and 104.30°–106°E. The distribution is closely related to the limestone mountain ranges in the provinces Ninh Binh, Thanh Hoa, Hoa Binh and Ha Nam. The area of occurrence comprises only about 400 km² (Nadler 1996, 2004).

During the decades following the discovery of Delacour's langur in 1930 there was only scanty information on its existence and distribution. In 1987, the first sightings of live animals were reported from Cuc Phuong National Park (Ratajszczak *et al.* 1990). Intensive surveys by the Frankfurt Zoological Society in the decade before 2000 confirmed 18 isolated populations with a total of 280 to 320 individuals. Five localities were found where local people reported that this species had been extirpated. *Trachypithecus delacouri* is Critically Endangered (Nadler *et al.* 2008), and the most important factor in the decline in numbers is poaching, which is not primarily for meat, but for bones, organs and tissues that are used in the preparation of traditional medicines. The recorded numbers of animals hunted over 10 years

(1990–1999) totalled 320, an annual loss of more than 30 individuals, but the real number is undoubtedly higher (Nadler 2004; Nadler *et al.* 2003).

Surveys in 2004 in two protected areas with important subpopulations—Cuc Phuong National Park and Pu Luong Nature Reserve—showed a decline in numbers of 20% in five years (2000 to 2004) (Luong Van Hao and Le Trong Dat 2008). The population in Ngoc Son Nature Reserve was extirpated (Le Trong Dat et al. 2008). Monitoring of populations and surveys carried out by the Endangered Primate Rescue Center (EPRC) shows a continuous and dramatic decline of populations. The existence of Delacour's langur could not be confirmed in eight of the formerly known 18 areas. Only four areas where Delacour's langurs now occur are protected: Cuc Phuong National Park, Pu Luong Nature Reserve, Hoa Lu Cultural and Historical Site, and Van Long Nature Reserve. In most protected areas poaching is also common and it is to be expected that populations in unprotected areas will disappear in the foreseeable future. A reasonable estimate of the current population indicates no more than 200 individuals (Nadler 2010). Van Long Nature Reserve harbours the largest remaining population of Delacour's langurs; these animals are well protected due to close cooperation between the provincial forest protection authorities, and a local guard unit paid and trained by the Frankfurt Zoological Society. Since the establishment of the Nature Reserve in 2001, the population of Delacour's langurs has grown by about 50%, and currently numbers 100 to 120 individuals (Ebenau 2011; Nadler 2010).

Efforts to save this species are one focus of the Vietnam Primate Conservation Program of the EPRC at Cuc Phuong National Park, established in 1993 primarily to safeguard the future of this and other endangered Vietnamese primate species. The EPRC is the only facility which keeps this species. The center started a breeding program with five confiscated animals, and 20 individuals have been born since 1996. The first reintroduction of three captive bred Delacour's langur was carried out in 2011 and continued in 2012 with the release of two individuals. This was the first reintroduction of leaf-eating langurs. It followed the IUCN guidelines for nonhuman primate reintroduction. The animals were equipped with GPS-radio collars and tracked for nearly one year. The reintroduced animals should strengthen the smaller subpopulation in the larger part of Van Long Nature Reserve to support the exchange of individuals of the fragmented area of the nature reserve (Nadler 2012, Elser and Nguyen Hong Chung 2013).

References

Ebenau, A. 2011. Conservation Genetics of the Critically Endangered Delacour's Langur (*Trachypithecus delacouri*). Diploma thesis, German Primate Centre, Göttingen, Germany.

Elser, S. K. and Nguyen Hong Chung. 2013. A survey to evaluate public opinion about the reintroduction of the 'critically endangered' Delacour's langur (*Trachypithecus delacouri*) in Van Long Nature Reserve, Ninh Binh Province, Vietnam. *Vietnamese Journal of Primatology* 2: 27–35.

Le Trong Dat, Do Quang Huy, Le Thien Duc, Luu Quang Vinh and Luong Van Hao. 2008. Survey report on vertebrate fauna of Ngoc Son-Ngo Luong Nature Reserve. Report, Foundation for Social Culture of Spain and Spanish Agency for International Cooperation and Development, Madrid.

Luong Van Hao and Le Trong Dat. 2008. The status of Delacour's langur *Trachypithecus delacouri* in Cuc Phuong National Park, Ninh Binh Province, Vietnam. Report, Cuc Phuong National Park and Frankfurt Zoological Society, Hanoi.

Nadler, T. 1996. Report on the distribution and status of Delacour's langur (*Trachypithecus delacouri*). *Asian Primates* 6: 1–4.

Nadler, T. 2004. Distribution and status of the Delacour's langur (*Trachypithecus delacouri*) and recommendations for its long-term conservation. In: *Conservation of Primates in Vietnam*, T. Nadler, U. Streicher and Ha Thang Long (eds.), pp.63–71. Frankfurt Zoological Society, Hanoi.

Nadler, T. 2010. Status of Vietnamese primates – complements and revisions. In: *Conservation of Primates in Indochina*, T. Nadler, B. M. Rawson and Van Ngoc Thanh (eds.), pp.3–16. Frankfurt Zoological Society, Hanoi.

Nadler, T. 2012. Reintroduction of the 'Critically Endangered' Delacour's langur (*Trachypithecus delacouri*) – preliminary report. *Vietnamese Journal of Primatology* 1: 67–72.

Nadler, T., Le Trong Dat and Luong Van Hao. 2004. A Primate Field survey at Pu Luong Nature Reserve with the Emphasis on Delacour's langur (*Trachypithecus delacouri*). Fauna and Flora International–Vietnam Conservation Support Programme and Forest Protection Department and Frankfurt Zoological Society, Hanoi.

Nadler, T., F. Momberg, Nguyen Xuan Dang and N. Lormee (eds.). 2003. *Vietnam Primate Conservation Status Review 2002. Part 2: Leaf Monkeys.* Fauna and Flora International and Frankfurt Zoological Society, Hanoi.

Nadler, T., Le Xuan Canh, Van Ngoc Thanh and Le Khac Quyet. 2008. *Trachypithecus delacouri*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 18 March 2014.

Ratajszczak, R., R. Cox and Ha Dinh Duc. 1990. A Preliminary Survey of Primates in North Vietnam. Report, World Wide Fund for Nature, Gland, Switzerland.

Golden-headed or Cat Ba Langur

Trachypithecus poliocephalus (Trouessart, 1911) Vietnam (2000, 2002, 2004, 2006, 2008, 2010, 2012)

Richard J. Passaro, Daniela Schrudde, Roswitha Stenke, Phan Duy Thuc & Martina Raffel



The Golden-headed or Cat Ba langur, *Trachypithecus poliocephalus*, is probably the most endangered of the Asian colobines, and is assessed as Critically Endangered (Bleisch *et al.* 2008). This species occurs only on the Island of Cat Ba in the Gulf of Tonkin, northeastern Vietnam (Stenke and Chu Xuan Canh 2004). The Cat Ba Archipelago is in the world-famous Ha Long Bay, a spectacular karst formation that was invaded by the sea. The Cat Ba langur inhabits tropical moist forest on limestone karst hills, a habitat preference it shares with the other six to seven taxa of the *T. francoisi* group.

While there are no systematic and reliable data available on the historic density of the langur population on Cat Ba Island reports of indigenous people suggest the entire island of Cat Ba (140 km²) and some smaller offshore islands were previously densely populated by langurs. Hunting has been identified as the sole cause for the dramatic and rapid population decline from an estimated 2,400–2,700 in the 1960s to approximately 50 individuals by 2000 (Nadler and Long 2000). The langurs were poached mainly for trade in traditional medicines and for sport. Since the implementation

of strict protection measures in 2000, the langur population on Cat Ba Island has stabilized and appears to be on the increase (Nadler *et al.* 2003).

Although the growth of the population is encouraging, the overall status of the species remains critical. As a result of habitat fragmentation, the remaining population had been divided into several isolated subpopulations some of which consist of all-female, nonreproducing social units. The total reproductive output of this species over the years has been accordingly low. However, after many years of planning and preparation, one group consisting of two females was successfully translocated from a small off-shore islet where they had become stranded to the relative safety of the strictly protected core zone of Cat Ba National Park. Here they quickly assimilated into existing groups containing males, thus allowing them the opportunity to reproduce for the first time ever. It is hoped that with continued protection efforts and additional population management interventions such as these the species' will soon begin to rebound.

The Cat Ba Archipelago is nationally and internationally recognized for its importance to biodiversity conservation. Cat Ba National Park was established in 1986. It presently covers more than half of the main island. The archipelago (some 1,500–2,000 large and small islands, cliffs and rocks) was designated a UNESCO Man and the Biosphere Reserve in 2004.

Despite this, nature and wildlife protection on Cat Ba Island is deficient, although environmental awareness and commitment among the local communities is slowly on the increase. Efforts to effectively conserve the langurs and their habitat, however, continue to face major obstacles due to the need to better address the local community's aspirations for development, a steadily increasing human population as well as severe deficiencies in law enforcement (Stenke 2005). As elsewhere in the region, poaching is driven by increasingly attractive commercial gains in satisfying the immense local and regional demand for wildlife and animal parts for food and dubious traditional medicines. The strictest protection regime possible is necessary then for the survival of all species on Cat Ba that are, like the langurs, targeted by the Asian wildlife trade.

A conservation program for the Cat Ba langur was initiated on Cat Ba Island in November 2000 by Allwetterzoo Münster and the Zoological Society for the Conservation of Species and Populations (ZGAP), Germany. The aim is to provide for the protection of the langurs and their habitat, conduct research that will help inform future population management decisions and to help contribute to the conservation of the overall biodiversity of the Cat Ba Archipelago in collaboration with Vietnamese authorities.

References

Bleisch, B., Le Xuan Canh, H. H. Covert and Long Yongcheng. 2008. *Trachypithecus poliocephalus* ssp. *poliocephalus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist. org>. Accessed 18 March 2014.

Nadler, T. and Ha Thang Long. 2000. The Cat Ba Langur: Past, Present and Future. The Definitive Report on Trachypithecus poliocephalus – the World's Rarest Primate. Frankfurt Zoological Society, Hanoi.

Nadler, T., F. Momberg, Nguyen Xuan Dang and N.

Lormee (eds.). 2003. *Vietnam Primate Conservation Status Review 2002. Part 2: Leaf Monkeys*, pp.55–67. Fauna and Flora International and Frankfurt Zoological Society, Hanoi.

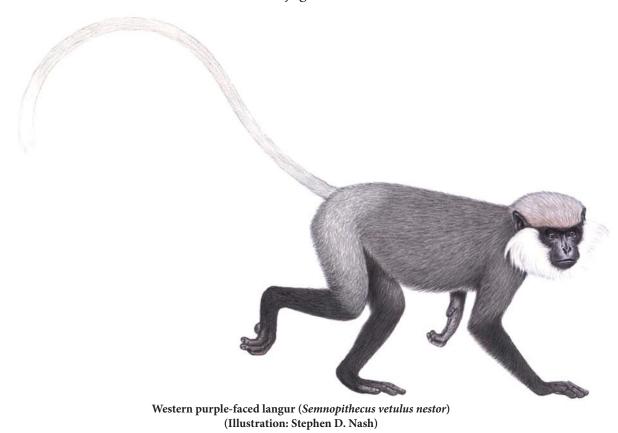
Stenke, R. 2005. Conservation of the Golden-headed Langur (*Trachypithecus poliocephalus*) on Cat Ba Island, Vietnam: Status of the Goldenheaded Langur and of Human Impact in Langur Areas. Report, People's Committee of Hai Phong (Provincial Government). Cat Ba, Hai Phong, Vietnam.

Stenke, R. and Chu Xuan Canh. 2004. The golden-headed langur (*Trachypithecus poliocephalus*) on Cat Ba Island – status, threat factors, and recovery options. In: *Conservation of Primates in Vietnam*, T. Nadler, U. Streicher and Ha Thang Long (eds.), pp.72–77. Frankfurt Zoological Society, Hanoi.

Western Purple-faced Langur

Semnopithecus vetulus nestor Bennett, 1833 Sri Lanka (2004, 2006, 2008, 2010, 2012)

Rasanayagam Rudran



Sri Lanka's Western purple-faced langur (Semnopithecus vetulus nestor) inhabits the most densely populated region around the country's capital, Colombo. As a result, haphazard urbanization severely threatens the long-term survival of this endemic monkey (Molur et al. 2003; Rudran et al. 2009; Mittermeier et al. 2012). Urbanization in its range has been so extensive that it is almost impossible to get a reliable count of its population. However, the size and distribution of its fragmented habitat in the urban landscape reflects the dire predicament this monkey is in. A 1,500-km survey conducted in 2007 through one-third of S. v. nestor's historical range (Hill 1934; Phillips 1935; Hill and Burn 1941) showed that nearly 81% of the areas surveyed consisted of deforested and human-dominated landscapes (Rudran 2007). This situation became even more alarming in 2009, when the country's 26-year civil war ended and the pent-up desire for economic progress

unleashed a flurry of development activities. A typical example is the super highway built between Sri Lanka's capital, Colombo, and the Southern city of Galle, which destroyed large areas of *S. v. nestor*'s natural habitat.

Deforestation has fragmented and drastically depleted the preferred habitat and principal food sources of the highly arboreal and folivorous *S. v. nestor*. Within the fragmented urban jungle *S. v. nestor* subsists mainly on fruits from home gardens (Dela 2007; Rudran 2007). The nutritional consequences of feeding on a low diversity diet of cultivated fruits are unclear. However, they are likely to be detrimental over the long term, because *S. v. nestor* is adapted to obtain its nutrients and energy from leaves with the help of a highly specialized stomach containing symbiotic bacteria (Bauchop and Martucci 1968).

Besides affecting *S. v. nestor*'s preferred habitat and diet, deforestation also causes other problems for this monkey's survival. For instance, fragmentation often forces this monkey to move on the ground, for which it is ill adapted, and makes young individuals vulnerable to capture as pets. While on the ground *S. v. nestor* also runs the risk of being killed by village dogs or speeding vehicles. Death by electrocution is another source of mortality when it climbs onto power lines and electricity cables (Parker *et al.* 2008). In some parts of its range *S. v. nestor* is occasionally shot and killed while feeding in home gardens (Dela 2004). Thus deforestation results in a host of human-induced fatalities, which reduce group sizes and undermine social organization.

The long-term effect of extensive deforestation resulting in local extinctions was also evident during the 2007 survey (Rudran 2007). The Western purple-faced langur was seen or recorded as present only in 43% of the sites surveyed in the eastern half of its historical range (N = 23), and 78% of the survey sites in the western half (N = 27). Moreover, the sites where it was seen or recorded as present were interspersed between areas where it was absent or rare, suggesting the occurrence of local extinctions.

Although facing a perilous future, there is hope that *S. v. nestor* can be conserved. One reason for hope is that most people living in the monkey's range follow Buddha's doctrine of compassion towards all living things. Therefore, promoting this doctrine and Buddha's own reverence of the forest could be an effective way of deterring deforestation in a country steeped in cultural traditions. Another reason for optimism stems from a recent decision taken by the Sri Lankan government to increase forest cover from 27% to 36% using native plants, to achieve the country's economic development goals (Yatawara 2011).

Even before the government announced its decision, my field staff and I had launched a research project to help reforest degraded habitats and establish safe havens for *S. v. nestor*. We were therefore delighted with the government's decision and have been continuing our efforts to help conserve *S. v. nestor*. The research on *S. v. nestor* commenced in June 2009 in the largest forest patch it now inhabits (about 21 km²). This forest became our study site due to its size and also because it surrounded two reservoirs (Kalatuwawa and Labugama) that supply water to 1.2 million inhabitants of Sri Lanka's capital. Because of its importance to people, this

forest was a secure safe haven for maintaining a viable population of *S. v. nestor* over the long term. In this safe haven, our field investigations focused on discovering the plants that were important for *S. v. nestor*'s survival, so that they could be used in a reforestation initiative to expand the size of this monkey's habitat. This research ended in December 2010 and the publication that resulted (Rudran *et al.* 2013) will be submitted to the Forest Department of Sri Lanka with a request for permission to initiate a reforestation program. The Forest Department's permission is pending at this time, but other activities that were launched along with field research are described below.

Public education was essential because the most serious threat to S. v. nestor's survival was extensive deforestation to satisfy human needs. Two programs were developed to promote conservation awareness among communities living around the study site. One focused on schoolchildren while the other was oriented towards educating an adult audience. A series of conservation-oriented lectures was presented to children attending primary, secondary and Sunday schools of different religions. Following presentations, several competitions were organized to test the knowledge that the children had gained from the lectures. These competitions encouraged children to express their environmental knowledge and concerns in the form of essays, drawings, cartoons and poetry. Over 600 schoolchildren participated in these competitions, and a panel of teachers judged their entries. This was followed by a public exhibition where the efforts of all the school children were on display, and well-known local dignitaries and conservationists awarded prizes to the winners of competitions. Details of the exhibition were then publicized via the newspapers to ensure that the project's conservation efforts reached a larger national audience. The above program was initially conducted for schools around the study site. Due to its success however, it has been continued in schools away from the study site.

Nature walks complemented the classroom lectures and brought school children closer to nature. They were conducted along forest trails and around wetlands by the project staff armed with field guides and binoculars. Every time an interesting animal was seen the project staff identified it and explained its life history, habitat preference, and the role it plays in nature. The students also learned about plants of the forest and wetlands, and the role they play in nutrient cycling, soil enrichment,

purifying water, and mitigating soil erosion and climate change. Informal discussions held after nature walks indicated that students learned more during outdoor sessions than in a classroom situation. Hence nature walks have been conducted regularly as part of the children's education program.

Discussions with the community's adults revealed that they were less willing than children to accept the need to conserve natural resources. Their indifference resulted mainly from poverty and anxieties about catering to the basic survival needs of their families. Hence, a program that dealt solely with conserving natural resources did not seem like an effective way to educate the adults. Instead, it had to be built on a platform of activities that focused on people's survival needs. In order to develop this platform, we conducted a community survey and the activities identified by it are discussed below.

A community-needs survey included interviews with residents of 250 homes around the study site. About 48% of the adults felt that their most important need was employment opportunities. This was understandable because the socioeconomic survey conducted in 2009 indicated that nearly half of the adults in this community were unemployed, while another 18% relied on meager pensions or unpredictable daily paid jobs. With over two-thirds of the community leading a hand-to-mouth existence, job opportunities was the community's most important need. Improving health services was ranked the second most important need, and vocational training was ranked third.

A meeting held to discuss employment opportunities showed that most people preferred self-employment to private sector or government employment. During the ensuing discussions the community requested a home-gardening program to help generate additional income, and also address the nutritional requirements of cash-strapped households. In return for the project's support, participants of the home gardening initiative agreed to set aside a plot within their vegetable patch to plant native species exploited by *S. v. nestor* so that these plants could be later used in the reforestation program. Thus a mechanism was developed for the community's adults to be personally involved and take pride in promoting the conservation of an endangered folivore.

The home gardening project was launched during a workshop where outside experts demonstrated techniques for creating plant beds, composting, and organic methods of pest and disease control. After the workshop people who had already established plant beds were given seeds and seedlings of vegetable plants to start their home gardens. The others were given vegetable seeds after they established plant beds. Two of our staff also received training in livestock management, mushroom farming, and bee keeping so that they could promote these income-generating activities in the community.

A healthcare programme began with assistance from HelpAge, a non-governmental organization that is devoted to elder care. HelpAge conducts eye clinics to treat cataracts and provide spectacles free of charge for the elderly. Before commencing this activity the project staff identified about 100 villagers with visual impairments. HelpAge staff then examined these villagers and diagnosed ten of them with cataracts and provided the rest with spectacles to remedy their impaired vision. However, the cataract patients had to be transported to a hospital 50 km away for surgery. The fact that the project staff made all arrangements with doctors, hospital, a transport agency and food suppliers to make the surgery possible was greatly appreciated by the local community.

In order to empower the community's women, two workshops were held to provide training in patchworkbag making. About ten trainees participated in the workshops, and some of them made and sold bags in the community for a reasonably attractive price. The project staff also contacted a well-known upscale sales outlet in Colombo to sell these bags to foreign tourists with a message explaining that the proceeds will go towards helping a community that is crucial for the conservation of *S. v. nestor*. Such messages were supposed to help expedite the sale of bags but marketing them has posed certain challenges to the community's bag makers.

The goodwill created by addressing community needs had a remarkable effect on the attitude of adults towards the project and its staff. These people finally began to realize that our project was interested in their welfare as much as it was interested in the future of this monkey, currently classified as Endangered (Dittus *et al.* 2008). Thus the people have become more receptive to the project's efforts to help promote the conservation of *S. v. nestor.* We expect this trend to continue as we strive to strengthen our bonds with the local community.

While the project strengthened its ties with local

communities another menace in the form of human-monkey conflicts began to cause serious concern. This problem has continued to intensify (Nahallage *et al.* 2008), and nearly 27% of the complaints (N = 371) received by the Department of Wildlife Conservation were about *S. v. nestor*, mainly from residents living around Colombo, the capital. Thus conserving this highly endangered endemic continues to pose challenges that must be addressed to prevent this animal from disappearing forever.

References

Bauchop, T. and R. W. Martucci. 1968. The ruminant-like digestion of the langur monkey. *Science* 161: 698–700.

Dela, J. 2004. Protecting the endemic purple-faced langur. *Loris* 23: 14–22.

Dela, J. D. S. 2007. Seasonal food use strategies of *Semnopithecus vetulus nestor* at Panadura and Piliyandala, Sri Lanka. *International Journal of Primatology* 28: 607–626.

Dittus, W., S. Molur and K. A. I. Nekaris. 2008. *Trachypithecus vetulus* ssp. *nestor*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 17 March 2014.

Hill, W. C. O. 1934. A monograph on the purple-faced leaf monkeys (*Pithecus vetulus*). Ceylon Journal of Science 29(1): 23–88.

Hill, W. C. O. and Y. Burn. 1941. Monkeys – mainly from Ceylon. *Loris II* 5: 247–252.

Mittermeier R. A., A. B. Rylands, C. Schwitzer, L. A. Taylor, F. Chiozza and E. A. Williamson (eds.). 2012. The World's 25 Most Endangered Primates 2010-2012. IUCN/Primate Specialist Group (PSG), International Primatological Society, and Conservation International, Arlington, VA.

Molur, S., D. Brandon-Jones, W. Dittus, A. A. Eudey, A. Kumar, M. Singh, M. M. Feeroz, M. Chalise, P. Priya and S. Walker (eds.). 2003. Status of South Asian Primates: Conservation Assessment and Management Plan (C.A.M.P). Workshop Report. Zoo Outreach Organization and Conservation Breeding Specialist Group (CBSG) – South Asia, Coimbatore, India.

Nahallage C. A. D., M. A. Huffman, N. Kuruppu and T. Weerasinghe. 2008. Diurnal primates in Sri Lanka and people's perceptions of them. *Primate Conservation* (23): 81–87.

Parker L., V. Nijman and K. A. I. Nekaris. 2008. When there is no forest left: fragmentation, local extinction, and small population sizes in the Sri Lankan western purple-faced langur. *Endangered Species Research* 5: 29–36

Phillips, W. W. A. 1935. *Manual of the Mammals of Ceylon*. Colombo Museum, Ceylon.

Rudran, R. 2007. A survey of Sri Lanka's endangered and endemic western purple-faced langur (*Trachypithecus vetulus nestor*). *Primate Conservation* (22): 139–144.

Rudran, R., K. Weerakoon and A. Wanasinghe. 2009. Western purple-faced langur *Trachypithecus* (*Semnopithecus*) vetulus nestor Bennett, 1833. In: The world's 25 most endangered primates 2008–2010, R. A. Mittermeier et al. (eds.), pp.24–25, 44. *Primate Conservation* (24): 1–58.

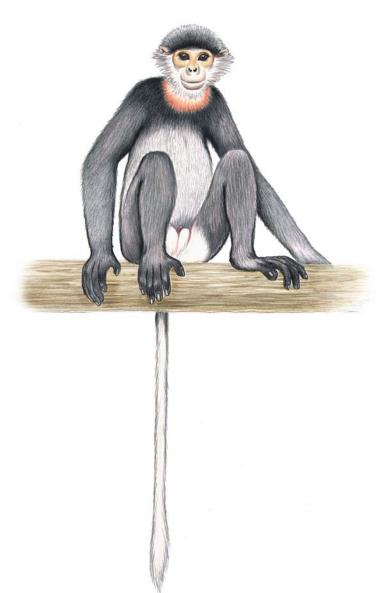
Rudran R., H. G. Salindra, K. Dayananda, D. D. Jayamanne and D. G. R. Sirimanne. 2013. Food habits and habitat use patterns of Sri Lanka's western purple-faced langur. *Primate Conservation* (27): 99–108.

Yatawara S.B. 2011. Let's preserve trees, before the last tree vanishes. *Sunday Observer* web article. Website: <www.sundayobserver.lk/2011/11/06/fea05.asp>. Accessed 27 February 2014.

Gray-shanked Douc

Pygathrix cinerea Nadler, 1997 Vietnam (2000, 2002, 2004, 2006, 2008, 2010, 2012)

Ha Thang Long & Tilo Nadler



Grey-shanked douc (*Pygathrix cinerea*) (Illustration: Stephen D. Nash)

The colobine monkeys of the genus *Pygathrix* are native to Indochina. The Gray-shanked douc was first described in 1997 as a subspecies of the Red-shanked douc (Nadler 1997), but genetic and morphological studies have since demonstrated a divergence at species level (Roos and Nadler 2001; Roos *et al.* 2007;

Stefen and Nadler 2012). It occurs in Central Vietnam between 13°30' and 16°N, and has been recorded in five provinces: Quang Nam, Quang Ngai, Kon Tum, Gia Lai and Binh Dinh (Ha Thang Long 2000, 2004; Nadler *et al.* 2003; Nguyen Thanh Tuan *et al.* 2010). Currently Gray-shanked doucs are known only from Vietnam, but photos of hunted animals from south-east Laos and genetic evidence of a Gray- and Red- shanked douc hybrid from far northeast Cambodia suggest that the species occurs also in very small areas in neighboring countries (Rawson 2010; Rawson and Roos 2008).

Surveys and research on the species in Vietnam have been conducted by the Frankfurt Zoological Society to gather information about its status, distribution and ecology. Gray-shanked douc populations are fragmented and estimated to total 600–700 individuals (Ha Thang Long, pers. obs.; Nadler 2010). Their occurrence has been confirmed in eight protected areas: Song Thanh Nature Reserve, Ngoc Linh Nature Reserve, Ba To Cultural and Historical Site, An Toan Nature Reserve, Kon Cha Rang Nature Reserve, Kon Ka Kinh National Park, Mom Ray National Park and A Yun Pa Nature Reserve. The largest known population occurs in Kon Ka Kinh National Park with 250–300 individuals (Ha Thang Long, pers. obs.).

Pygathrix cinerea is Critically Endangered (Ngoc Thanh 2008). Hunting is the principal threat to the species and is still a problem, particularly inside national parks and nature reserves. Snares are the most commonly used method of hunting, since gun confiscation programs have been carried out in a number of the areas. Often hundreds of traps are installed in trees frequently used by the douc langur groups, as well as on the ground where they are seen crossing between small forest patches. Trapped animals are often severely injured and mutilated. Forest loss in at least part of the species' range is attributable to the expansion of agriculture, illegal logging and firewood collection. Almost 10,000 ha of forest are destroyed every year in the Central Highlands.

Since 1995, the Endangered Primate Rescue Centre (EPRC) at Cuc Phuong National Park has received 68 Gray-shanked doucs confiscated from poachers and the illegal animal trade. The EPRC is the only facility which keeps this species. Despite intensive health care, approximately one-third of animals died during the days after arrival due to heavy injuries, digestion disorders, or extreme dehydration, despite intensive health care. Based on information from villagers and forest protection authorities, less than one-quarter of the hunted animals are confiscated alive. The captive breeding program at EPRC was started with the confiscated animals to provide stock for reintroduction, and 18 individuals have been born at the center. However, reintroduction can only start if hunting is eliminated.

Frankfurt Zoological Society carries out a long-term protection and monitoring project for the species in the Central Highlands of Vietnam to improve the protection of habitats and to identify possible reintroduction areas.

References

Ha Thang Long. 2000. Records of Grey-shanked Douc Langur (*Pygathrix cinerea*) in the Central Highlands of Vietnam. Frankfurt Zoological Society-Vietnam Primate Conservation Programme. Report, Frankfurt Zoological Society, Hanoi.

Ha Thang Long. 2004. Distribution and status of the grey-shanked douc (*Pygathrix cinerea*) in Vietnam. In: *Conservation of Primates in Vietnam*, T. Nadler, U. Streicher and Ha Thang Long (eds.), pp.52–57. Frankfurt Zoological Society, Hanoi.

Nadler, T. 1997. A new subspecies of Douc langur, *Pygathrix nemaeus cinereus* ssp. nov. *Zool. Garten* 67: 165–176.

Nadler, T. 2010. Status of Vietnamese primates – complements and revisions. In: *Conservation of Primates in Indochina*, T. Nadler, B. M. Rawson and Van Ngoc Thinh (eds.), pp.3–16. Frankfurt Zoological Society and Conservation International, Hanoi.

Nadler, T., F. Momberg, Nguyen Xuan Dang and N. Lormee. 2003. *Vietnam Primate Conservation Status Review 2002. Part 2: Leaf Monkeys*, pp.113–143. Fauna and Flora International and Frankfurt Zoological Society, Hanoi.

Ngoc Thanh, V., L. Lippold, T. Nadler and R. J. Timmins. 2008. *Pygathrix cinerea*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 17 March 2014.

Nguyen Thanh Tuan, Le Vu Khoi and Le Khac Quyet. 2010. New data on the distribution of grey-shanked douc langurs (*Pygathrix cinerea*) in Quang Ngai Province, Vietnam. In: *Conservation of Primates in Indochina*, T. Nadler, B. M. Rawson and Van Ngoc Thinh (eds.), pp.63-69. Frankfurt Zoological Society and Conservation International, Hanoi.

Rawson, B. M. 2010. The status of Cambodian primates. In: *Conservation of Primates in Indochina*, T. Nadler, B. M. Rawson and Van Ngoc Thinh (eds.), pp.17–25. Frankfurt Zoological Society and Conservation International, Hanoi.

Rawson, B and C. Roos. 2008. A new primate species record for Cambodia: *Pygathrix nemaeus*. *Cambodian Journal of Natural History* 1: 7–11.

Roos, C. and T. Nadler. 2001. Molecular evolution of the douc langurs. *Der Zoologische Garten* 71: 1–6.

Roos, C., Vu Ngoc Thanh, L. Walter and T. Nadler. 2007. Molecular systematics of Indochinese primates. *Vietnamese Journal of Primatology* 1: 41–53

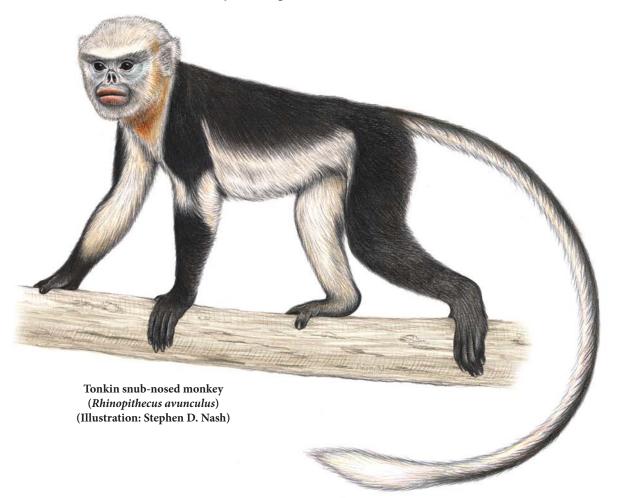
Stefen, C. and T. Nadler. 2012. Comparative cranial morphology of douc langurs (*Pygathrix cinerea*, *P. nemaeus*, *P. nigripes*). *Vietnamese Journal of Primatology* 1: 7–24.

Tonkin Snub-nosed Monkey

Rhinopithecus avunculus Dollman, 1912 Vietnam

(2000, 2002, 2004, 2006, 2008, 2010, 2012)

Le Khac Quyet, Dong Thanh Hai & Tilo Nadler



The Tonkin snub-nosed monkey is one of five unusual, large, Asian colobine monkeys of the genus Rhinopithecus, all of which have a characteristic turnedup nose. Three species are endemic to China and the newly discovered Burmese snub-nosed monkey, R. strykeri, is found in Myanmar and China. The Tonkin snub-nosed monkey is found only in northeastern Vietnam. Rhinopithecus avunculus was discovered in 1911, collected on perhaps no more than two occasions over the course of the subsequent 50 to 60 years, and consequently presumed to be extinct by a number of primatologists until it was rediscovered in 1989. Historically the species occurs only east of the Red River between about 21°09'-23°N. Due to widespread deforestation and intensive hunting in recent decades, its distribution has become severely restricted (Nadler et al. 2003).

Rhinopithecus avunculus is Critically Endangered (Le Xuan Canh et al. 2008). Recent evidence suggests there are only five known locations where Tonkin snub-nosed monkeys occur, and these are completely isolated. In 1992, a population was found in Na Hang-Chim Hoa region, Tuyen Quang Province. As a result of the discovery, Na Hang Nature Reserve was established in 1994 (Thach Mai Hoang 2011). The nature reserve comprises two separate areas: the Ban Bung and Tat Ke sectors. A study in 1993 estimated a population of between 95 and 130 individuals in each sector, respectively, which was probably an overestimation. The most recent field surveys in 2010 found and estimated only 5-10 individuals in the Tat Ke sector, and 13-16 individuals in Ban Bung sector. Hunting is still the main threat to the monkeys in the Na Hang Nature Reserve.

During surveys in 2010, local hunters, hunter shelters and gunshots were recorded commonly in both Tak Ke and Ban Bung Sectors. Conservation activities carried out by several organizations have been unsuccessful, and it has resulted in a reduction of this population (Thach Mai Hoang 2011).

A population of about 70 individuals was estimated for Cham Chu Nature Reserve in 2001, also in Tuyen Quang Province (Dong Thanh Hai *et al.* 2006). Based on interviews of local people during a survey that was reported in 1992, the population was believed to have dropped to only 20–40 individuals. A survey in 2006 provided no sightings and no reliable evidence of the survival of the population. Local reports indicate, however, a small group of 8–12 individuals still in the area. The current threats to the populations of the monkeys are hunting and habitat destruction. Conservation efforts should target reducing human activities inside the reserve.

A population of about 60 Tonkin snub-nosed monkeys was discovered in 2001 and a census in October 2013 confirmed 108–113 individuals in the Tonkin Snub-nosed Monkey Species/Habitat Conservation Area at Khau Ca, Ha Giang Province. This is the only population that is not immediately threatened. There, population and habitat monitoring, conservation education, public awareness and community participatory activities are being linked to increased protection efforts under the supervision of the University of Colorado Boulder, Fauna and Flora International (FFI), and Denver Zoo.

In 2007, a new population of about 20 Tonkin snubnosed monkeys was discovered in a small forest patch in Tung Vai Commune of Quan Ba District close to the border with China (Le Khac Quyet and Covert 2010). This is the second population of Tonkin snubnosed monkey discovered in Ha Giang Province. The newly discovered population at Tung Vai appears to be threatened by hunting and habitat loss due to timber exploitation, shifting cultivation and the collection of non-timber forest products for commercial purposes. The immediate conservation measures are likely to be training and establishing patrol groups, awareness-raising, more survey work to locate other groups and assessment of the range of the monkeys, and of the impact of cardamom production on the habitat.

The total population of the Tonkin snub-nosed monkey is believed to be less than 200 individuals.

References

Dong Thanh Hai, Do Quang Huy, Lu Quang Vinh, Nguyen Duc Manh, Nguyen Hai Ha, Ngo Duy Bach and Vu Duc Kham. 2006. A Survey of Distribution and Population Status of Tonkin Snub-nosed Monkey (*Rhinopithecus avunculus*) in Cham Chu Nature Reserve. Report, Department of Wildlife Manangement, Forest University of Vietnam, Xuan Mai, Vietnam.

Le Khac Quyet and H. H. Covert. 2010. Another population of the Tonkin snub-nosed monkey (*Rhinopithecus avunculus*) discovered in Ha Giang Province, Vietnam. *Vietnamese Journal of Primatology* 4: 19–25.

Le Xuan Canh, Le Khac Quyet, Dong Thanh Hai and R. Boonratana. 2008. *Rhinopithecus avunculus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. www.iucnredlist.org. Accessed 17 March 2014.

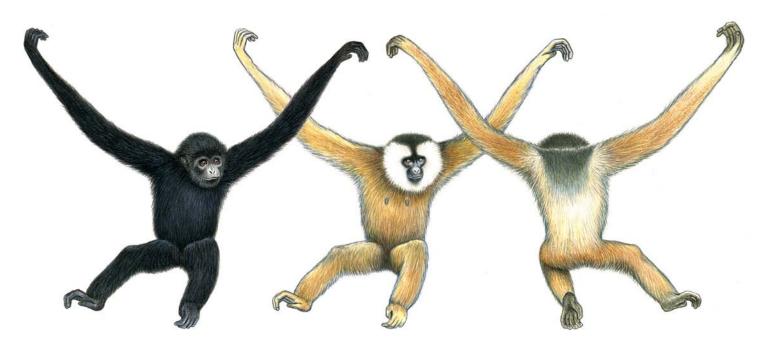
Nadler, T., F. Momberg, Nguyen Xuan Dang and N. Lormee. 2003. *Vietnam Primate Conservation Status Review 2002. Part 2: Leaf Monkeys*, pp.145-164. Fauna and Flora International and Frankfurt Zoological Society, Hanoi.

Thach Mai Hoang. 2011. Primate Survey Prioritising Tonkin Snub-Nosed Monkey (*Rhinopithecus avunculus*) and Francois' Langur (*Trachipythecus francoisi*) in Na Hang Nature Reserve, Tuyen Quang Province. Report, People Resources and Conservation Foundation, Hanoi, Vietnam.

Cao Vit or Eastern Black-crested Gibbon

Nomascus nasutus (Kunkel d'Herculais, 1884) China, Vietnam (2008, 2010, 2012)

Long Yongcheng & Tilo Nadler



Cao Vit or Eastern black-crested gibbon (*Nomascus nasutus*). Male (left) and front and back views of female (center and right) (Illustrations: Stephen D. Nash)

The Eastern black-crested gibbon or Cao Vit gibbon (*Nomascus nasutus*) was rediscovered in 2002 (La Quang Trung *et al.* 2002; Nadler 2003; La Quang Trung and Trinh Ding Hoang 2004) after being considered extinct in the 1960s (Tan 1985; Geissmann *et al.* 2003). *Nomascus nasutus* was formerly believed to comprise two subspecies (*N. n. nasutus* and *N. n. hainanus*); the first occurring in Vietnam and the second on China's Hainan Island. The subspecies have since been elevated to full species status, based on genetic differences (Roos *et al.* 2007), also supported by vocalizations and fur colouration (Geissmann *et al.* 2000; Mootnick and Fan Pengfei 2011; Van Ngoc Thinh *et al.* 2010, 2011).

Historically, *N. nasutus* was widespread east of the Red River in northern Vietnam and southern China (Geissmann *et al.* 2003; Rawson *et al.* 2011). Early surveys recorded only a few groups along the Vietnamese-Chinese border (Geissmann *et al.* 2002, 2003; Trinh Dinh Hoang 2004; Chan Bosco Pui Lok 2008; Rawson *et al.* 2011). A first simultaneous trans-boundary census of the gibbon population in 2007 recorded 18 groups

totalling approximately 110 individuals at the Chinese-Vietnam border at the Bangliang limestone forest in Jingxi County, Guangxi Province, China, and Phong Nam-Ngoc Khe Communes in the northernmost Trung Khanh District, Cao Bang Province, Vietnam (Le Trong Dat and Le Huu Oanh 2007).

Nomascus nasutus inhabits lower montane and limestone forests in a wet tropical monsoon climate at elevations of 500–900 m (Geissmann et al. 2000). The species is now known to occur in an extremely restricted area with only one surviving population in a small karst forest patch along the Chinese-Vietnam border. The area comprises only about 48 km², around 22°55′N/106°30′E, including the northern Phong Nam-Ngoc Khe forests (about 30 km²) of Trung Khanh District, Cao Bang Province, Vietnam (Rawson et al. 2011), and an immediately adjacent area (about 18 km²) in Jingxi County, Guangxi Zhuang Autonomous Region, China (Chan Bosco Pui Lok et al. 2008). A census in 2013 recorded 24 groups and 129 individuals, which represents an increase of the population by 17%

since 2007 (Nguyen The Cuong 2013). A population increase is also estimated for the groups on the Chinese side of the border (Fan Pengfei 2010).

Conservation efforts resulted in the designation of the "Cao Vit Gibbon Conservation Area" in Trung Khanh District, Vietnam, in 2007, and the Bangliang Nature Reserve, Jingxi County, China in 2009. Despite the establishment of protected areas there are still threats to this species through habitat loss and disturbance. The habitat is in danger of being cleared for cultivation, pasture for livestock, logging, firewood collection, and charcoal-production. The species is also Critically Endangered (Bleisch and Geissmann 2008) due to problems intrinsic to small populations such as inbreeding effects, genetic drift, poor mate-choice, and human or natural disasters. The latest study indicates that the gibbon population is approaching the carrying capacity of its current habitat and the carrying capacity has a significant impact on population dynamics (Fan Pengfei et al. 2013).

References

Banjie Tan. 1985. The status of primates in China. *Primate Conservation* (5): 63-81.

Bleisch, B. and Geissmann, T. 2008. *Nomascus nasutus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. www.iucnredlist.org. Accessed 17 March 2014.

Chan Bosco Pui Lok, Tan Xue-feng and Tan Wu-jing. 2008. Rediscovery of the critically endangered eastern black-crested gibbon *Nomascus nasutus* (Hylobatidae) in China, with preliminary notes on population size, ecology and conservation status. *Asian Primates Journal* 1: 17–25.

Fan Pengfei. 2010. Conservation status of *Nomascus* gibbons in China. *Primate Research*. 26 (suppl.): 89. Congress of the International Primatological Society, Kyoto. (Abstract).

Fan Pengfei, Guo-Peng Ren, Wei Wang, M. B. Scott, Chang-Yong Ma, Han-Lan Fei, Lin Wang, Wen Xiao and Jian-Guo Zhu. 2013. Habitat evaluation and population viability analysis of the last population of Cao Vit gibbon (*Nomascus nasutus*): implications for conservation. *Biological Conservation* 161: 39–47.

Geissmann T., Nguyen Xuan Dang, N. Lormée and F. Momberg 2000. *Vietnam Primate Conservation Status Review 2000. Part 1: Gibbons.* Fauna and Flora International, Hanoi.

Geissmann, T., La Quang Trung, Trinh Dinh Hoang; Dang Ngoc Can; Pham Duc Tien and Vu Dinh Thong. 2002. Report on an Overall Survey of the Cao Vit Gibbon Population (*Nomascus* sp. cf. *nasutus*) in Trung Khanh District, Cao Bang Province (second overall survey). Report, Fauna and Flora International, Hanoi.

Geissmann, T., Q. T. La, D. H. Trinh, D. T. Vu, N. C. Dang, and D. T. Pham. 2003. Rarest ape species rediscovered in Vietnam. *Asian Primates* 8: 8–10.

La Quang Trung and Trinh Ding Hoang, B. Long and T. Geissmann. 2002. Status review of black crested gibbons (*Nomascus concolor* and *Nomascus* sp. cf. *nasutus*) in Vietnam. In: Caring for Primates, Abstracts of the XIXth Congress of the International Primatological Society, pp.131–132.

La Quang Trung and Trinh Dinh Hoang. 2004. Status review of the Cao vit black-crested Gibbon (*Nomascus nasutus nasutus*) in Vietnam. In: *Conservation of Primates in Vietnam*, T. Nadler, U. Streicher and Ha Thang Long (eds.), pp.90–94. Frankfurt Zoological Society, Hanoi.

Le Trong Dat and Le Huu Oanh. 2007. The Cao Vit Gibbon *Nomascus nasutus* Population in Phong Nam and Ngoc Khe Communes, Trung Khanh District, Cao Bang Province, Vietnam. Report, Fauna and Flora International, Hanoi.

Mootnick, A. and Fan Pengfei. 2011. A comparative study of crested gibbons (*Nomascus*). *American Journal of Primatology* 73: 135–154.

Nadler, T. 2003. Rediscovery of the eastern black crested gibbon *Nomascus nasutus* in Vietnam. *The Gibbon's Voice* 6: 1–3.

Nguyen The Cuong. 2013. Survey and status review of the 'Critically Endangered' Cao Vit gibbon (Nomascus nasutus). 3rd International Conference on the Conservation of Primates in Indochina. Cuc Phuong National Park, Vietnam. Abstract.

Rawson, B.M., P. Insua-Cao, Nguyen Manh Ha, Van Ngoc Thinh, Hoang Minh Duc, S. Mahood, T. Geissmann and C. Roos. 2011. *The Conservation Status of Gibbons in Vietnam*. Fauna and Flora International and Conservation International, Hanoi.

Roos, C., Vu Ngoc Thanh, L. Walter and T. Nadler. 2007. Molecular systematics of Indochinese primates. *Vietnamese Journal of Primatology* 1: 41–53.

Trinh Dinh Hoang. 2004. Gibbon Monitoring Survey and Training in Trung Khanh, Cao Bang Province. Report, Fauna and Flora International, Hanoi.

Van Ngoc Thinh, T. Nadler, C. Roos and K. Hammerschmidt. 2010. Taxon-specific vocal characteristics of crested gibbons (*Nomascus* spp.). In: *Conservation of Primates in Indochina*, T. Nadler, B. M. Rawson and Van Ngoc Thinh (eds.), pp.121-132. Frankfurt Zoological Society and Conservation International, Hanoi.

Van Ngoc Thinh, C. Hallam, C. Roos and K. Hammerschmidt. 2011. Concordance between vocal and genetic diversity in crested gibbons. *BMC Evolutionary Biology* 11: 36.

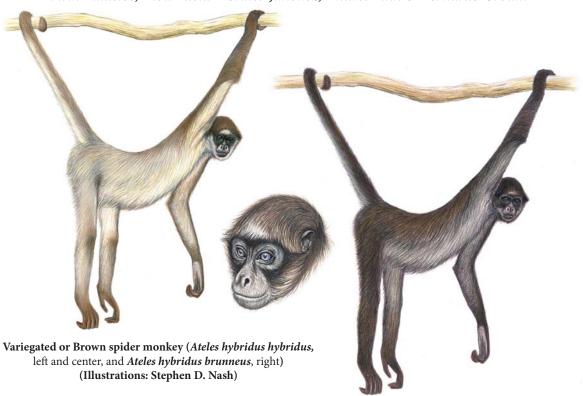
Neotropics



Variegated or Brown Spider Monkey

Ateles hybridus (I. Geoffroy Saint Hilaire, 1829) Colombia, Venezuela (2004, 2006, 2008, 2010, 2012)

Erwin Palacios, Alba Lucia Morales-Jiménez, Andrés Link & Bernardo Urbani



There are two subspecies of the variegated or brown spider monkey. *Ateles hybridus brunneus* Gray, 1870, is restricted to Colombia, occurring between the lower ríos Cauca and Magdalena in the departments of Bolívar, Antioquia and Caldas. *Ateles h. hybridus* occurs east from the right bank of the Río Magdalena extending into western Venezuela. Both subspecies are Critically Endangered because of habitat loss, hunting and the pet trade (Morales-Jiménez *et al.* 2008a, 2008b).

The large size, slow reproductive rate (single offspring at 3-4 year intervals) and generally low population densities of spider monkeys make them especially vulnerable to hunting. Historically, *A. hybridus* has suffered from habitat destruction, and only 0.67% of the current remaining *A. hybridus* distribution is protected. Most of its range has been converted to farms for agriculture and cattle (Portillo-Quintero and Velásquez 2006).

Ateles h. brunneus has a small geographic range in a region where forest loss, degradation and fragmentation

is widespread (Defler 2003). The remaining populations are surrounded by human populations, compounding the already high level of threat. Only 9% of their potential range remains as continuous forest, and legal and illegal mining as well as habitat destruction for large scale monoculture (e.g., oil palm) pose an imminent threat for the remaining populations. This subspecies has been seen recently in the lowland forests of eastern Antioquia, Caldas and Bolivar, although there are no protected areas in their current distribution. Potential refuges remain, however. The Serranía San Lucas, southern Bolívar, and some parts of Nechí have been identified as important areas for the creation of national parks. A protected area is highly necessary for this subspecies. It would include besides two other threatened endemic primates, the white-footed tamarin, Saguinus leucopus, and the woolly monkey, Lagothrix lugens.

Ateles h. hybridus is also extremely endangered due to habitat destruction in both Colombia and Venezuela. Habitat alteration appears to be the most important threat to brown spider monkeys across their current

range (Cordero-Rodríguez and Biord 2001). The forests of Magdalena river valley in Colombia, the Catatumbo area in Colombia, and the lowland forests in the state of Zulia and the piedmont of the Perijá Mountains in Venezuela are heavily destroyed because of expansionist cattle-ranching activities. In the Perijá Mountains only 30% of the forest is relatively well preserved and protected (Portillo-Quintero and Velásquez 2006). The rest is affected by rapid human expansion and land clearing, poor protection and increasing fragmentation, putting potential corridors at risk in most of its extent. Also in the Perijá Mountains, brown spider monkeys seem to be favoured game (Lizarralde 2002). In central Venezuela, some areas that had populations in 2001 were resurveyed in 2007 without any sightings; most of the areas are already covered by secondary vegetation (Cordero-Rodríguez and Biord 2001; Duque 2007). The lowland forest in the eastern part of the Andean Mountains at San Camilo, Ticoporo and Caparo, are extensively logged (Congdon 1996).

Ateles hybridus can be found in at least six zoos in Colombia, presenting problems of surplus animals and consanguinity. This species is suffering also from the pet trade; about 20 confiscated individuals are in residence in four rescue centers and need to be relocated. There is an urgent need for surveys to establish areas with populations of this species and to propose conservation measures. An *ex situ* breeding program is also necessary to maintain healthy and viable captive populations.

References

Congdon, E. R. 1996. A preliminary study of distribution, habitat use, and activity patterns of primates within Caparo Forestry Reserve, Venezuela. Report, Cleveland Metro Park Zoo and University of the Andes-Mérida/PROFAUNA/MARNR. Caracas.

Cordero-Rodríguez, G. A. and H. J. Biord F. 2001. Distribution and conservation of the spider monkey (*Ateles hybridus*) in the coastal range of northern Venezuela. *Neotropical Primates* 9: 8–11.

Defler, T. R. 2003. *Primates de Colombia*. Conservación Internacional, Serie de Guías Tropicales de Campo, Conservación Internacional Colombia, Bogotá.

Duque, D. 2007. Abundancia y distribución del mono araña *Ateles hybridus* en una región del sur-este del Parque Nacional el Ávila. Report, Provita/Programa IEA, Caracas, Venezuela.

Duque, D. 2013. Spider Monkey Conservation Project (and papers cited therein). <www.spidermonkeyproject. org>.

Hernández-Camacho, J. I. and R. W. Cooper. 1976. The nonhuman primates of Colombia. In: *Neotropical Primates Field Studies and Conservation*, R. W. Thorington Jr. and P. G. Heltne, (eds.), pp.35–69. National Academy of Sciences, Washington, DC.

Lizarralde, M. 2002. Ethnoecology of monkeys among the Barí of Venezuela: perception, use and conservation. In: *Primates Face to Face: Conservation Implications of Human and Nonhuman Primate Interconnections*, A. Fuentes and L. D. Wolfe (eds.), pp. 85–100. Cambridge University Press, Cambridge, UK.

Morales-Jiménez, A.L., A. Link and P. Stevenson. 2008a. *Ateles hybridus* ssp. *brunneus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 16 March 2014.

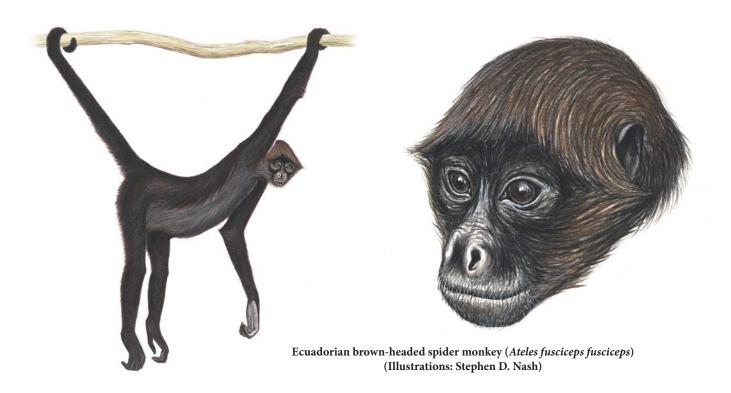
Morales-Jiménez, A.L., A. Link, P. Stevenson and B. Urbani. 2008b. *Ateles hybridus* ssp. *hybridus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 16 March 2014.

Portillo-Quintero, C. and Y. Velásquez. 2006. Disponibilidad y Estado del Hábitat de Tres Especies de Primates Amenazadas de Extinción en la Sierra de Perijá: Generación de Información Biogeográfica Base para el Diseño de Proyectos de Investigación para la Conservación (*Ateles hybridus*, *Cebus albifrons* y *Aotus trivirgatus*). Report to Provita/Programa IEA, Caracas.

Ecuadorian Brown-headed Spider Monkey

Ateles fusciceps fusciceps Gray, 1866 Ecuador, Colombia (2012)

Diego G. Tirira, Alba Lucia Morales-Jiménez & Paola Moscoso-R.



Ateles fusciceps lives in Central and South America, from southeast Panama to Ecuador, west of the Andes along the Chocó Eco-region. It is a diurnal species that inhabits mostly evergreen humid tropical and subtropical forests. It mainly inhabits large continuous forest patches in primary or secondary forest and prefers the highest levels of the canopy. The species lives in groups of up to 35 individuals. Its diet comprises mainly ripe fruits, although it supplements its diet with leaves, flowers, seeds, aerial roots, invertebrates, fungi, decaying wood, mud and termitaria. The subspecies Ateles fusciceps fusciceps inhabits the Pacific Coast of Ecuador and possibly southern Colombia, in an altitudinal range of 100 to 1,700 m above sea level.

This subspecies is classified as Critically Endangered on the IUCN Red list (Cuarón *et al.* 2008) and in the *Red Book of Mammals of Ecuador* (Peck 2011), because of its restricted distribution range and the small size of the natural populations. Extensive and ongoing deforestation and hunting are the main threats for the species in Ecuador; destruction of the humid tropical

and subtropical rainforest in western Ecuador has surpassed 80% of its original area. Tirira (2004) presented information on the historical and current distribution of the subspecies, reporting several localities where it is locally extinct, including the type locality (Hacienda Chinipamba, west of Ibarra, Intag Valley, Imbabura Province), the whole central coast of Ecuador, and the surroundings of the ríos Cayapas, San Miguel, Ónzole and Santiago, in the Esmeraldas Province. Currently, the subspecies is concentrated in the interior part of Esmeraldas Province, and adjacent regions of Imbabura and Carchi Provinces, as well as a small portion of northwest Pichincha Province. Priority areas for its conservation are the Cotacachi-Cayapas Ecological Reserve and its area of influence (mainly within a private protected forest, Los Cedros Biological Reserve), Corredor Awacachi, the Awa Ethnic Forest Reserve, north of the Mira River and close to the Colombian border, and the buffer and surrounding areas of these reserves (Moscoso et al. 2011). Population density estimates in the buffer areas of the Cotacachi-Cayapas Ecological Reserve report are 0.2-8.5 individuals/km²

(Gavilánez-Endara 2006; Cueva 2008, Estévez-Noboa 2009; Moscoso 2010). The presence of *Ateles fusciceps fusciceps* in Colombia is uncertain, but there is a record of *A. fusciceps* in Barbacoas, Nariño, that needs to be confirmed.

References

Cuarón, A. D., A. Morales, A. Shedden., E. Rodríguez-Luna and P. C. de Grammont. 2008. *Ateles fusciceps* ssp. *fusciceps*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 16 March 2014.

Cueva, X. 2008. Parámetros Demográficos de *Ateles fusciceps fusciceps* y *Alouatta palliata aequatorialis* en el Noroccidente Ecuatoriano. Bachelor's thesis, Universidad Central del Ecuador, Ouito.

Estévez-Noboa, M. 2009. Estudio Poblacional y Uso de Hábitat de *Alouatta palliata*, *Ateles fusciceps* y *Cebus capucinus* en el Bosque Protector Los Cedros, Provincia de Imbabura. Bachelor's thesis, Universidad Central del Ecuador, Quito.

Gavilánez-Endara, M. M. 2006. Demografía, Actividad y Preferencia de Hábitat de Tres Especies de Primates (*Alouatta palliata*, *Ateles fusciceps* y *Cebus capucinus*) en un Bosque Nublado del Noroccidente Ecuatoriano. Bachelor's thesis, Pontificia Universidad Católica del Ecuador, Quito.

Moscoso, R. P. 2010. Estado Poblacional del Mono Araña de Cabeza Café (*Ateles fusciceps*) en el Noroccidente del Ecuador, con Notas Ecológicas de una Relación Interespecífica con *Alouatta palliata*. Bachelor's thesis, Pontificia Universidad Católica del Ecuador, Quito.

Moscoso, P., V. A. Burbano and J. F. M. y Freile. 2011. *Guía de Observación de Primates en Áreas Naturales del Ecuador*. Ministerio de Turismo. Quito.

Peck, M. R. 2011. Brown-headed spider monkey (*Ateles fusciceps*). *In: Red Book of Mammals of Ecuador*, 2nd edition, D. G. Tirira (ed.), pp.73–75. Fundación Mamíferos y Conservación, Pontificia Universidad Catolica del Ecuador y Ministerio del Ambiente del Ecuador, Quito.

Tirira, D. G. 2004. Estado actual del mono araña de cabeza café (*Ateles fusciceps* Gray, 1866) (Primates: Atelidae) en el Ecuador. *Lyonia* 6: 17–24.

Ka'apor Capuchin

Cebus kaapori Queiroz, 1992 Brazil (2012)

Fay Clark, Fabiano R. Melo & Maurício Talebi



Ka'apor capuchin (Cebus kaapori) (Illustration: Stephen D. Nash)

The Ka'apor capuchin (Cebus kaapori), first described only recently, is found in northeast Brazil, in the state of Maranhão and the south of the state of Pará (Queiroz 1992). Its range extends from the east of the lower Rio Tocantins to the Rio Grajaú where it enters the Zona dos Cocais (Queiroz 1992; Ferrari and Queiroz 1994; Ferrari and Souza 1994; Silva and Cerqueira 1998; Carvalho et al. 1999; Cunha et al. 2007). It has been observed only in tall lowland terra firma forest, below 200 m above sea level, and has not been recorded in seasonally inundated forest or secondary forest (Rylands and Mittermeier 2013). The birth season is from June to July. This capuchin is generally seen in small groups of up to seven individuals, sometimes accompanying bearded sakis (Chiropotes satanas) (Ferrari and Lopes 1996; Carvalho et al. 1999; Rylands and Mittermeier 2013).

The precise range of *C. kaapori* is unknown, but is suspected to include an area of around 15,000 km² in the most densely populated region (Carvalho *et al.* 1999), with the highest level of deforestation and habitat degradation, in the entire Brazilian Amazon. More than 50% of the forest has been destroyed in the process of converting land to farmland and pasture (Carvalho *et al.* 1999). Deforestation continues, and

most of the remaining forests now comprise isolated, usually hunted and degraded, patches on farmland. *Cebus kaapori* occurs in only two protected areas: the Gurupí Biological Reserve and the Lago de Tucuruí Environmental Protection Area. A large part of the forest of the Gurupí Biological Reserve has been logged and destroyed since its creation in 1988. Ferrari and Lopes (1996) estimated a density of 0.98 individuals/km² in this reserve. Another survey revealed a relative abundance of 0.99 groups/10 km in the Fazenda Cauaxi in Paragominas (Carvalho *et al.*, 1999). Lopes (1993) saw three groups in 480 km walked in the Gurupí Biological Reserve.

Due to the threats of habitat loss and hunting, and a drastic population reduction (more than 80% over the past three generations (48 years), *C. kaapori* is classified as Critically Endangered on the IUCN Red List (Kierulff and Oliveira 2008). Lopes and Ferrari (1993) and Ferrari and Queiroz (1994) concluded that *C. kaapori* is one of the most threatened of all the Amazonian primates. It would seem that the Ka'apor Capuchin is naturally rare; it is hunted and is susceptible to any, even light, disturbance or degradation of its habitat. For example, selective logging of trees providing fruit, which forms a significant part of the diet, is a considerable threat

for this species (Lopes 1993). Why it is so rare may be related to competition with the sympatric Guianan brown capuchin (*Sapajus apella*) and naturally low densities may reflect the need for large home ranges. *Cebus kaapori* is not found in any zoological institutions (M. Richardson, pers. comm.). Guajá Indians, however, keep them as pets (Queiroz 1992).

More recently, researchers from the National Research and Conservation Centre for Brazilian Primates (CPB) of the Chico Mendes Institute for Biodiversity Conservation (ICMBio), Ministry of the Environment, are inventorying primates that inhabit the "arch of deforestation" in the Brazilian Amazon, including *Cebus kaapori*. Partial results show that this species has a healthy population found in the Gurupí Biological Reserve (L. Jerusalinsky, pers. comm.).

References

Carvalho Jr. O., A. C. B. de Pinto and M. Galetti. 1999. New observations on *Cebus kaapori* in eastern Brazilian Amazonia. *Neotropical Primates* 7: 41–43.

Cunha, F. A., M. A. Lopes, S. de M., Dantas, N. A. S. do Carmo and S. do S. B. da Silva. 2007. Registro de ocorrência de *Cebus kaapori* (Cebidae: Primates) na APA Lago de Tucuruí. *Neotropical Primates* 14: 84–85.

Ferrari, S. F. and M. A. Lopes. 1996. Primate populations in eastern Amazonia. In: *Adaptive Radiations of Neotropical Primates*, M. A. Norconk, A. L. Rosenberger and P. A. Garber (eds.), pp.53–67. Plenum Press, New York.

Ferrari, S. F. and H. L. Queiroz. 1994. Two new Brazilian primates discovered, endangered. *Oryx* 28: 31–36.

Ferrari, S. F. and A. P. de Souza Jr. 1994. More untufted capuchins in southeastern Amazonia? *Neotropical Primates* 21: 9–10.

Kierulff, M. C. M. and M. M. de Oliveira. 2008. *Cebus kaapori*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. www.iucnredlist.org. Accessed 16 March 2014.

Lopes, M. A. 1993. Conservação do Cuxiú-preto, *Chiropotes satanas satanas* (Cebidae: Primates) e de Outros Mamíferos na Amazônia Oriental. Master's thesis, Universidade Federal do Pará, Belém.

Lopes, M. A. and S. F. Ferrari. 1993. Primate conservation in eastern Brazilian Amazonia. *Neotropical Primates* 1: 8–9.

Queiroz, H. L. 1992. A new species of capuchin monkey, genus *Cebus* Erxleben 1977 (Cebidae, Primates), from eastern Brazilian Amazonia. *Goeldiana Zoologia* 15: 1–3.

Rylands, A. B. and R. A. Mittermeier. 2013. Ka'apor capuchin *Cebus kaapori*. In: *Handbook of the Mammals of the World. Volume 3. Primates*, R. A. Mittermeier, A. B. Rylands and D. E. Wilson (eds.), p.410. Lynx Edicions, Barcelona.

Silva Jr., J. S. and R. Cerqueira. 1998. New data and a historical sketch on the geographical distribution of the Ka'apor capuchin, *Cebus kaapori* Queiroz, 1992. *Neotropical Primates* 6: 118–121.

San Martín Titi Monkey

Callicebus oenanthe Thomas, 1924 Peru (2012)

Jan Vermeer



San Martín titi monkey (Callicebus oenanthe) (Illustrations: Stephen D. Nash)

The San Martín titi monkey was discovered in 1924, but until 2007 was only known from six museum specimens and scarce observations, all from the Alto Mayo Valley in northeastern Peru (Thomas 1924, 1927; Hershkovitz 1990; Mark 2003; Rowe and Martinez 2003; De Luycker 2006). Extensive surveys by the team of Proyecto Mono Tocón have shown that the distribution of the species extends from the Alto Mayo Valley in the south, restricted largely (but not completely) by mountains ranges in the west, south and north, and the Río Huallaga in the east (Boveda-Penalba *et al.* 2009). It inhabits the lowland forest on the eastern foothills of the Andes, rarely occurring at altitudes above 1,000 m above sea level.

Callicebus oenanthe is endemic to the department of San Martín, which has the highest deforestation rates in Peru. Although its original range was estimated to have been approximately 14,000 km², its habitat has been reduced to less than 6,500 km², of which only 1,900 km² is thought to be covered with good habitat (Shanee *et al.* 2013). Considering that the forest cover data used for this study were from 2007/2008 and the high deforestation rate in the lowlands, it is very likely that the situation is even worse today.

The San Martín titi monkey is highly variable in colouration (Boveda-Penalba *et al.* 2009, Vermeer *et al.* 2011). Most animals in the north are brownish with a white mask, while in the south many lack the typical mask and have a darker or more orange color (Proyecto Mono Tocón, unpubl. data).

Only small and isolated populations that are probably not viable have been encountered during extensive surveys in its range (Boveda-Penalba *et al.* 2009). Connecting isolated forest patches is mostly impossible due to human presence. The situation is even more complicated as the San Martín titi monkey seems to have a preference for the edges between primary and secondary forest, where human pressure is often very high (Proyecto Mono Tocón, unpubl. data). The species

can be found on the borders of some protected areas. Although a number of (relatively) small conservation concessions and private conservation areas have been created in the range of the San Martín titi monkey, only two may harbour viable populations. Unfortunately, most of its habitat is still unprotected, and is in danger of being destroyed for agriculture and logging.

The San Martín titi monkey is Critically Endangered (Veiga *et al.* 2011) as it is estimated that a population reduction of \geq 80% has occurred over the last 25 years. The isolation of unviable populations in small forest patches increases the risk for the species. More support from national and regional governments and (international) conservation organizations is urgently needed to save this species from extinction.

References

Boveda-Penalba, A. J., J. Vermeer, F. Rodrigo and F. Guerra-Vasquez, F. 2009. Preliminary report on the distribution of *Callicebus oenanthe* on the eastern feet of the Andes. *International Journal of Primatology* 30: 467–480.

De Luycker, A. M. 2006. Preliminary report and conservation status of the Río Mayo Titi Monkey, *Callicebus oenanthe* (Thomas, 1924) in the Alto Mayo Valley, northeastern Peru. *Primate Conservation* (21): 33–39.

Hershkovitz, P. 1990. Titis, New World monkeys of the genus *Callicebus*: a preliminary taxonomic review. *Fieldiana Zoology, new series* 55: 1–109.

Mark, M. 2003. Some observations on *Callicebus* oenanthe in the upper Rio Mayo Valley, Peru. *Neotropical Primates* 11: 183–187.

Rowe, N. and W. Martinez. 2003. *Callicebus* sightings in Bolivia, Peru and Ecuador. *Neotropical Primates* 11: 32–35.

Shanee, S., J. C. Tello-Alvarado, J. Vermeer and A. J. Bóveda-Penalba. 2013. GIS Risk Assessment and GAP analysis for the Andean titi monkey (*Callicebus oenanthe*). *Primate Conservation* (26): 17–23.

Thomas, O. 1924. New *Callicebus*, *Conepatus*, and *Oecomys* from Peru. *Annals and Magazine of Natural History*, *Series* 9, 14: 286–288.

Thomas, O. 1927. The Godman-Thomas Expedition to Peru - V. On mammals collected by Mr. R. W. Hendee in the province of San Martín, N. Peru, mostly at Yurac Yacu. *Annals and Magazine of Natural History, Series 9*, 19: 361–363.

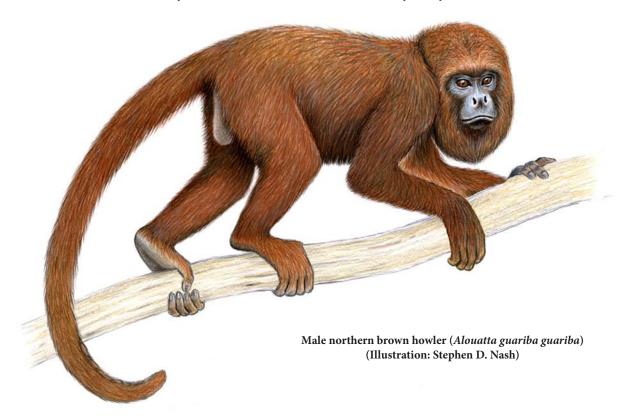
Veiga, L., A. Bóveda-Penalba, J. Vermeer, J. C. Tello-Alvarado and F. Cornejo. 2011. *Callicebus oenanthe*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. www.iucnredlist.org. Accessed 01 February 2014.

Vermeer, J., J. C. Tello-Alvarado, S. Moreno-Moreno and F. Guerra-Vásquez. 2011. Extension of the geographical range of white-browed titi monkeys (*Callicebus discolor*) and evidence for sympatry with San Martín titi monkeys (*Callicebus oenanthe*). *International Journal of Primatology* 32: 924–930.

Northern Brown Howler Monkey

Alouatta guariba guariba (Humboldt, 1812) Brazil (2012)

Leonardo Gomes Neves, Fay E. Clark, Fabiano R. Melo, Anthony B. Rylands & Maurício Talebi



The brown howler is separated into two subspecies, the northern brown howler, Alouatta guariba guariba, and southern brown howler, A. g. clamitans (Rylands et al. 2000; Groves 2001, 2005). Following a study of the morphology of the cranium and hyoid apparatus of the two forms, Gregorin (2006) considered them to be full species, using the name A. fusca (É. Geoffroy Saint-Hilaire, 1812) rather than A. guariba (Humboldt, 1812) for the northern form, following the recommendation of Hershkovitz (1963). Rylands and Brandon-Jones (1998; p.895) argued that the correct name is in fact guariba. Kinzey (1982) concluded that A. g. guariba occurred north of the Rio Doce; clamitans to the south. Rylands et al. (1988) observed what they believed to be A. g. clamitans further north, in the middle Jequitinhonha valley, and indicated that the Rio Jequitinhonha basin, not the Rio Doce, divided the two howlers. The extreme rarity of brown howlers north of the Jequitinhonha has confounded attempts to clarify the taxonomy. Only recently have few and minuscule populations been located in southern Bahia. Gregorin (2006) argued that the original range of the northern brown howler in fact

extended from Bahia (Rio Paraguaçú) south along the coastal forest to the state of Rio de Janeiro (crossing as such the lower and middle Rio Doce), and that *clamitans*, the southern form, occurs inland north as far as the upper and middle Jequitinhonha. This would be compatible with the findings of Rylands *et al.* (1988) in the Jequitinhonha valley and, in this case, some of the populations surveyed by Chiarello (1999) may have been of the northern subspecies *A. g. guariba*. Here, we maintain the names and subspecific classification as used by Rylands *et al.* (2000), Groves (2001, 2005), and Glander (2013).

Both sexes of *A. g. guariba* are a red-fawn colour, the females being rather duller in colour. *Alouatta g. guariba* inhabits lowland, submontane and montane Brazilian Atlantic forest. It is a folivore-frugivore, including more fruit in its diet according to seasonal availability (Neville *et al.* 1988; Mendes 1989; Chiarello 1994; Glander 2013; Rylands and Mittermeier 2013). As such, brown howler monkeys are important seed dispersers for numerous plant species (Chiarello and Galetti 1994). While the

parent species *Alouatta guariba* is widely distributed and is classified as Least Concern on the IUCN Red List, *A. g. guariba* has a considerably more restricted range and is Critically Endangered (Mendes *et al.* 2008). The primary threats are widespread forest loss and fragmentation throughout its range, due to logging and agriculture (Horwich 1998), hunting (Melo 2005; Canale *et al.* 2012), and disease epidemics such as yellow fever brought from Africa (Holzmann *et al.* 2010).

An action plan for 27 threatened mammals of the Brazilian Atlantic Forest, including A. g. guariba, was drawn up in 2010 by the National Research and Conservation Centre for Brazilian Primates (CPB) of the Chico Mendes Institute for Biodiversity Conservation (ICMBio) (Brazil, MMA, ICMBio-CPB 2010). A conservation project for A. g. guariba is now ongoing as an immediate effect of this federal conservation public policy. Surveys carried out since 2012, by the Instituto de Estudos Sócioambientais do Sul da Bahia (IESB) and the State University of Santa Cruz (UESC) with the support of Conservation International and the Rainforest Trust, have attempted to locate and count surviving populations, understand better the threats to their survival, and establish the limits to its geographic distribution. To date, eight populations in small and widely separated forest patches have been found: 1) Itajú de Colônia - two groups and one individual seen; 2) Itarantim – two groups heard; 3) Caatiba – three groups totalling nine individuals; 4) Itapetinga – two groups heard; 5) Macarani - one group, one individual seen; 6) Ribeirão Largo - one group heard; 7) Pouso Alegre - one group, two individuals seen; and 8) Itambé - two groups heard (L. G. Neves, unpubl. data). The surveys indicate that most of the surviving populations are those in the valleys of the Rio Pardo and Rio Jequitinhonha. Further north, in the cacao-growing region of southern Bahia, they have been largely hunted out.

There are a number of protected areas in the northern brown howlers range in Bahia and northeastern Minas Gerais, all created since 1980. They have been reported in the Mata Escura Biological Reserve (51.046 ha, created in 2003), just north of the middle Rio Jequitinhonha (Melo 2005). Adding the locations in the lower reaches of Jequitinhonha basin reported by Rylands *et al.* (1988), the known population today is unlikely to number more than 250 mature individuals, and no subpopulation is believed to exceed 50 mature individuals. Howlers have not been seen further north in the Una Biological Reserve (18,500 ha, created in

1980) for more than 60 years. It is not known if they still occur in the submontane and montane forest of the Serra das Lontras National Park (11,336 ha, created in 2010). Future surveys will target protected areas and the limits of their supposed range—the Rio Paraguaçú in the north to the Río Doce in the south, and protected areas in southern Bahia.

Hunting has resulted in the surviving populations being very small and isolated and a metapopulation management plan for the future will need to incorporate translocation of threatened populations. A promising initiative underway at the Serra Bonita Private Reserve, Camacan, Bahia, owned by Vitor Becker, and managed by the NGO Instituto Uiraçú, is the successful release, with the collaboration of ICMBio, of two confiscated pets—an incipient reintroduction of the species that has not been seen or heard there for more than 50 years.

References

Brazil, MMA, ICMBio-CPB. 2010. Sumário Executivo do Plano de Ação Nacional para a Conservação dos Mamíferos da Mata Atlântica Central. Ministério do Meio Ambiente (MMA), Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio), Centro de Proteção de Primatas Brasileiros (CPB), Brasília.

Canale, G. R., C. A. Peres, C. E. Guidorizzi, C. A. F. Gatto and M. C. M. Kierulff. 2012. Pervasive defaunation of forest remnants in a tropical biodiversity hotspot. *PLoS ONE* 7: e41671. doi:10.1371/journal.pone.0041671

Chiarello, A. G. 1994. Diet of the brown howler monkey *Alouatta fusca* in a semi-deciduous forest fragment of southeastern Brazil. *Primates* 35: 25–34.

Chiarello, A. G. 1999. Effects of fragmentation of the Atlantic forest on mammal communities in south-eastern Brazil. *Biological Conservation* 89: 71–82.

Chiarello, A. G. and M. Galetti. 1994. Conservation of the brown howler monkey in south-east Brazil. *Oryx* 28: 37–42.

Glander, K. E. 2013. Brown howler *Alouatta guariba*. In: *Handbook of the Mammals of the World. Volume 3*. *Primates*, R. A. Mittermeier, A. B. Rylands and D. E. Wilson (eds.), p.531. Lynx Edicions, Barcelona.

Gregorin, R. 2006. Taxonomy and geographic variation

of species of the genus *Alouatta* Lacépède (Primates, Atelidae) in Brazil. *Revista Brasileira de Zoologia* 23: 64–144.

Groves, C. P. 2001. *Primate Taxonomy*. Smithsonian Institution Press, Washington, DC.

Groves, C. P. 2005. Order Primates. In: *Mammal Species of the World*, D. E. Wilson and D. M. Reeder (eds), pp.111–184. The Johns Hopkins University Press, Baltimore, MD.

Hershkovitz, P. 1963. *Primates: Comparative Anatomy and Taxonomy, V, Cebidae, Part B.* A Monograph by W. C. Osman Hill, Edinburgh University Press, 1962, xxix 537pp., 34pls., 94 figs., 3 maps. A critical review with a summary of the volumes on New World primates. *American Journal of Physical Anthropology* 21: 391–398.

Holzmann, I., I. Agostini, J. I. Areta, H. Ferreyra, P. Beldomenico and M. S. Di Bitetti. 2010. Impact of yellow fever outbreaks on two howler monkey species (*Alouatta guariba clamitans* and *A. caraya*) in Misiones, Argentina. *American Journal of Primatology* 72: 475–480.

Horwich, R. H. 1998. Effective solutions for howler conservation. *International Journal of Primatology* 19: 579–598.

Kinzey, W. G. 1982. Distribution of primates and forest refuges. In: *Biological Diversification in the Tropics*, G. T. Prance (ed.), pp.455–482. Columbia University Press, New York.

Melo, F. R. 2005. A Reserva Biológica Federal da Mata Escura e sua importância como unidade de conservação para os primatas do médio rio Jequitinhonha, Minas Gerais. *Neotropical Primates* 13: 26–29.

Mendes, S. L. 1989. Estudo ecológico de *Alouatta fusca* (Primates: Cebidae) na Estação Biológica de Caratinga, MG. *Revista Nordestina de Biologia* 6: 71–104.

Mendes, S. L., A. B. Rylands, M. C. M. Kierulff and M. M. de Oliveira. 2008. *Alouatta guariba*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.2. <www.iucnredlist.org>. Accessed 16 March 2014.

Neville, M. K., Glander, K., Braza, F. and Rylands, A. B. 1988. The howling monkeys, genus *Alouatta*. In: *Ecology and Behavior of Neotropical Primates*, Vol. 2, R. A. Mittermeier, A. B. Rylands, A. F. Coimbra-Filho and G. A. B. da Fonseca (eds.), pp.349–453. World Wildlife Fund, Washington, DC.

Rylands, A. B. and D. Brandon-Jones. 1998. The scientific nomenclature of the red howlers from the northeastern Amazon in Brazil, Venezuela and the Guianas. *International Journal of Primatology* 19: 879–905.

Rylands, A. B. and R. A. Mittermeier, 2013. Family Atelidae (howlers, spider and woolly monkeys and muriquis). In: *Handbook of the Mammals of the World. Volume 3. Primates*, R. A. Mittermeier, A. B. Rylands and D. E. Wilson (eds.), pp.484–523. Lynx Edicions, Barcelona.

Rylands, A. B., W. R. Spironelo, V. L. Tornisielo, R. M. Lemos de Sá, M. C. M. Kierulff and I. B. Santos. 1988. Primates of the Rio Jequitinhonha valley, Minas Gerais, Brazil. *Primate Conservation* (9): 100–109.

Rylands, A. B., H. Schneider, A. Langguth, R. A. Mittermeier, C. P. Groves and E. Rodríguez-Luna. 2000. An assessment of the diversity of New World primates. *Neotropical Primates* 8: 61–93.

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There are **Regional Vice Chairs** for the principal areas where primates occur, as follows: **AFRICA SECTION** – *W. Scott McGraw*, The Ohio State University, Columbus, OH, USA, *David N. M. Mbora*, Whittier College, Whittier, California, USA, and *Janette Wallis*, Interdisciplinary Perspectives on the Environment, University of Oklahoma, Oklahoma, OK, USA; **MADAGASCAR SECTION** – *Christoph Schwitzer*, Bristol Zoological Society, Bristol Zoo Gardens, Bristol, UK, and *Jonah Ratsimbazafy*, Durrell Wildlife Conservation Trust – Madagascar Programme, Antananarivo, Madagascar; **NEOTROPICAL SECTION** – **Mesoamerica** – *Liliana Cortés-Ortiz*, University of Michigan, Ann Arbor, MI, USA; **Andean Countries** – *Erwin Palacios*, Conservación Internacional Colombia, Bogotá, Colombia, and *Eckhard W. Heymann*, Deutsches Primatenzentrum, Göttingen, Germany; **Brazil and the Guianas** – *M. Cecília M. Kierulff*, Instituto Pri-Matas para a Conservação da Biodiversidade, Belo Horizonte, Minas Gerais, Brazil, *Fabiano Rodrigues de Melo*, Universidade Federal de Goiás, Jataí, Goiás, Brazil, and *Maurício Talebi*, Universidade Federal de São Paulo, Diadema, São Paulo, Brazil; **ASIA SECTION** – **China** – *Long Yongcheng*, The Nature Conservancy, China; **Southeast Asia / Indochina** – *Jatna Supriatna*, Conservation International Indonesia Program, Jakarta, Indonesia, *Christian Roos*, Deutsches Primatenzentrum, Göttingen, Germany, *Benjamin M. Rawson*, Fauna and Flora International, Hanoi, Vietnam, and *Ramesh Boonratana*, Mahidol University International College, Salaya, Nakhon Pathom, Thailand; **South Asia** – *Sally Walker*, Zoo Outreach Organization, Coimbatore, India, and *Sanjay Molur*, Wildlife Information Liaison Development, Coimbatore, Tamil Nadu, India.

INTERNATIONAL PRIMATOLOGICAL SOCIETY (IPS)

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There are six Vice-Presidents: Treasurer and Vice-President for Membership – Steven Schapiro, Department of Veterinary Sciences, UTMDACC, Bastrop, Texas, USA; Vice-President for Communications – Claudia Fichtel, Behavioral Ecology & Sociobiology Unit, German Primate Center (DPZ), Kellnerweg 4, D - 37077 Göttingen, Germany; Vice-President for Conservation – Janette Wallis, Interdisciplinary Perspectives on the Environment (IPE), The University of Oklahoma, 630 Parrington Oval, Monnet Hall, Rm 555, Norman, OK 73019-4036, USA; Vice-President for Captive Care – Christoph Schwitzer, Bristol Zoological Society, c/o Bristol Zoo Gardens, Clifton, Bristol, BS8 3HA, UK; Vice-President for Education and Outreach – Elizabeth Lonsdorf, Department of Psychology, Biological Foundations of Behavior Program, Franklin and Marshall College, P.O. Box 3003, Lancaster, PA 17604, USA; and Vice President for Research – Joanna Setchell, Durham University, Department of Anthropology, Dawson Building, South Road, Durham, DH1 3LE, UK.

The **Species Survival Commission (SSC)** is one of six volunteer commissions of IUCN, a union of sovereign states, government agencies and non-governmental organizations. SSC's mission is to conserve biological diversity by developing and executing programs to save, restore and wisely manage species and their habitats. Survival of the world's living primate species and subspecies is the principal mission of the IUCN/SSC Primate Specialist Group (PSG), over 400 volunteer professionals who represent the front line in international primate conservation. The PSG website is www.primate-sg.org.

The **International Primatological Society (IPS)** was created to encourage all areas of non-human primatological scientific research, to facilitate cooperation among scientists of all nationalities engaged in primate research, and to promote the conservation of all primate species. The Society is organized exclusively for scientific, educational and charitable purposes. For more information about IPS, visit www.internationalprimatologicalsociety.org.

Conservation International (CI). Building upon a strong foundation of science, partnership and field demonstration, CI empowers societies to responsibly and sustainably care for nature, our global biodiversity, for the well-being of humanity. With headquarters in Arlington, VA, CI works in more than 40 countries on four continents. For more information about CI, visit www.conservation.org.

Bristol Zoological Society (BZS) runs Bristol Zoo Gardens and the Wild Place Project. BZS undertakes conservation action and conservation research in both the UK and the developing world. Its mission is to save wildlife through conservation action and engaging people with the natural world. For more information about BZS, visit www.bristolzoo.org.uk.

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